

Wound Healing Potential of Some Medicinal Plants

*Girija Pashikanti¹, Chowdary Saithi²

Head Department of Pharmacology. Vaagdevi College of Pharmacy, Hanamkonda, Warangal-506001, Telangana, India.
 Department of Pharmacology, Vaagdevi College of Pharmacy, Hanamkonda, Warangal, 506001, Telangana, India.
 *Corresponding author's E-mail: lckgr3@gmail.com

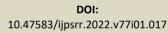
Received: 23-08-2022; Revised: 20-10-2022; Accepted: 26-10-2022; Published on: 15-11-2022.

ABSTRACT

Wound healing is the process by which skin repairs itself. Wound healing can be divided into 4 phases: haemostasis, inflammation, proliferation, and remodelling. In humans, keratinocytes re-form a functional epidermis (re-epithelialization) as rapidly as possible, closing the wound and reestablishing tissue homeostasis. Dermal fibroblasts migrate into the wound bed and proliferate, creating granulation tissue rich in extracellular matrix proteins and supporting the growth of new blood vessels. Ultimately, this is remodelled over an extended period, returning the injured tissue to a state similar to that before injury. Dysregulation in any phase of the wound healing cascade delays healing and may result in various skin pathologies, including non-healing, hypertrophic scarring and chronic ulceration. Various plant products have been used in the treatment of wounds over the years. Recognizing the important role of traditional plants, we have undertaken an extensivesurvey of literature reporting the use of medicinal plants for wounds. We describe the activeingredients, bioactivities, clinical uses of 8 medicinal plant species. Several species including *curcuma longa*, honey, *Terminalia chebula, Aloe vera, Centella asiatica, Arctium lappa, Commiphora myrrha*, showing wound healing activities by their anti-inflammatory and antioxidant mechanisms.

Keywords: Transforming growth factor beta 1 ECM, Extracellular matrix, Interleukin-1, Matrix metallo proteases, Tumour necrosis factor-α, Natural factor kappa B, inducible nitric oxide synthase.

QUICK RESPONSE CODE →





DOI link: http://dx.doi.org/10.47583/ijpsrr.2022.v77i01.017

INTRODUCTION

ounds are disruptions to the continuity of cells due to a physical, chemical, thermal, infectious or immunological injury to the skin. Effective wound healing is defined by the restoration of functional tissue integrity. Proper wound healing is achieved by adequate activation and infiltration of inflammatory cells, neutrophils and macrophages, which produce proinflammatory cytokines such as tumor necrosis factor α (TNF- α) and interleukin-1 (IL-1)¹. These inflammatory cytokines result in the activation of growth factors such as transforming growth factor (TGF)- β , and several fibroblast growth factors, resulting in the proliferation and infiltration of activated fibroblasts to the wound site¹. However, these natural healing process is impaired with aging, obesity, and endocrine abnormalities such as diabetes mellitus².

wound healing is the process by which skin repairs itself following injury caused by surgery trauma and burns. The healing process is classically divided into 4 phases: coagulation (a.k.a. haemostasis), inflammation, proliferation (a.k.a. granulation), and remodelling (a.k.a. maturation)³. Upon injury, a fibrin clot rapidly forms to restore haemostasis^{4,5}. Platelets present in the blood trigger the clotting cascade and secrete several growth factors, initiating wound healing⁶. In the following inflammation phase, neutrophils migrate into the wound site engulfing foreign debris and killing bacteria by phagocytosis and releasing proteolytic enzymes^{6,7}. Coincidently, blood monocytes infiltrate the injury site and differentiate into macrophages, releasing proteases to debride the wound⁶, and secrete a mixture of bioactive molecules, including transforming growth factor beta 1 (TGF- β 1), that stimulates the migration of fibroblasts and epithelial cells⁸. The proliferation phase usually starts about 3 days after wounding; it involves diverse activities including angiogenesis (by endothelial cells), granulation tissue formation (by fibroblasts), and re-epithelialization (by keratinocytes)^{9,10}. In this stage, fibroblasts produce a large amount of extracellular matrix (ECM), mainly collagen, to form the granulation tissue which replaces the damaged tissue. Meanwhile, the keratinocytes migrate, proliferate, differentiate, and re-form a functional epidermis (re-epithelialization), closing the lesion and protecting underlying tissues from further trauma¹¹. As the wound matures, the characteristic disorganized ECM of granulation tissue is actively remodeled by the dermal fibroblast cell population¹², whose numbers are progressively reduced through apoptosis¹³. The outcome of wound healing is scar tissue (aka fibrosis) with sparsely distributed fibroblasts within a collagen-rich ECM.



Available online at www.globalresearchonline.net

©Copyright protected. Unauthorised republication, reproduction, distribution, dissemination and copying of this document in whole or in part is strictly prohibited.

Compared to the original tissue, scar tissue, having distinct texture and reduced biomechanical and functional properties, is characteristically altered¹⁴.

CLASSIFICATION OF WOUNDS

Wounds are generally classified according to the underlying cause of the development of wounds.

Acute wounds

In acute wounds, there is tissue damage/injury that generally occurs through an orderly and time-reparative phase that results in the anatomical and functional integrity being restored sustainably. Acute wounds are typically caused by the cuts or surgical incisions¹⁵.

Closed wounds

The blood escapes from the circulatory system in closed wounds but stays inside the body. It becomes evident in the form of bruises.

Open wounds

Blood leaks from the body through an open wound and bleeding is clearly noticeable. The open wound may be further divided into categories according to the source causing the wound.

Incised wounds

This is a wound with no loss of tissue and minor damage to tissue. It is caused primarily by sharp objects like a scalpel or knife.

Tear or laceration wounds

This is the non-chirurgical injury in conjunction with other types of trauma which results in tissue loss and damage.

Puncture wounds

These are caused by an object like a nail or a needle, which punctures the skin. Since dirt may penetrate deep into the wound, chances of infection are common on them.

Abrasive or superficial wounds

Sliding slip onto a rough surface induces abrasion. During this time, abrasion is scrapped off the top layer of the skin, i.e., epidermis which exposes nerve endings resulting in a painful injury.

Penetration wounds

Penetration wounds are chiefly caused by an object like a knife going in and out of the skin.

Gunshot wounds

They are typically produced by bullet or similar projectile which drives through or into the body.

Chronic wounds

Chronic wounds are the wounds that have not gone through the usual healing stages and hence reach a state of pathologic inflammation. They need extended healing time¹⁶.

FACTORS AFFECTING WOUND HEALING

Oxygenation

Oxygen is essential for the metabolism of cells, particularly the production of energy through ATP, and is necessary for almost all wound healing processes. It protects wounds from infection, causes angiogenesis, increases differentiation of keratinocytes, migration and reepithelialization, improves proliferation of fibroblasts and synthesis of collagen, and facilitates contraction of wounds. The microenvironment of the early wound is deprived of oxygen and is very hypoxic owing to ingestion by metabolically active cells. several systemic disorders will produce reduced vascular flow, including advancing age and diabetes, thereby setting the stage for inadequate oxygenation of the tissue. This superposition of inadequate perfusion produces a hypoxic wound in the sense of recovery. Chronic wounds are hypoxic in particular: tissue oxygen concentrations were measured transcutaneous in chronic wounds of 5 to 20 mm Hg, relative to control tissue concentrations of 30 to 50 mm Hg.

Infections

Micro-organisms that are typically sequestered on the skin surface gain access to the underlying tissues until the skin is wounded. If the wound is listed as having inflammation, colonization, local invasion/critical colonization, and/or spreading invasive infection determines the state of infection and replication status of the micro-organisms. Contamination is the presence of non-replicating microbes on a wound, while colonization is characterized as the presence without tissue damage of replicating microorganisms on the wound. An intermediate stage is local infection/critical colonization, with proliferation of microorganisms and the beginning of local tissue responses. The involvement of replicating organisms inside a wound with subsequent damage to the host is known as invasive infection. Inflammation is a natural part of the wound healing process and is necessary for the elimination of micro-organisms that are infected. However, inflammation can be prolonged in the absence of successful because microbial clearance decontamination, is incomplete. The sustained elevation of pro-inflammatory cytokines such as interleukin-1 (IL- 1) and TNF- α will contribute to both bacteria and endotoxins and elongate the inflammatoryprocess. The wound can reach a chronic state and refuse to heal if this persists. In addition, this prolonged inflammation contributes to an elevated level of matrix metallo-proteases (MMPS), a protease family that can degrade the ECM. A decreased level of the naturally occurring protease inhibitors occurs in combination with the increased protease content. This change in protease equilibrium may cause the rapid deterioration of growth factors that occur in chronic wounds.

Age

The elderly population (people over 60 years of age) is growing more than any other age group (a significant risk factor for delayed wound healing). Several cellular and



molecular- level clinical and animal studies have explored age-related changes and delays in wound healing. It is widely accepted that the impact of aging induces a transient pause in wound healing in stable older people, but not a genuine disability in terms of the consistency of healing.

Stress

Stress has a considerable influence on human well-being and social behaviour. Stress is associated with multiple disorders, such as cardiovascular disease, cancer, compromised wound healing, and diabetes. Several studies have reported that stress-induced neuroendocrine immune equilibrium dysfunction is critical for well-being. Stressed people are more likely to have risky behaviours, including irregular sleep schedules, insufficientdiet, less exercise, and a higher risk for the consumption of alcohol, nicotine, and other medications, in addition to the direct effects of anxiety and depression on endocrine and immune function.

Body type

Body form can also influence the healing of wounds. For instance, an obese patient can experience a compromise in wound healing due to low adipose tissue blood supply. In addition, there is some protein malnutrition in some obese patients, which further impedes recovery. Conversely, the absence of oxygen and nutrition stores can interfere with wound healing when a patient is thin and weak.

Chronic diseases

A few of chronic conditions that can compromise wound healing are coronary heart disease, stroke, peripheral vascular disease, and diabetes mellitus. To have the right plan, patients with chronic illness should be monitored closely through their course of care.

Vascular insufficiency

Various wounds or ulcers such as arterial, diabetic pressure, and venous ulcers can affect the lower extremities. Decreased blood supply is the common cause of these ulcers. The clinician must identify the type of ulcer to ensure appropriate topical and supportive therapies.

Nutrition

Food has been recognized for more than 100 years as a very significant aspect that impacts wound healing. The most apparent thing is that malnutrition or specific nutritional shortages following trauma and surgery can have a profound impact on the wound healing. Special nutrients are also needed in patients with chronic or nonhealing wounds and with nutritional deficiencies. The metabolism of energy, carbohydrates, proteins, fats, vitamins, and minerals will all affect the healing process².

TRADITIONAL USE OF MEDICINAL PLANTS IN WOUND HEALING

Medicinal plants are good source of compounds which could serve as leads for drug discovery for wound healing¹⁷. The wound healing activities displayed by medicinal plants are attributed to the presence of bioactive chemicals such as phenols, alkaloids, triterpenes and flavonoids. In wound healing, these bioactive compounds have been reported to have antioxidant and antimicrobial activities, improve collagen deposition and increase the proliferation of both fibroblasts and keratinocytes^{18,19}.

Curcumin

One of the most extensively studied phytochemicals for wound healing is curcumin, which is chemical compound present in the Asian spice turmeric or *curcuma longa*. Apart from usage in Indian and Chinese cuisine, turmeric has been used topically for cutaneous wounds including ulcers, traditionally in the Indian subcontinent²⁰. The main mechanism by which curcumin impacts wound healing is through its anti-inflammatory properties. Invitro studies have been demonstrated the suppression of TNF- α and IL-1 production by human macrophages²¹. Moreover, curcumin is also a potent inhibitor of phosphorylase kinase (PhK)and NF- κ B activation^{22,23}. This make curcumin a great phytochemical candidate for the treatment of hyperinflammatory wounds such as chronic diabetic wounds and burns.

Since curcumin is a hydrophobic compound, its dermal delivery is minimal²⁴. Consequently, different formulations has been created to enhance topical usage of curcumin such as gels²⁵, polymetric bandages²⁶, collagen films²⁷, alginate foams²⁸. Moreover, lipid-core nano capsule (LCN) significantly enhances the dermal delivery of curcumin²⁹. Thus, LCN-based delivery systems of curcumin show significant promise for topical applications of curcumin.

Curcumin and chronic wounds

Chronic wounds are hyper-inflammatory and highly proteolytic environments. Thus, controlling this dysregulated inflammation is crucial to ensure adequate wound healing. Topical curcumin shows promise in the management of chronic non-healing wounds. Topical curcumin treatment of wounds of streptozotocin-induced diabetic rats showed faster re-epithelialization, increased migration of fibroblasts to the wound bed, improved vascularization and significantly higher collagen content than control animals³⁰. Diabetic wounds have diminished angiogenic potential, thus prolonging wound healing³¹. It is interesting to note that topical curcumin treatment on the wounds of diabetic rats also showed enhanced angiogenesis demonstrated by significant upregulation in VEGF³².

Curcumin and hypertrophic scarring

In hypertrophic scarring and keloids, there is an abundance of TGF- β 1 expression, fibroblast proliferation, and excess



collagen and extracellular matrix (ECM) synthesis³³. Apart from being a potent inhibitor of NF- κ B, curcumin inhibits TGF- β 1 signaling in keloid fibroblasts and also diminishes ECM production³⁴. Therefore, topical curcumin may show promise in hypertrophic scar prevention.

Honey

Honey has been a component of traditional medicine in diverse parts around the globe. Oneof its most common usages has been topical treatment for chronic wounds and burns³⁵.

Since the primary components of honey are plant-based, honey has been extensively studied for its phytochemical properties^{36,37}. Studies have found that types of honey may differ in their wound healing properties depending on their phytochemical profile, which depends on their floral sources³⁵. Honey has some antibacterial effects³⁸. This is particularly important for burns and chronic wounds. Apart from its antimicrobial effects, honey also have immunomodulatory effects that are useful for the management of chronic wounds. Honey is also shown to promote angiogenesis and fibroblast proliferation in human clinical trials³⁹.

Honey and chronic wounds

The antibacterial effects of honey, which include both bacteriostatic and bactericidal activities, make it of use to eliminate pathogens whilst having a moist environment favorable to wound healing⁴⁰. In order to achieve wound healing in diabetic ulcers, debridement of old cells and tissues is crucial. Honey contains protease enzymes that facilitate debridement of wounds⁴⁰. However, since chronic wounds have a hyper- inflammatory microenvironment, without controlling inflammation there is little chance of achieving wound repair. Honey exerts its anti-inflammatory effects by the inhibition of cyclooxygenase-2 (cox-2), inducible nitric oxide synthase (iNOS), TNF- α and IL-6 expression⁴¹. Honey is also shown to inhibit MMP9, which may help reduce the degradation of ECM in chronic wounds⁴². Furthermore, honey contains various compounds including flavonoids, phenolic acids, catalase, peroxidase, carotenoids, and ascorbic acid, which possess antioxidant properties that can counteract the abundance of free-radicals found in chronic wounds^{43,44}.

Honey and burns

Honey has been used for burns in various ancient societies. Greek and roman physicians, for instance, used honey for the treatment of burn wounds⁴⁵. In rat models of partialthickness burn injuries, honey formulations shortened the period of epithelialization and increased wound contraction compared to vehicle controls⁴⁶. In humans a systemic review of randomized controlled trials of eight studies comparing the efficacy of honey to silver sulphadiazine-impregnated gauze showed that honey had a superior healing effect⁴⁷. However, this was limited to superficial and partial thickness burns only.

Terminalia chebula

T. chebula is reported to enhance extracellular matrix deposition in granulation tissues in rat excision wound models⁴⁸. *T. chebula* extracts have been shown to enhance growth keratinocytes fibroblasts invitro⁴⁹ and Furthermore, rat wounds treated with T. chebula had significantly reduced lipid peroxide levels, suggesting the antioxidant role of *T. chebula* topical treatment⁴⁸, which was confirmed by electronic spin resonance (ESR)-2,2diphenyl-1-picrylhydrazyl (DPPH) assays^{48,49}. Tannins extracted from T. chebula alsopromote angiogenesis in wounds of rat models shown by the upregulation of vascular endothelial growth factor (VEGF) A expression and increased new vessel formation in the inflammatory phase⁵⁰. It is also possible that the wound healing effects of *T. chebula* are also due to its anti-inflammatory effects.

Chebulagic acid (CA), an antioxidant compound extracted from *T. chebula*, when cultured with macrophages in vitro, significantly suppressed NF- κ B activation as well as TNF- α and cox-2 expression⁵¹. It is possible that topical application of *T. chebula* would be beneficial in hyperinflammatory wounds such as chronic diabetic wounds or burns. Supporting this concept, increased wound healing in streptozotocin-induced diabetic rats with the topical application of *T. chebula* extract has been shown⁵². Moreover, *T. chebula* extract accelerates wound healing in burn wounds in comparison to 1% silver sulfadiazine in rat models⁵³.

Aloe vera

Aloe vera, applied to wounds for over 5000 years by Egyptians, romans, indigenous peoples of Africa Asia, and the Americans, Aloe vera continues to be a first-line treatment for burns, ulcers, and surgical wounds⁵⁴. Aloe vera contains many natural bioactive compounds, including pyrocatechol, saponins, acemannan, anthraquinones, glycosides, oleic acid, phytol, as well as simple and complex water-soluble polysaccharides⁵⁵. Acetone extracts from the leaves of Aloe vera exhibit stronger antimicrobial activity than alcohol and aqueous extracts. Gram-positive bacterial species appear to be more sensitive than gram-negative species to Aloe vera⁵⁶. Compounds with known antimicrobial activity are saponins, acemannan, and anthraquinone derivatives⁵⁷.

Acemannan, a major mucopolysaccharide (mesoglycan) from *Aloe vera*, is a potent stimulator of macrophage and T-cell activity and induces the transcription of proinflammatory mRNAs (including IL-1 α , IL-1 β , IL-6, TNF- α , PGE2, and nitrous oxide)⁵⁸. Mesoglycan moieties bind and captureendogenous mitogen inhibitors and reactive oxygen species and promote phagocytosis. Coincidentally, glycans stabilize secreted cytokines, growth factors, and other bioactives, prolonging their activity. Topically applied acemannan has been reported to significantly reduce the time to wound closure in a rat wound healing model, acting via cyclin D1 and AKT/mTOR signal pathways⁵⁹. *Aloe vera* glycans are also reported to



significantly improve denovo formation of granulation tissue by an unknown mechanism⁶⁰.

Centella asiatica

Centella asiatica, also known as Asiatic pennywort, has been used to promote wound healing for eons⁶¹. Extracts from the aerial parts of *Centella asiatica* are reported to improve he healing of chronic ulcers in sprague-Dawley rats in terms of width, depth, and length⁶¹. Wounds associated with acute radiation dermatitis in rats were observed to heal earlier when treated with extracts from *Centella asiatica* compared to the no-treatment control group⁶².

A triterpene glycoside compound Asiaticoside, isolated from Centella asiatica, is commonly known for its significant wound healing properties that have been studied in normal as well as diabetic wound healing. A topical application of 0.4% solution of asiaticoside over the wound of streptazotocin-induced diabetic rats increased the tensile strength, hydroxyproline content, protein content and epithelialization thereby facilitating the wound healing⁶³. In guinea pig, 0.2% solution of asiaticoside was applied topically which produced an increase in hydroxyproline, tensile strength and quick healing. Asiaticoside promoted angiogenesis in the CAM (chick chorioallantoic membrane) model at 40 µg/disk concentration. Enhanced wound healing activity was achieved by asiaticoside has been attributed to angiogenesis, collagen formation increased remodelling of the collagen matrix and stimulation of glycosaminoglycan synthesis in a rat wound chamber model^{64,65}. Since antioxidants play an important role in the wound healing process, the effects of asiaticoside on the levels of antioxidants in the wound were reported in many researchers to explore the possible mechanism of asiaticoside in wound healing.

Topical application of asiaticoside (0.2%) in cutaneous wounds in rats led to increased enzymatic and nonenzymatic antioxidants such as glutathione peroxidase, superoxide dismutase, catalase, vitamin E and vitamin C (ascorbic acid) in newly formed tissues and decrease in lipid peroxide levels. Studies revealed that asiaticoside enhanced induction of antioxidant levels at an initial phase of wound healing. All these reports indicate that asiaticoside exhibits significant wound healing activity in normal as well as delayed healing models⁶⁶.

Citrullus lanatus

Watermelon (*Citrullus lanatus*) is an important horticultural crop which belongs to the Cucurbitaceae family. Watermelon has been used to treat various ailments, such as cardio- vascular diseases, aging related ailments, obesity, diabetes, ulcers, and various types of cancers. The medicinal properties of watermelon are attributed by the presence of important phytochemicals with pharmaceutical values such as lycopene, citrulline, and other polyphenolic compounds. Watermelon acts as vital source of L-citrulline, a neutral- alpha amino acid which is the precursor of L-arginine, an essential aminoacid necessary for protein synthesis. Supplementation of Lcitrulline and lycopene displayed numerous health benefits in invitro and *in vivo* studies.

In particular, watermelon can be considered as an excellent functional food due to its rich lycopene, vitamin A, vitamin C contents and antioxidant potentials^{67,68}. Bioactive compounds present in watermelon render numerous health benefits, such as decreasing therisk of cardio-vascular disease, aging related ailments, obesity, diabetes, and various cancer alleviating effects have been reported⁶⁹⁻⁷⁴. In 1930, Wada⁷⁵ determined and isolated citrulline, a non-essential amino acid from watermelon which is involved in the synthesis of arginine. The amino acid arginine is vital for the endogenous synthesis of nitric oxide, a crucial signaling molecule involved in various neurological and immune responses in animals and humans⁷⁶.

However, direct intake of L-citrulline and L-arginine could lead to gastro-intestinal discomforts such as nausea and diarrhea^{77,78}. Therefore, the consumption of fruits rich in Lcitrulline (precursor of L-arginine, an essential amino acid for protein synthesis)-such as watermelon is important to obtain the necessary nutrition. Supplementation of whole watermelon in powder form improved the lipid profiles, antioxidant status, and anti- inflammatory properties of high fat fed rats⁷⁹. Moreover, the ingestion of watermelon regulated the expression of genes associated with lipid metabolism⁷⁹. In detail, the augmentation of watermelon and L-arginine enhanced the regulation of hepatic gene expression of endothelial nitric oxide synthase. Nitric oxide (NO) is a ubiquitous signaling molecule vital for the relaxation of blood vessels and it also reduces the atherosclerosis by influencing the lipid metabolism⁷⁹⁻⁸¹. On the other hand, watermelon supplementation downregulated the genes involved in lipid metabolism such as fatty acid synthase (FAS), 3- hydroxy-3-methyl glutaryl-coA reductase (HMGCR), sterol regulatory element binding protein (SERB) 1, SERB 2, cyclooxygenase-2 (COX2), and nuclear factor -кВ (NF-кВ) in rats⁷⁹.

Moreover, watermelon also down regulated the expression of Cox-2 enzyme responsible for the synthesis of pro-inflammatory prostaglandins. Furthermore, Hong *et al.*⁸², illustrated that the watermelon supplementation exhibited similar mechanism to non- steroidal anti-inflammatory drugs that inhibits the activity of Cox-2 and reduces the inflammatory response. Previous studies suggest that the direct influence of NO and L- arginine on the up-regulation of PPAR- γ whereas ulcerative colitis leads to the reduction in the levels of PPAR- $\gamma^{83,84}$. The important enzymes such as Cox-2, iNOS, and NF- κ B involved in the generation of reactive oxygen species are inhibited by PPAR- γ ; on the other hand, the activities of antioxidant enzymes are enhanced by PPAR- γ^{85} .

Similarly, pigments such as lycopene and β -carotene present in watermelon also displayed antioxidant properties. Among the carotenoids, lycopene consists of



strong antioxidants, for instance, the free radical scavenging rate of lycopene is higher in comparison with carotenoids such as β-carotene and tocopherol. According to previous reports, the capability of lycopene to guench the singlet oxygen is ten times higher than tocopherol and two-fold higher than β-carotene^{86,87}. Polyphenolic compounds are vital antioxidants classified into phenolic acids, flavonoids, lignans and stilbenes. According toTilili et al.⁷¹, in watermelon the occurrence of polyphenols is responsible for the hydrophilic antioxidant activity, and the fresh juice of watermelon is reported to have 16.94-20.23 mg Gallic acid equivalent (GAE)/100 mL of polyphenols. Therefore, the intake of watermelon as dietary snack or in beverage form can induce the antioxidant potentials in the human body and helps in the improvement of cell signaling, adhesion, and other biological activities. In addition, the watermelon rind powder consisted of different polyphenolic substances such as 4-hydroxybenzoic acid, vanillin, and coumaric acid⁸⁸. The presence of polyphenolic compounds in watermelon rind powder significantly increased the efficiency of 1,1- diphenyl-2-picrylhydrazyl (DPPH) radical scavenging⁸⁸.

Arctium lappa

Arctium lappa, commonly known as burdock, is a widely herb⁸⁹. Scientific analyses cultivated perennial demonstrate Arctium lappa has antioxidant⁹⁰, antiinflammatory⁹¹, antidiabetic⁹², antimicrobial⁹³, antiviral⁹⁴, anticancer⁹⁵, and hepatoprotective⁹⁶ properties. The root extract of Arctium lappa has been shown to significantly improve dermal ECM metabolism, affecting glycosaminoglycan turnover and reducing visible wrinkles in human skin in vivo⁹⁷. Arctium lappa is also reported to regulate cell adhesion and gene expression in canine dermal fibroblasts, affecting the Wnt/ β -catenin signalling pathway, known to be a key regulator of wound healing⁹⁸. In a pilot study of one commercial preparation including Arctium lappa, Burns and Wounds[™] topical ointment (B&W), pain and healing of first and second-degree burns in humans was demonstrated to be managed more effectively thanthe control treatment⁹⁹.

Commiphora myrrha

Myrrh, the resinous exudate produced by commiphora myrrha¹⁰⁰, has well-documented antioxidant¹⁰¹, antiantibacterial¹⁰³, inflammatory¹⁰², and analgesic¹⁰⁴ activities. Medicinal applications of myrrh include the treatment of gastrointestinal diseases, fractures, arthritis, obesity, parasitic infections, and as an anticoagulant¹⁰⁵⁻¹⁰⁷. Myrrh has been used topically to clean wounds, reduce edema, and provide pain relief (analgesia)¹⁰⁸. Myrrh is commonly used in combination with other ingredients. Galehdari et al. showed that the combination of myrrh, adiantum capillus-veneris, Aloe vera, and Lawsonia inermis, significantly improved wound healing in diabetic mice¹⁰⁹. The short-term application of myrrh effectively reduces pain and controls the recurrence of mouth ulcers in humans¹¹⁰. In common with several other herbal preparations described here, myrrh is found to modify the expression of TGF- β 1 and VEGF in mouse dermal fibroblasts in vitro, suggesting a common mechanism of action¹¹¹.

Camellia sinensis

Green tea, an aqueous extract made from the leaves of camellia sinensis, is revered throughout Asia for its reputed health benefits¹¹². Centuries of anecdotal evidence has been experimentally validated by demonstrating that camellia sinensis has antioxidant¹¹³, anti- inflammatory¹¹⁴, antimicrobial¹¹⁵, anticarcinogenic¹¹⁶, antiaging¹¹⁷, antiobesity^{118,119}, cardioprotective¹²⁰, and neuroprotective¹²¹ activities. Catechins, the polyphenolic compounds from *camellia sinensis*, are primarily responsible for these pharmacological activities¹²². The major catechin, (-)-epigallocatechin-3-gallate (EGCG)¹¹², stimulates the proliferation and differentiation of keratinocytes¹²³. Klass et al. found that EGCG suppresses TGF-β signaling, reducing MMP-1 and MMP-2 expression, and attenuating synthesis of collagen type 1 in human dermal fibroblasts. These properties suggest that EGCG is a potential anti-scarring agent¹²⁴. In addition, EGCG was demonstrated to induce keloid shrinkage¹²⁵ and inhibit growth and pathological features of keloids by suppressing STAT3 signaling¹²⁶. Methanol extracts from camellia sinensis reportedly increase fibroblast proliferation and collagen synthesis¹¹⁵. Furthermore, invivo studies have been demonstrated that camellia sinensis significantly improves wound healing by increasing angiogenesis in rats^{122,127}. Extracts from camellia sinensis are also reported to improvewound healing in a diabetic mouse model¹²⁸.

Cinnamomum cassia

Cinnamomum cassia is a commonly used spice and flavouring agent, and the bark of Cinnamomum cassia is also used to increase blood circulation and as analgesic¹²⁹. Cinnamomum cassia is frequently formulated with other herbs; it is one of the seven botanical components of shexiang baoxan pill (SBP), a well known traditional chinese medicine (TCM) prescribed for chest pain and discomfort associated with coronary artery disease¹³⁰. SBP is currently the subject of a randomized double-blinded clinical trial for the treatment of coronary artery disease not amenable to revascularization¹³¹. Attention is also focussed on SBP anti-inflammatory¹³² and anticancer activities^{133,134}, as well as its impact on hypertension, insulin resistance, and noninsulin-dependent diabetes mellitus¹³⁵. In vitro and In vivo studies indicate that cinnamaldehyde, a bioactive component from Cinnamomum cassia, is a natural insecticide, is an antimicrobial, antidiabetic, antilipidemic, antiinflammatory, and neuroprotective agent¹³⁶, and activates PI3K/AKT and MAPK signaling pathways, increasing VEGF expression, and stimulating angiogenesis in human umbilical veinendothelial cells¹²⁹. Cinnamaldehyde is also reported to improve wound healing in zebrafish¹²⁹.

[©]Copyright protected. Unauthorised republication, reproduction, distribution, dissemination and copying of this document in whole or in part is strictly prohibited.

Hibiscus rosa-sinensis

Hibiscus rosa-sinensis or shoeblack plant, is an evergreen shrub native to tropical south eastern Asia¹³⁷. The flowers of *Hibiscus rosa-sinensis* are edible. Traditional texts describe preparations of the leaves of flowers promote hair growth and prevent greying¹³⁸. Alcoholic extracts of *Hibiscus rosa-sinensis* flowers are claimed to provide women with control of their fertility¹³⁹. Extracts from *Hibiscus rosa-sinensis* have also been found to have antibacterial¹⁴⁰ and wound healing properties¹⁴¹. They attenuate inflammation, enhance fibroblast proliferation, and collagen deposition, as well as upregulate VEGF and TGF- β 1 expression in rat excisional wounds¹⁴².

Paeonia suffruticosa

Paeonia suffruticosa, also known as moutan peony, has been bred for millennia¹⁴³; over 1000 distinct cultivars are now available. The root bark of *Paeonia suffruticosa* is the source for bioactive ingredients used for TCM preparations. Pharmacological investigation of *Paeonia suffruticosa* has demonstrated it has antioxidant¹⁴⁴, neuroprotective¹⁴⁵, antitumour¹⁴⁶, anti-inflammatory¹⁴⁷, and antidiabetic¹⁴⁸ properties. The dried root of *Paeonia suffruticosa* is commonly applied to cracked skin to assist healing and relieve pain¹⁴⁹. When tested invitro at low concentrations (<10 µg/mL), *Paeonia suffruticosa* is found to stimulate the viability and proliferation of human primary dermal fibroblasts and HaCaT keratinocytes, suggesting its potential use as a wound healing therapy¹⁵⁰.

In this review, we show that there is anti-inflammatory and antioxidant phytochemicals are involved in wound healing. The main mechanism by which curcumin impacts wound healing is through its anti-inflammatory properties and it is also a potent inhibitor of phosphorylase kinase (Phk) and NF-KB activation. It also involves in the suppression of TNF- α and IL-1 production by human macrophages. This makes the curcumin effective in hypertrophicscarring and chronic diabetic wounds. Honey involves in wound healing activity as it exerts anti-inflammatory effects by the inhibition of cyclooxygenase-2 (Cox-2), inducible nitric oxide synthase (iNOS), TNF- α and IL-6 expression. Honey has greater wound healing potential in chronic wounds by its antioxidant properties that can counteract the abundance of free-radicals and also by inhibiting MMP9, which may help reduce the degradation of ECM in chronic wounds. Chebulagic acid derived from T. chebula has potent antiinflammatory effects such as the inhibition of Cox-2 and NF-KB. Also, tannic acid strengthenscollagen scaffolds and inhibits the MMP-mediated destruction of ECM which is useful in chronic wounds and hypertrophic scarring. Aloe vera exhibits its wound healing activity by its antimicrobial activity and also by the stimulation of macrophage and T-cell activity and induces the transcription of proinflammatory mRNAs (including IL-1 α , IL-1 β , IL-6, TNF- α , PGE2 and nitrous oxide). Asiaticoside enhances induction of antioxidant levels at an initial phase of wound healing this indicates that asiaticoside exhibits significant wound healing activity in normal as well as delayed healing models. It also increases hydroxy proline content, tensile strength and promote angiogenesis, collagen formation, epithelialization thereby facilitating wound healing. Arctium lappa has been shown to improve dermal ECM metabolism and also regulate cell adhesion and gene expression, affecting the Wnt/ β -catenin signaling pathway which is a key regulator of wound healing. Commiphora myrrha exhibit a common mechanism of action in wound healing by modifying the expression of TGF-B1 and VEGF. Watermelon inhibits the activity of Cox-2 and reduces the inflammatory response similar to the mechanism of nonsteroidal anti-inflammatory drugs. Polyphenolic compounds and pigments such as lycopene and Bcarotene present in watermelon induce antioxidant potentials and helps in the improvement of cell signaling, adhesion, and other biological activities.

Epigallocatechin-3-gallate (EGCG) found in camellia sinensis stimulates the proliferation and differentiation of keratinocytes and suppresses TGF-B receptors by modifying TGF-β signaling, reducing MMP-1 and MMP-2 expression and attenuating synthesis of collagen type 1 in human dermal fibroblasts. These properties suggest that EGCG is a potential anti scarring agent. Cinnamomum cassia enhances wound healing by its antimicrobial antiinflammatory properties and also by activating PI3K/AKT and MAPK signaling, increasing VEGF expression and stimulating angiogenesis. Hibiscus rosa-sinensis involves in woundhealing through its antibacterial properties and also it attenuate inflammation, enhance fibroblast proliferation and collagen deposition as well as upregulate VEGF and TGF-B1 expression. Along with antioxidant and antiinflammatory properties, Paeonia suffruticosa found to stimulate the viability and proliferation of human primary dermal fibroblasts and HaCaT keratinocytes, suggesting its potential use in wound healing.

Wound healing is a biological process that starts with trauma and ends with scar formation. The present review clearly revealed that provides huge number of plants that show significant wound healing activities. These natural substances are rich for the development of alternatives to synthetic drugs. However, there is a need for scientific validation, standardization and safety evaluation of plants of the traditional medicine before these could be recommended for healing of the wounds.

REFERENCES

- Portou, MJ, Baker D, Abraham D, Tsui J. The innate immune system, toll-like receptors and dermal wound healing: A review. Vascul. Pharmacol. 2015; 71: 31–36.
- Guo S, Dipietro LA. Factors affecting wound healing. J. Dent. Res. 2010; 89: 219–229.
- Eming SA, Martin P, and Tomic-Canic M. Wound repair and regeneration: mechanisms, signaling and translation. Science Translational Medicine. 2014; 265.
- Arya AK, Tripathi K, and Das P. Promising role of ANGPTL4 gene in diabetic wound healing. The International Journal of Lower Extremity Wounds 2014; 13(1):58–63.
- Golebiewska EM, Poole AW. Platelet secretion: from haemostasis to wound healing and beyond Blood Reviews. 2015; 29(3):153–



162.

- Enoch S, Leaper DJ. Basic science of wound healing. Surgery (Oxford) 2008; 26(2):31–37.
- Martin P, Leibovich SJ. Inflammatory cells during wound repair: the good, the bad and the ugly. Trends in Cell Biology. 2005; 15(11):599–607.
- Delavary BM, van der Veer WM, van Egmond M, Niessen FB, Beelen RHJ. Macrophages in skin injury and repair. Immunobiology. 2011; 216(7):753–762.
- 9. Kasuya A, Tokura Y. Attempts to accelerate wound healing. Journal of DermatologicalScience. 2014; 76(3):169–172.
- Fan D, Takawale A, Lee J, Kassiri Z. Cardiac fibroblasts, fibrosis and extracellular matrix remodeling in heart disease. Fibrogenesis & Tissue Repair. 2012; 5(1):15.
- 11. Kioka N, Ito T, Yamashita H, *et al*. Crucial role of vinexin for keratinocyte migration in vitro and epidermal wound healing in vivo. Experimental Cell Research 2010; 316(10):1728–1738.
- Sarkar SK, Marmer B, Goldberg G, Neuman KC. Single-molecule tracking of collagenase on native type I collagen fibrils reveals degradation mechanism. Current Biology. 2012; 22(12):1047– 1056.
- 13. Akasaka Y, Ono I, Kamiya T, *et al*. The mechanisms underlying fibroblast apoptosis regulated by growth factors during wound healing. The Journal of Pathology. 2010; 221(3):285–299.
- McDougall S, Dallon J, Sherratt J, Maini P. Fibroblast migration and collagen deposition during dermal wound healing: mathematical modelling and clinical implications. Philosophical Transactions of the Royal Society A: Mathematical, Physical and Engineering Sciences. 2006; 364(1843):1385–1405.
- 15. Schultz GS. Molecular regulation of wound healing in: acute and chronic wounds: nurmngmt. Br, RA (Ed), 2nd edn. 1999; 413–429.
- 16. Nagori BP, Salonki R. Role of medicinal in wound healing. Res J Med Plant. 2011; 5(4):392–405.
- Yahaya E, Cordier W, Steenkamp P, Steenkamp V. Effect of ethnomedicinal extracts used for wound healing on cellular migration and intracellular reactive oxygen species release in SC-1 fibroblasts. South Afr. J. Bot. 2018; 118:11–17.
- Agyare C, Boakye YD, Bekoe EO, Hensel A, Dapaah SO, Appiah T. Review: African medicinal plants with wound healing properties. J. Ethnopharmacol. 2016; 177: 85–100.
- 19. Farahpour MR. Medicinal plants in wound healing. Wound Heal. Curr. Perspect.2019; 2019: 33–47.
- Biswas TK, Mukherjee B. Plant medicines of indian origin for wound healing activity: A review. Int. J. Low Extrem. Wounds. 2003; 2:25–39.
- 21. Chan MM. Inhibition of tumor necrosis factor by curcumin, a phytochemical. Biochem. Pharmacol. 1995; 49:1551–1556.
- Bierhaus A, Zhang Y, Quehenberger P, Luther T, Haase M, Muller M, et al. The dietarypigment curcumin reduces endothelial tissue factor gene expression by inhibiting binding of AP-1 to the DNA and activation of NF-κB. Thromb. Haemost. 1997; 77: 772–782.
- 23. Singh S, Aggarwal BB. Activation of transcription factor NF- κB is suppressed by

curcumin (diferuloylmethane). J. Biol. Chem. 1995; 270: 24995-25000.

- El-Refaie WM, Elnaggar YS, El-Massik MA, Abdallah OY. Novel curcumin-loaded gel- core hyaluosomes with promising burnwound healing potential: Development. in vitro appraisal and in vivo studies. Int. J. Pharm. 2015; 486: 88–98.
- 25. Heng MC. Wound healing in adult skin: Aiming for perfect regeneration. Int. J. Dermatol. 2011; 50:1058–1066.
- Mohanty C, Das M, Sahoo SK. Sustained wound healing activity of curcumin loaded oleic acid based polymeric bandage in a rat model. Mol. Pharm. 2012; 9:2801–2811.
- Gopinath D, Ahmed MR, Gomathi K, Chitra K, Sehgal PK, Jayakumar R. Dermal woundhealing processes with curcumin incorporated collagen films. Biomaterials. 2004; 25:1911–1917.

- Hegge AB, Andersen T, Melvik JE, Bruzell E, Kristensen S, Tonnesen HH. Formulation and bacterial phototoxicity of curcumin loaded alginate foams for wound treatment applications: Studies on curcumin and curcuminoides xlii. J. Pharm. Sci. 2011; 100:174– 185.
- Friedrich RB, Kann B, Coradini K, Offerhaus HL, Beck RC, Windbergs M. Skin penetration behavior of lipid-core nanocapsules for simultaneous delivery of resveratrol and curcumin. Eur. J. Pharm. Sci. 2015; 78:204–213.
- Sidhu GS, Mani H, Gaddipati JP, Singh AK, Seth P, Banaudha KK, et al. Curcumin enhances wound healing in streptozotocin induced diabetic rats and genetically diabetic mice. Wound Repair Regen. 1999; 7:362–374.
- Martin A, Komada MR, Sane DC. Abnormal angiogenesis in diabetes mellitus. Med. Res. Rev. 2003; 23:117–145.
- Kant V, Gopal A, Kumar D, Pathak NN, Ram M, Jangir BL, et al. Curcumin-induced angiogenesis hastens wound healing in diabetic rats. J. Surg. Res. 2015; 193: 978–988.
- Arno AI, Amini-Nik S, Blit PH, Al-Shehab M, Belo C, Herer E, *et al.* Effect of human wharton's jelly mesenchymal stem cell paracrine signaling on keloid fibroblasts. StemCells Transl. Med. 2014; 3: 299–307.
- 34. Hsu YC, Chen MJ, Yu YM, Ko SY, Chang CC. Suppression of TGF- β 1/smad pathway andextracellular matrix production in primary keloid fibroblasts by curcuminoids: Its potential therapeutic use in the chemoprevention of keloid. Arch. Dermatol. Res. 2010; 302: 717–724.
- 35. Majtan J. Honey: An immunomodulator in wound healing. Wound Repair Regen. 2014; 22: 187–192.
- Alvarez-Suarez JM, Gasparrini M, Forbes-Hernandez TY, Mazzoni L, Giampieri F. The composition and biological activity of honey: A focus on manuka honey. Foods. 2014;3: 420–432.
- Jerkovic I, Kus PM, Tuberoso CI, Sarolic M. Phytochemical and physical-chemical analysis of polish willow (Salix spp.) honey: Identification of the marker compounds. Food Chem. 2014; 145: 8–14.
- Blair SE, Cokcetin NN, Harry EJ, Carter DA. The unusual antibacterial activity of medical-grade leptospermum honey: Antibacterial spectrum, resistance and transcriptome analysis. Eur. J. Clin. Microbiol. Infect. Dis. 2009; 28:1199–1208.
- Molan PC. Potential of honey in the treatment of wounds and burns. Am. J. Clin. Dermatol. 2001; 2:13–19.
- Alam F, Islam MA, Gan SH, Khalil MI. Honey: A potential therapeutic agent for managing diabetic wounds. Evid. Based Complement. Altern. Med. 2014; 2014: 169130.
- 41. Hussein SZ, Mohd Yusoff K, Makpol S, Mohd Yusof YA. Gelam honey inhibits the production of proinflammatory, mediators NO, PGE2, TNF- α , and IL-6 in carrageenan-induced acute paw edema in rats. Evid. Based Complement. Altern. Med. 2012; 2012: 109636.
- Majtan J, Bohova J, Garcia-Villalba R, Tomas-Barberan FA, Madakova Z, Majtan T, *et al*. Fir honeydew honey flavonoids inhibit TNF-β-induced MMP-9 expression in human keratinocytes: A new action of honey in wound healing. Arch. Dermatol. Res. 2013; 305: 619–627.
- Henriques A, Jackson S, Cooper R, Burton N. Free radical production and quenching in honeys with wound healing potential. J. Antimicrob. Chemother. 2006; 58:773–777.
- Gheldof N, Wang XH, Engeseth NJ. Identification and quantification of antioxidant components of honeys from various floral sources. J. Agric. Food Chem. 2002; 50:5870–5877.
- Pecanac M, Janjic Z, Komarcevic A, Pajic M, Dobanovacki D, Miskovic SS. Burnstreatment in ancient times. Med. Pregl. 2013; 66: 263–267.
- Iftikhar F, Arshad M, Rasheed F, Amraiz D, Anwar P, Gulfraz M. Effects of acacia honey on wound healing in various rat models. Phytother. Res. 2010; 24: 583–586.
- 47. Wijesinghe M, Weatherall M, Perrin K, Beasley R. Honey in the



International Journal of Pharmaceutical Sciences Review and Research

treatment of burns: A systematic review and meta-analysis of its efficacy. N. Z. Med. J. 2009; 122: 47–60.

- Suguna L, Singh S, Sivakumar P, Sampath P, Chandrakasan G. Influence of terminalia chebula on dermal wound healing in rats. Phytother. Res. 2002; 16: 227–231.
- Singh D, Singh D, Choi SM, Zo SM, Painuli RM, Kwon SW, et al. Effect of extracts of terminalia chebula on proliferation of keratinocytes and fibroblasts cells: An alternative approach for wound healing. Evid. Based Complement. Altern. Med. 2014; 2014;701656.
- Li K, Diao Y, Zhang H, Wang S, Zhang Z, Yu B, *et al.* Tannin extracts from immature fruits of terminalia chebula fructus retz. Promote cutaneous wound healing in rats. BMC Complement. Altern. Med. 2011; 11: 86.
- Reddy DB, Reddanna P. Chebulagic acid (CA) attenuates LPSinduced inflammationby suppressing NF-κB and MAPK activation in raw 264.7 macrophages. Biochem. Biophys. Res. Commun. 2009; 381:112–117.
- Soni R, Mehta NM, Srivastava DN. Healing potential of ethyl acetate soluble fraction of ethanolic extract of terminalia chebula on experimental cutaneous wounds in streptozotocin induced diabetic rats. Asian J. Biomed. Pharm. Sci. 2013; 3:32–36.
- Nasiri E, Hosseinimehr SJ, Azadbakht M, Akbari J, Enayati-Fard R, Azizi S. The effect of terminalia chebula extract vs. Silver sulfadiazine on burn wounds in rats. J. Complement. Integr. Med. 2015; 12:127–135.
- Garcia-Orue I, Gainza G, Gutierrez FB, et al. Novel nanofibrous dressings containing rhEGF and Aloe vera for wound healing applications. International Journal of Pharmaceutics. 2017; 523(2):556–566.
- 55. Salehi B, Albayrak S, Antolak H, *et al*. Aloe genus plants: from farm to food applications and phytopharmacotherapy. International Journal of Molecular Sciences. 2018; 19(9):2843.
- Lawrence R, Tripathi P, Jeyakumar E. Isolation, purification and evaluation of antibacterial agents from *Aloe vera*. Brazilian Journal of Microbiology. 2009; 40(4):906–915.
- Mart'inez-Romero D, Alburquerque N, Valverde JM, et al. Postharvest sweet cherry quality and safety maintenance by Aloe vera treatment: a new edible coating. Postharvest Biology and Technology. 2006; 39(1):93–100.
- Ali P, Chen YF, Sargsyan E. Chapter 12-bioactive molecules of herbal extracts with anti-infective and wound healing properties. in Microbiology for Surgical Infections 2014; 205–220.
- Xing W, Guo W, Zou CH, *et al*. Acemannan accelerates cell proliferation and skin wound healing through AKT/ mTOR signaling pathway. Journal of Dermatological Science. 2015; 79(2):101–109.
- 60. Jettanacheawchankit S, Sasithanasate S, Sangvanich P, Banlunara W, Thunyakitpisal
- P. Acemannan stimulates gingival fibroblast proliferation; expressions of keratinocyte growth factor-1, vascular endothelial growth factor, and type I collagen;and wound healing. Journal of Pharmacological Sciences. 2009; 109(4):525-531.
- Somboonwong J, Kankaisre M, Tantisira B, Tantisira MH. Wound healing activities of different extracts of *Centella asiatica* in incision and burn wound models: an experimental animal study, BMC Complementary and Alternative Medicine. 2012; 12:103.
- 62. Chen YJ, Dai YS, Chen BF, *et al.* The effect of tetrandrine and extracts of *Centella asiatica* on acute radiation dermatitis in rats. Biological & Pharmaceutical Bulletin. 1999; 22(7):703–706.
- Maquart FX, chastang F, Simeon A, Birembaut P, Gillery P. Triterpenes from *Centella asiatica* stimulate extracellular matrix accumulation in rat experimental wounds. EurJ Dermatol. 1999; 9(4):289-296.
- Rosen H, Blumenthal A, McCallum J. Effect of asiaticoside on wound healing in the rat. Proc Soc Exp Biol Med. 1967; 125(1): 279-280.
- 65. Liu M, Dai Y, Li Y, Luo Y, Huang F, *et al*. Madecassoside isolated from *Centella asiatica* herbs facilitates burn wound healing in

mice. Planta Med. 2008; 74(8): 809-15.

- Shukla A, Rasik AM, Dhawan BN. Asiaticoside-induced elevation of antioxidant levels in healing wounds. Phytother Res. 1999;13(1):50-54.
- 67. U.S. Department of Agriculture ARS. USDA National Nutrient Database for Standard Reference, Release 27; 2015.
- 68. Rimando AM, Perkins-Veazie PM. Determination of citrulline in watermelon rind. J. Chromatogr. A 2005; 1078:196–200.
- 69. Rao AV, Agarwal S. Role of antioxidant lycopene in cancer and heart disease. J. Am. Coll. Nutr. 2000; 19:563–569.
- Romero MJ, Platt DH, Caldwell RB, Caldwell RW. Therapeutic use of citrulline in cardiovascular disease. Cardiovasc. Drug Rev. 2006; 24:275–290.
- Tlili I, Hdider C, Lenucci MS, Riadh I, Jebari H, Dalessandro G. Bioactive compounds and antioxidant activities of different watermelon (Citrullus lanatus (Thunb.) Mansfeld) cultivars as affected by fruit sampling area. J. Food Compos. Anal. 2011; 24:307–314.
- Tomes ML, Johnson KW, Hess M. The carotene pigment content of certain red fleshed watermelons. Inproc. Am. Soc. Hortic. Sci. 1963; 82:460–464.
- Wan X, Liu W, Yan Z, Zhao S, He N, Liu P. Changes of the contents of functional substances including lycopene, citrulline and ascorbic acid during watermelon fruits development. Sci. Agric. Sin. 2011; 44:2738–2747.
- Tarazona-Díaz MP, Viegas J, Moldao-Martins M, Aguayo E. Bioactive compounds from flesh and by-product of fresh-cut watermelon cultivars. J. Sci. Food Agric. 2011; 91:805–812.
- Wada M. Uber Citrullin, eine neue Aminosaure im PreBsaft der Wassermelone, Citrullus vulgaris schrad. Biochemische Zeitschrift. 1930; 224;420–429.
- 76. Wu G, Collins JK, Perkins VP, Siddiq M, Dolan KD, Kelly KA, et al. Dietary supplementation with watermelon pomace juice enhances arginine availability and ameliorates the metabolic syndrome in Zucker diabetic fatty rats. J. Nutr. 2007; 137:2680–2685.
- Evans RW, Fernstrom JD, Thompson J, Morris SM, Kuller LH. Biochemical responses of healthy subjects during dietary supplementation with l-arginine. J. Nutr. Biochem. 2004; 15:534– 539.
- Wu G, Meininger CJ. Arginine nutrition and cardiovascular function. J. Nutr. 2000; 130: 2626–2629.
- Hong MY, Hartig N, Kaufman K, Hooshmand S, Figueroa A, Kern M. Watermelon consumption improves inflammation and antioxidant capacity in rats fed an atherogenic diet. Nutr. Res. 2015; 35:251– 258.
- Jobgen WS, Fried SK, Fu WJ, Meininger CJ, Wu G. Regulatory role for the arginine– nitric oxide pathway in metabolism of energy substrates. J. Nutr. Biochem. 2006; 17:571–588.
- Jobgen W, Fu WJ, Gao H, Li P, Meininger CJ, Smith SB, *et al*. High fat feeding and dietary l-arginine supplementation differentially regulate gene expression in rat white adipose tissue. Amino Acids. 2009; 37:187–198.
- Hong MY, Tseng YT, Kalaba M, Beidler J. Effects of watermelon powder supplementation on colitis in high-fat diet-fed and dextran sodium sulfate-treated rats. J. Funct. Foods. 2019; 54:520–528.
- Ptasinska A, Wang S, Zhang J, Wesley RA, Danner RL. Nitric oxide activation ofperoxisome proliferator-activated receptor gamma through a p38 MAPK signaling pathway. FASEB J. 2007; 21:950– 961.
- Liu Y, Huang J, Hou Y, Zhu H, Zhao S, Ding B, *et al*. Dietary arginine supplementation alleviates intestinal mucosal disruption induced by Escherichia coli lipopolysaccharide in weaned pigs. Br. J. Nutr. 2008; 100:552–560.
- Cai W, Yang T, Liu H, Han L, Zhang K, Hu X, *et al.* Peroxisome proliferator-activated receptor γ (PPARγ): A master gatekeeper in CNS injury and repair. Prog. Neurobiol. 2018; 163:27–58.
- 86. Naz A, Butt MS, Sultan MT, Qayyum MM, Niaz RS. Watermelon



International Journal of Pharmaceutical Sciences Review and Research

lycopene and allied health claims. EXCLI J. 2014; 13:650.

- Kyriacou MC, Leskovar DI, Colla G, Rouphael Y. Watermelon and melon fruit quality: The genotypic and agro-environmental factors implicated. Sci. Hortic. 2018; 234:393–408.
- Al-Sayed HM, Ahmed AR. Utilization of watermelon rinds and sharlyn melon peels as a natural source of dietary fiber and antioxidants in cake. Ann. Agric. Sci. 2013; 58:83–95.
- Lin SC, Lin CH, Lin CC, *et al.* Hepatoprotective effects of *Arctium lappa* linne on liver injuries induced by chronic ethanol consumption and potentiated by carbon tetrachloride. Journal of Biomedical Science. 2002; 9(5):401–409.
- Fierascu RC, Georgiev MI, Fierascu I, *et al.* Mitodepressive, antioxidant, antifungal and anti-inflammatory effects of wildgrowing Romanian native *Arctium lappa* L. (Asteraceae) and Veronica persica Poiret (Plantaginaceae). Food and Chemical Toxicology. 2018; 111:44–52.
- de Almeida ABA, Sanchez-Hidalgo M, Martın AR, et al. Antiinflammatory intestinal activity of Arctium lappa L. (Asteraceae) in TNBS colitis model. Journal of Ethnopharmacology. 2013; 146(1):300–310.
- Ahangarpour A, Heidari H, Oroojan AA, Mirzavandi F, Nasr Esfehani K, DehghanMohammadi Z. Antidiabetic, hypolipidemic and hepatoprotective effects of *Arctium lappa* root's hydroalcoholic extract on nicotinamide-streptozotocin induced type 2 model of diabetes in male mice. Avicenna Journal of Phytomedicine. 2017; 7(2):169–179.
- Pereira JV, Bergamo DCB, Pereira J, França SDC, Pietro RCLR, Silva-Sousa YTC. Antimicrobial activity of *Arctium lappa* constituents against microorganisms commonly found in endodontic infections. Brazilian Dental Journal. 2005; 16(3):192–196.
- Dias MM, Zuza O, Riani LR, *et al.* In vitro schistosomicidal and antiviral activities of *Arctium lappa* L. (Asteraceae) against Schistosoma mansoni and Herpes simplex virus-1. Biomedicine & Pharmacotherapy. 2017; 94:489–498.
- Sun Q, Liu K, Shen X, *et al*. Lappaol F, a novel anticancer agent isolated from plant *Arctium lappa* L. Molecular Cancer Berapeutics. 2014;13(1):49–59.
- de Souza Predes F, da Silva Diamante MA, Foglio MA, et al. Hepatoprotective effectof Arctium lappa root extract on cadmium toxicity in adult Wistar rats. Biological Trace Element Research. 2014; 160(2):250–257.
- 97. Knott A, Reuschlein K, Mielke H, *et al*. Natural *Arctium lappa* fruit extract improves the clinical signs of aging skin. Journal of Cosmetic Dermatology. 2008; 7(4):281–289.
- Pomari E, Stefanon B, Colitti M. Effect of Arctium lappa (burdock) extract on canine dermal fibroblasts. Veterinary Immunology and Immunopathology. 2013; 156(3- 4):159–166.
- Amish Burn Study G, Kolacz NM, Jaroch MT, et al. The effect of Burns & Wounds (B&W)/burdock leaf therapy on burn-injured Amish patients: a pilot study measuringpain levels, infection rates, and healing times. Journal of Holistic Nursing. 2014; 32(4):327– 340.
- Walsh ME, Reis D, Jones T. Integrating complementary and alternative medicine: use of myrrh in wound management. Journal of Vascular Nursing. 2010; 28(3):102.
- Fatani AJ, Alrojayee FS, Parmar MY, Abuohashish HM, Ahmed MM, Al-Rejaie SS. Myrrh attenuates oxidative and inflammatory processes in acetic acid-induced ulcerative colitis. Experimental and Berapeutic Medicine. 2016; 12(2):730–738.
- Manjula N, Gayathri B, Vinaykumar KS, Shankernarayanan NP, Vishwakarma RA, Balakrishnan A. Inhibition of MAP kinases by crude extract and pure compound isolated from Commiphora mukul leads to down regulation of TNF-α, IL-1β and IL-2. International Immunopharmacology. 2006; 6(2):122–132.
- Shuaib M, Ali A, Ali M, Panda B, Ahmad M. Antibacterial activity of resin rich plant extracts. Journal of Pharmacy and Bioallied Sciences. 2013; 5(4):265–269.
- 104. Shalaby M, Hammouda A. Analgesic, anti-inflammatory and

antihyperlipidemic activities of *Commiphora molmol* extract (myrrh). Journal of Intercultural Ethnopharmacology. 2014; 3(2):56–62.

- Al-Harbi MM, Qureshi S, Raza M, Ahmed MM, Afzal M, Shah AH. Gastric antiulcer and cytoprotective effect of *Commiphora molmol* in rats. Journal of Ethnopharmacology. 1997; 55(2):141–150.
- Abdul-Ghani RA, Loutfy N, Hassan A. Myrrh and trematodoses in Egypt: an overviewof safety, efficacy and effectiveness profiles. Parasitology International. 2009; 58(3):210–214.
- Shen T, Li GH, Wang XN, Lou HX. The genus Commiphora: a review of its traditional uses, phytochemistry and pharmacology. Journal of Ethnopharmacology. 2012; 142 (2):319–330.
- 108. Nomicos EYH. Myrrh, Holistic Nursing Practice. 2007; 21(6):308–323.
- 109. Galehdari H, Negahdari S, Kesmati M, Rezaie A, Shariati G. Effect of the herbal mixture composed of *Aloe vera*, Henna, Adiantum capillus-veneris, and Myrrha on wound healing in streptozotocininduced diabetic rats. BMC Complementary and Alternative Medicine. 2016; 16(1):386.
- Mansour G, Ouda S, Shaker A, Abdallah HM. Clinical efficacy of new *Aloe vera*- and myrrh-based oral mucoadhesive gels in the management of minor recurrent aphthous stomatitis: a randomized, double-blind, vehicle-controlled study. Journal of Oral Pathology & Medicine. 2014; 43(6):405–409.
- 111. Negahdari S, Galehdari H, Kesmati M, Rezaie A, Shariati G. Wound healing activity of extracts and formulations of *Aloe vera*, Henna, Adiantum capillus-veneris, and Myrrh on mouse dermal fibroblast cells. International Journal of Preventive Medicine. 2017; 8:18.
- 112. Yang CS, Chen G, and Wu Q. "Recent scientific studies of a traditional Chinese medicine, tea, on prevention of chronic diseases." Journal of Traditional and Complementary Medicine. 2014; 4(1):17–23.
- 113. Espinosa C, Lopez-Jimenez JA, Perez-Llamas F *et al.* "Long-term intake of white tea prevents oxidative damage caused by adriamycin in kidney of rats." Journal of the Science of Food and Agriculture. 2016; 96(9):3079–3087.
- 114. Chen BT, Li WX, He RR *et al.* "Anti-inflammatory effects of a polyphenols-rich extract from tea (Camellia sinensis) flowers in acute and chronic mice models," Oxidative Medicine and Cellular Longevity. Article ID 537923. 2012; 2012:7.
- 115. Anwar Ibrahim D and Noman Albadani R. "Evaluation of the potential nephroprotective and antimicrobial effect of Camellia sinensis leaves versus Hibiscus sabdariffa (in vivo and in vitro studies)." Advances in Pharmacological Sciences. Article ID 389834. 2014; 2014:5.
- Er S and Dikmen M. "Camellia sinensis increased apoptosis on U2OS osteosarcoma cells and wound healing potential on NIH3T3 fibroblast cells," Cytotechnology. 2017; 69(6):901–914.
- 117. Jadoon S, Karim S, Bin Asad MHH *et al.* "Anti-aging potential of phytoextract loaded-pharmaceutical creams for human skin cell longetivity," Oxidative Medicine and Cellular Longevity. Article ID 709628. 2015; 2015:17.
- He RR, Chen L, Lin BH, Matsui Y, Yao XS, Kurihara H. "Beneficial effects of oolong teaconsumption on diet-induced overweight and obese subjects." Chinese Journal of Integrative Medicine. 2009; 15(1):34–41.
- 119. Hasani-Ranjbar S, Jouyandeh Z, Abdollahi M. "A systematic review of anti-obesity medicinal plants-an update." Journal of Diabetes & Metabolic Disorders. 2013; 12(1):28.
- Khan G, Haque SE, Anwer T, Ahsan MN, Safhi MM, Alam MF. "Cardioprotective effect of green tea extract on doxorubicininduced cardiotoxicity in rats." Acta Poloniae Pharmaceutica. 2014; 71(5):861–868.
- Levites Y, Weinreb O, Maor G, Youdim MBH, Mandel S. "Green tea polyphenol (-)- epigallocatechin-3- gallate prevents N-methyl-4phenyl-1,2,3,6-tetrahydropyridine- induced dopaminergic neurodegeneration." Journal of Neurochemistry. 2001; 78(5):1073–1082.

115

R

International Journal of Pharmaceutical Sciences Review and Research

Available online at www.globalresearchonline.net

©Copyright protected. Unauthorised republication, reproduction, distribution, dissemination and copying of this document in whole or in part is strictly prohibited.

- 122. Hajiaghaalipour F, Kanthimathi MS, Abdulla MA, Sanusi J. "The effect of *Camellia sinensis* on wound healing potential in an animal model," Evidence-Based Complementary and Alternative Medicine. 2013; 2013:7.
- Hsu S, Bollag WB, Lewis J *et al.* "Green tea polyphenols induce differentiation and proliferation in epidermal keratinocytes," Journal of Pharmacology and Experimental Berapeutics. 2003; 306(1):29–34.
- 124. Klass BR, Branford OA, Grobbelaar AO, Rolfe KJ. "The effect of epigallocatechin-3- gallate, a constituent of green tea, on transforming growth factor-β1-stimulated wound contraction," Wound Repair and Regeneration. 2010; 18(1):80–88.
- Syed F, Bagabir RA, Paus R, Bayat A. "Ex vivo evaluation of antifibrotic compounds inskin scarring: EGCG and silencing of PAI-1 independently inhibit growth and induce keloid shrinkage," Laboratory Investigation. 2013; 93(8):946–960.
- 126. Park G, Yoon BS, Moon JH *et al.* "Green tea polyphenol epigallocatechin-3-gallate suppresses collagen production and proliferation in keloid fibroblasts via inhibition of the STAT3-signaling pathway." Journal of Investigative Dermatology. 2008; 128(10):2429–2441.
- 127. Asadi SY, Parsaei P, Karimi M *et al.* "Effect of green tea (*Camellia sinensis*) extract on healing process of surgical wounds in rat." International Journal of Surgery. 2013; 11(4): 332–337.
- 128. Kim H, Kawazoe T, Han DW *et al.* "Enhanced wound healing by an epigallocatechin gallate-incorporated collagen sponge in diabetic mice," Wound Repair and Regeneration. 2008; 16(5):714–720.
- 129. Yuan X, Han L, Fu P *et al.* "Cinnamaldehyde accelerates wound healing by promotingangiogenesis via up-regulation of PI3K and MAPK signaling pathways," Laboratory Investigation. 2018; 98(6):783–798.
- 130. Zhang KJ, Zhu JZ, Bao XY, Zheng Q, Zheng GQ, Wang Y. "Shexiang baoxin pills for coronary heart disease in animal models: preclinical evidence and promoting angiogenesis mechanism," Frontiers in Pharmacology. 2017; 8:404.
- 131. Tian PP, Li J, Gao J, Li Y. "Efficacy and safety of the Shexiang baoxin Pill for the treatment of coronary artery disease not amenable to revascularisation: study protocol for a randomised, placebocontrolled, double-blinded trial," BMJ Open. 2018; 8(2):18-26.
- Lee SH, Lee SY, Son DJ et al. "Inhibitory effect of 2'hydroxycinnamaldehyde on nitric oxide production through inhibition of NF-κB activation in RAW 264.7 cells," Biochemical Pharmacology. 2005; 69(5):791–799.
- 133. Koppikar SJ, Choudhari AS, Suryavanshi SA *et al.* "Aqueous cinnamon extract (ACE-c)from the bark of Cinnamomum cassia causes apoptosis in human cervical cancercell line (SiHa) through loss of mitochondrial membrane potential," BMC Cancer. 2010; 10:210.
- Kwon HK, Jeon WK, Hwang JS *et al.* "Cinnamon extract suppresses tumor progression by modulating angiogenesis and the effector function of CD8 + Tcells," Cancer Letters. 2009; 278(2):174–182.
- 135. Ye H, Du J, Shen D *et al.* "[Effect of *Shexiang baoxin* pill on the function of vascular endothelium in patients with diabetes mellitus type 2 complicated with angina pectoris]." 2004; 24(12):1077–1079.
- 136. Rao PV, Gan SH. "Cinnamon: a multifaceted medicinal plant,"

Evidence-Based Complementary and Alternative Medicine. 2014; 2014:12.

- Bhaskar A, Nithya V. "Evaluation of the wound-healing activity of *Hibiscus rosa sinensis* L (Malvaceae) in Wistar albino rats," Indian Journal of Pharmacology. 2012; 44(6):694–698.
- 138. Adhirajan N, Ravi Kumar T, Shanmugasundaram N, Babu M. "In vivo and in vitro evaluation of hair growth potential of Hibiscus rosa-sinensis Linn.," Journal of Ethnopharmacology. 2003; 88(2-3):235–239.
- Jadhav VM, Kamble SS, Kadam VJ. "Herbal medicine: Syzygium cumini: a review," Journal of Pharmacy Research. 2009; 2(8):1212–1219.
- 140. Khan ZA, Naqvi SA, Mukhtar A *et al.* "Antioxidant and antibacterial activities of Hibiscus Rosa-sinensis Linn flower extracts," Pakistan Journal of PharmaceuticalSciences. 2014; 27(3):469–474.
- 141. Shivananda Nayak B, Sivachandra Raju S, Orette FA, Chalapathi Rao AV. "Effects of *Hibiscus rosa sinensis* L (Malvaceae) on wound healing activity: a preclinical study in a sprague dawley rat," The International Journal of Lower Extremity Wounds. 2007; 6(2):76– 81.
- Shen HM, Chen C, Jiang JY et al. "The N-butyl alcohol extract from Hibiscus rosa- sinensis L. flowers enhances healing potential on rat excisional wounds," Journal of Ethnopharmacology. 2017; 198:291–301.
- 143. Zhou SL, Zou XH, Zhou ZQ *et al.* "Multiple species of wild tree peonies gave rise to the 'king of flowers', Paeonia suffruticosa Andrews," Proceedings of the Royal Society B: Biological Sciences. 2014; 281(1797).
- 144. Rho S, Chung HS, Kang M *et al.* "Inhibition of production of reactive oxygen species and gene expression profile by treatment of ethanol extract of Moutan Cortex Radicis in oxidative stressed PC12 cells," Biological & Pharmaceutical Bulletin. 2005; 28(4):661–666.
- 145. Kim HG, Park G, Piao Y *et al.* "Effects of the root bark of Paeonia suffruticosa on mitochondria-mediated neuroprotection in an MPTP-induced model of Parkinson's disease," Food and Chemical Toxicology. 2014; 65:293–300.
- Xing G, Zhang Z, Liu J, Hu H, Sugiura N. "Antitumor effect of extracts from moutan cortex on DLD-1 human colon cancer cells in vitro," Molecular Medicine Reports. 2010; 3(1):57–61.
- Wu M, Gu Z. "Screening of bioactive compounds from moutan cortex and their anti-inflammatory activities in rat synoviocytes," Evidence-Based Complementary and Alternative Medicine. 2009; 6(1):63.
- 148. Hong H, Wang QM, Zhao ZP *et al.* "Studies on antidiabetic effects of cortex Moutan polysaccharide-2b in type 2 diabetes mellitus rats." 2003; 38(4):255–259.
- 149. He DY, Dai SM. "Anti-inflammatory and immunomodulatory effects of *paeonia lactiflora* pall., a traditional Chinese herbal medicine," Frontiers in Pharmacology. 2011; 2:10.
- 150. Wang R, Lechtenberg M, Sendker J, Petereit F, Deters A, Hensel A. "Wound-healing plants from TCM: in vitro investigations on selected TCM plants and their influence on human dermal fibroblasts and keratinocytes," Fitoterapia. 2013; 84:308–317.

Source of Support: The author(s) received no financial support for the research, authorship, and/or publication of this article.

Conflict of Interest: The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

For any question relates to this article, please reach us at: globalresearchonline@rediffmail.com
New manuscripts for publication can be submitted at: submit@globalresearchonline.net and submit_jpsrr@rediffmail.com



International Journal of Pharmaceutical Sciences Review and Research

Available online at www.globalresearchonline.net

©Copyright protected. Unauthorised republication, reproduction, distribution, dissemination and copying of this document in whole or in part is strictly prohibited