



Polyphenols as Next-Generation Bioactive: Integrating Plant Metabolism, Human Health, and Biotechnological Innovation

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ABSTRACT

Polyphenols are a diverse class of plant-derived compounds characterized by multiple phenolic structures, contributing significantly to their wide-ranging biological activities. This review provides a comprehensive overview of polyphenol chemistry, classification, biosynthesis, dietary sources, and mechanisms of action. Major subclasses include flavonoids and non-flavonoids such as phenolic acids, stilbenes, and lignans, synthesized primarily via the shikimic acid and phenylpropanoid pathways. Polyphenols exhibit potent antioxidant, anti-inflammatory, antimicrobial, anticancer, cardioprotective, and neuroprotective effects through modulation of key molecular pathways, including NF- κ B and Nrf2 signaling. Despite strong preclinical evidence, their clinical application is limited by poor bioavailability and rapid metabolism. Advances in nanotechnology-based delivery systems, microbiome research, and precision nutrition are emerging to overcome these challenges. Overall, polyphenols represent a promising area in biotechnology and therapeutic development, with significant potential for improving human health and preventing chronic diseases.

Keywords: Polyphenols, Antioxidant activity, Phenylpropanoid pathway, Bioavailability, Therapeutic potential.

1. INTRODUCTION

1.1 Definition and Chemical Overview

Polyphenols constitute a heterogeneous category of naturally occurring organic compounds characterized by the presence of at least one aromatic ring bearing one or more hydroxyl substituents, with the majority of biologically relevant members possessing multiple phenolic units. The term “polyphenol” was formally defined by Quideau and colleagues as molecules of plant origin that possess at least two phenol units, distinguishing them from simple phenols and from other phenolic metabolites that lack the structural complexity associated with pronounced biological activity. This molecular architecture, while seemingly straightforward, gives rise to an exceptional degree of chemical diversity: variations in the number and arrangement of hydroxyl groups, the degree of methylation, glycosylation, acylation, and polymerization collectively generate the thousands of structurally distinct compounds that populate the polyphenol class ¹.

1.2 Historical Background

The scientific investigation of plant polyphenols has its intellectual roots in nineteenth-century organic chemistry, when chemists such as Braconnot and Chevreul began isolating and characterizing tannins and other phenolic substances from bark and fruit sources. However, it was not

until the mid-twentieth century, with the development of paper chromatography and ultraviolet spectroscopy, that systematic classification of the flavonoid subclasses became feasible. The discovery in the 1990s that red wine consumption correlated inversely with cardiovascular disease incidence in French populations — the so-called “French Paradox” documented by Renaud and de Lorgeril — catalysed a major expansion of research interest into the biological activities of dietary polyphenols, particularly resveratrol and the flavonols of grape skin. Since that period, advances in mass spectrometry, genomics, and cell biology have enabled increasingly precise mechanistic characterization of polyphenol activity at the molecular level ².

1.3 Importance in Plants and Human Health

Within the context of plant biology, polyphenols fulfil indispensable roles in structural support, pigmentation, chemical signaling, and environmental stress adaptation. Lignins, which are among the most abundant polyphenols on Earth, provide the mechanical rigidity of plant cell walls and contribute to vascular tissue integrity. Anthocyanins confer the red, violet, and blue pigmentation of flowers and fruits, attracting pollinators and seed-dispersing fauna. Condensed tannins serve as feeding deterrents against herbivores, while stilbenes and phytoalexins are induced in response to pathogen challenge. In the context of human



health, epidemiological evidence accumulated over several decades has consistently associated high dietary polyphenol intake with reduced risk of cardiovascular disease, type 2 diabetes mellitus, neurodegenerative disorders, and certain malignancies. The mechanistic basis for these associations is multifaceted and continues to be actively investigated, involving direct antioxidant activity, modulation of gene expression through epigenetic mechanisms, and interaction with the gut microbiome³.

1.4 Scope and Objectives

The present review was undertaken to provide an integrated, graduate-level account of polyphenol science that bridges fundamental chemistry and biosynthetic biology with applied biotechnology and clinical nutrition. The scope encompasses structural classification, biosynthetic pathways, natural sources, physicochemical properties, biological activities, mechanisms of action, extraction and analytical techniques, bioavailability, and current and emerging biotechnological applications. Particular attention is given to recent developments in nanotechnology-based delivery systems, the gut microbiome's role in polyphenol bioactivation, and the prospects for precision nutritional interventions based on individual metabolic phenotyping⁴.

2. CLASSIFICATION OF POLYPHENOLS

2.1 Flavonoids

Flavonoids constitute the largest and most extensively studied subclass of polyphenols, with over six thousand individual compounds identified to date. Their shared structural framework consists of a fifteen-carbon diphenylpropane skeleton (C6-C3-C6) comprising two benzene rings (designated rings A and B) connected by a three-carbon chain that is typically cyclized into a central pyran ring (ring C). Variations in the oxidation state of ring C, the substitution pattern on rings A and B, and the degree of hydroxylation give rise to the major flavonoid subclasses.

Flavonols, which include quercetin, kaempferol, myricetin, and fisetin, are the most widely distributed flavonoids in the plant kingdom, characterized by a 3-hydroxyflavone backbone with a double bond between C2 and C3. Quercetin in particular has been the subject of extensive pharmacological investigation owing to its potent antioxidant and anti-inflammatory activities. Flavones, such as apigenin and luteolin, share a similar unsaturated C2–C3 bond but lack the 3-hydroxyl group; they are abundant in celery, parsley, and chamomile. Flavanones, represented by naringenin and hesperidin, are characteristically concentrated in citrus fruits.^[7] Isoflavones, notably genistein and daidzein, are predominantly found in leguminous plants, particularly soybean (*Glycine max*), and are distinguished by the migration of the B ring from the C2 to the C3 position of the chromane skeleton; their structural similarity to oestradiol underpins their classification as phytoestrogens and their investigated role in hormone-dependent pathologies. Anthocyanins are water-soluble glycosides of

anthocyanidin aglycones such as cyanidin, delphinidin, and pelargonidin; their colour is pH-dependent, ranging from red in acidic environments to blue in alkaline conditions, and they are most abundant in berries, grapes, red cabbage, and purple sweet potato⁵.

2.2 Non-Flavonoid Polyphenols

Phenolic acids represent the most numerically abundant non-flavonoid polyphenols in the human diet and are subdivided into two principal classes: hydroxycinnamic acids (e.g., caffeic acid, ferulic acid, chlorogenic acid) and hydroxybenzoic acids (e.g., gallic acid, protocatechuic acid, syringic acid). Chlorogenic acid, formed by the esterification of caffeic acid with quinic acid, is among the most abundant polyphenols in the human diet, present at high concentrations in coffee, where it contributes both to flavour complexity and to antioxidant capacity.^[9] Ferulic acid, esterified to arabinoxylans in the cell walls of cereal grains, constitutes a significant antioxidant in whole-grain diets and has additionally demonstrated photoprotective activity in skin cells.

Stilbenes are a smaller but pharmacologically significant non-flavonoid class, characterized by a trans- or cis-1,2-diphenylethylene backbone. Resveratrol (3,5,4'-trihydroxystilbene), found principally in grape skin, red wine, and peanuts, has attracted exceptional research attention since the identification of its ability to activate the longevity-associated deacetylase SIRT1 and to inhibit the NF-κB inflammatory pathway. Pterostilbene, a dimethylated analogue of resveratrol with superior oral bioavailability, has shown promise in preclinical models of metabolic syndrome and cognitive ageing. Lignans are diphenolic compounds formed through the oxidative coupling of two cinnamic acid residues and are abundant in flaxseed, sesame, and whole-grain cereals. Secoisolariciresinol diglucoside (SDG) and matairesinol from flaxseed are converted by colonic bacteria to the mammalian lignans enterodiol and enterolactone, which exhibit weak oestrogenic activity and have been associated with reduced breast cancer risk in prospective cohort studies⁶.

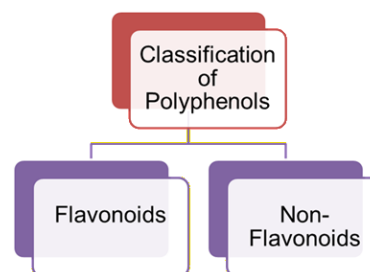


Figure 1: Classification of polyphenol..

3. BIOSYNTHESIS OF POLYPHENOLS

3.1 The Shikimic Acid Pathway

The biosynthesis of the majority of plant polyphenols originates in the shikimic acid pathway, a seven-step metabolic route confined to plants, fungi, and

microorganisms that converts simple carbohydrate precursors — specifically phosphoenolpyruvate (PEP) from glycolysis and erythrose-4-phosphate (E4P) from the pentose phosphate pathway — into chorismate, the last common intermediate of aromatic amino acid biosynthesis. The pathway proceeds through 3-deoxy-D-arabino-heptulosonate-7-phosphate (DAHP), dehydroquinate, shikimate, shikimate-3-phosphate, and 5-enolpyruvylshikimate-3-phosphate (EPSP) before yielding chorismate. The enzyme EPSP synthase, notably the molecular target of the herbicide glyphosate, catalyses the penultimate step of this pathway. Chorismate is subsequently converted to prephenate and thence to the aromatic amino acids phenylalanine and tyrosine, which serve as the direct precursors for polyphenol biosynthesis in the phenylpropanoid pathway ⁷.

3.2 The Phenylpropanoid Pathway

The phenylpropanoid pathway is initiated by the action of phenylalanine ammonia-lyase (PAL), which catalyses the non-oxidative deamination of phenylalanine to trans-cinnamic acid with the release of ammonia. PAL is considered a pivotal regulatory enzyme of polyphenol biosynthesis and is transcriptionally induced by diverse environmental stimuli, including UV radiation, wounding, pathogen attack, and nutrient deficiency. Trans-cinnamic acid is subsequently hydroxylated at the para position by the cytochrome P450 enzyme cinnamate-4-hydroxylase (C4H) to yield p-coumaric acid, which is then activated to p-coumaroyl-CoA by 4-coumarate: CoA ligase (4CL). p-Coumaroyl-CoA serves as a key branch-point metabolite: condensation with three molecules of malonyl-CoA by chalcone synthase (CHS) initiates the flavonoid biosynthetic branch, while successive hydroxylation and reduction reactions generate the phenylpropanoid esters, lignin monomers, and stilbene precursors. Within the flavonoid branch, chalcone synthase catalyses a polyketide condensation reaction to produce naringenin chalcone, which is subsequently isomerised to naringenin (a flavanone) by chalcone isomerase (CHI).

Naringenin then serves as the precursor for the diverse flavonoid subclasses: flavanone 3-hydroxylase (F3H) converts naringenin to dihydrokaempferol, which is further hydroxylated by flavonoid 3'-hydroxylase (F3'H) and flavonoid 3',5'-hydroxylase (F3',5'H) to yield dihydroquercetin and dihydromyricetin, respectively. Flavonol synthase (FLS) oxidises these dihydroflavonols to the corresponding flavonols, while dihydroflavonol 4-reductase (DFR) and leucoanthocyanidin dioxygenase (LDOX/ANS) direct carbon flux toward the anthocyanin branch.

The regulatory control of this pathway is largely mediated by MYB transcription factors acting in concert with basic helix-loop-helix (bHLH) proteins and WD40-repeat proteins, forming the MBW ternary complex that activates structural gene expression in a tissue- and stimulus-specific manner ⁸.

4. SOURCES OF POLYPHENOLS

4.1 Natural Dietary Sources

Polyphenols are ubiquitously distributed across plant food sources, and estimates of mean daily intake in Western diets range from approximately 1,000 to 2,000 mg per day, making them quantitatively among the most abundant bioactive compounds in the human diet. Fruits contribute the most diverse range of polyphenol classes: pomegranate (*Punica granatum*) is one of the richest sources of ellagitannins and punicalagins; berries, including blueberries, blackcurrants, and raspberries, provide high concentrations of anthocyanins and ellagic acid; apples contain quercetin glycosides and chlorogenic acid; and grapes are the primary dietary source of resveratrol and procyanidins. Among vegetables, onions are exceptional sources of quercetin, artichokes are rich in cynarin and chlorogenic acid, and members of the Brassica family provide kaempferol and various hydroxycinnamic acid derivatives. Legumes, particularly soybeans, chickpeas, and lentils, are the principal dietary sources of isoflavones in populations consuming traditional Asian diets.

Beverages constitute a quantitatively significant contribution to total polyphenol intake in many populations. Coffee is the single largest contributor to dietary polyphenol intake in European and North American populations, primarily through its high content of chlorogenic acids (200–550 mg per cup in filtered coffee). Green tea is an exceptionally rich source of catechins, particularly epigallocatechin gallate (EGCG), which has been extensively investigated for its anticancer and metabolic properties. Red wine provides resveratrol, quercetin, and myricetin from grape skin, as well as polymeric procyanidins from seeds; the antioxidant capacity of red wine substantially exceeds that of white wine owing to the greater polyphenol extraction during skin contact fermentation. Dark chocolate (≥70% cocoa) is among the most polyphenol-dense foods available, containing high concentrations of procyanidins, epicatechin, and catechin, and human intervention studies have documented acute improvements in endothelial function and blood pressure following consumption ⁹.

5. CHEMICAL PROPERTIES AND STRUCTURE–ACTIVITY RELATIONSHIP

5.1 Functional Groups and Physicochemical Properties

The physicochemical properties of polyphenols are largely determined by the nature, number, and spatial distribution of their hydroxyl groups, as well as by the presence of conjugated double bonds, carbonyl functionalities, and glycosidic substituents. The phenolic hydroxyl groups are weakly acidic, with pKa values typically ranging from 8 to 10, and confer moderate water solubility; however, glycosylation substantially increases hydrophilicity, whereas methylation and prenylation reduce polarity and improve membrane permeability. The aromatic ring systems contribute to the planar molecular geometry that facilitates intercalation with nucleic acids and interaction



with protein binding pockets, a property of relevance to both the antimicrobial and anticancer activities of this compound class ¹⁰.

5.2 Antioxidant Mechanisms and Structure-Activity Relationships

The antioxidant activity of polyphenols is primarily attributable to their capacity to donate hydrogen atoms or electrons to free radicals, thereby interrupting oxidative chain reactions. This activity is mechanistically classified into two principal modes: hydrogen atom transfer (HAT), in which a phenolic O–H bond is homolytically cleaved to yield a stable phenoxyl radical, and single electron transfer (SET), in which an electron is donated to a radical acceptor with concomitant formation of a phenolate cation. The bond dissociation enthalpy (BDE) of the phenolic O–H bond is the primary thermodynamic determinant of HAT-based antioxidant activity, and lower BDE values, associated with greater radical stabilisation through resonance delocalisation and intramolecular hydrogen bonding, correspond to superior antioxidant potency.

Structure-activity relationship studies have established several key structural determinants of antioxidant efficacy. The presence of a catechol moiety (ortho-dihydroxy substitution) on ring B, as found in quercetin, catechin, and caffeic acid, is strongly associated with high antioxidant activity owing to the ability of the resulting semiquinone radical to be stabilised by electron delocalisation and intramolecular hydrogen bonding. The 2,3-double bond in conjugation with a 4-oxo group in flavones and flavonols further enhances radical delocalisation across the entire molecular framework. Additionally, hydroxyl groups at C3 and C5, through intramolecular chelation with divalent metal ions such as Fe²⁺ and Cu²⁺, enable polyphenols to inhibit metal-catalysed Fenton-type reactions that generate the highly reactive hydroxyl radical (•OH) from hydrogen peroxide. This metal chelation capacity is considered an important secondary mechanism of antioxidant protection, particularly relevant to the prevention of lipid oxidation in biological membranes and food matrices ¹¹.

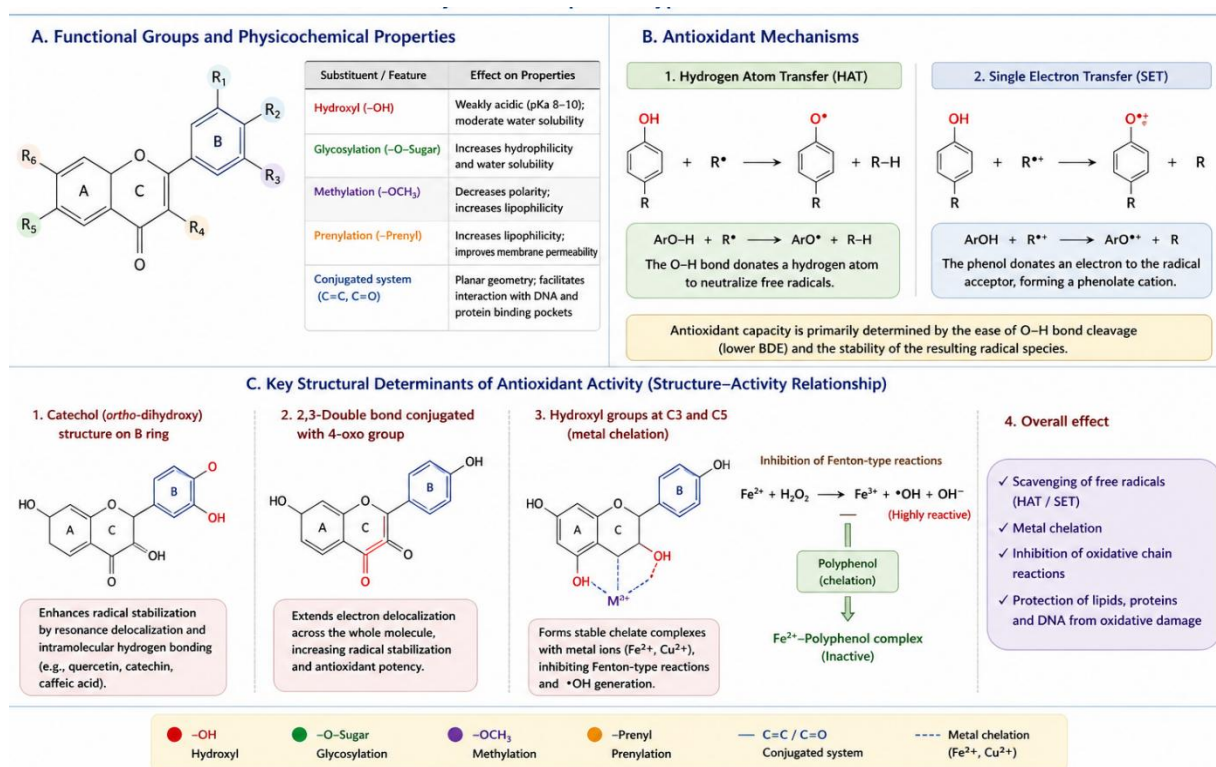


Figure 2: Chemical Properties and Structure–Activity Relationship.

6. BIOLOGICAL ACTIVITIES

6.1 Antioxidant Activity

The antioxidant properties of polyphenols have been documented across a wide range of in vitro and in vivo experimental systems. In cultured cell models, polyphenol treatment consistently reduces intracellular reactive oxygen species (ROS) levels, as quantified by fluorescent probes such as 2',7'-dichlorodihydrofluorescein diacetate (DCFH-DA), and activates the Nrf2-Keap1 transcriptional pathway, which governs the expression of cytoprotective and

antioxidant genes including heme oxygenase-1 (HO-1), NAD(P)H quinone oxidoreductase 1 (NQO1), and the glutamate-cysteine ligase subunits (GCLC and GCLM). Human intervention studies using biomarkers of oxidative stress, such as plasma F2-isoprostanes and 8-hydroxy-2'-deoxyguanosine (8-OHdG), have provided in vivo evidence of antioxidant efficacy for quercetin, EGCG, and anthocyanin-rich extracts, though the magnitude of effects varies considerably with dose, matrix, and individual metabolic factor ¹².



6.2 Anti-inflammatory Effects

Polyphenols interfere with inflammatory signaling at multiple regulatory nodes, with the NF- κ B and MAPK pathways representing the most frequently reported molecular targets. NF- κ B, a master transcriptional regulator of inflammatory gene expression, is maintained in an inactive cytoplasmic state by the inhibitory protein I κ B α ; upon phosphorylation and proteasomal degradation of I κ B α , NF- κ B translocates to the nucleus and activates the transcription of pro-inflammatory mediators including TNF- α , IL-1 β , IL-6, cyclooxygenase-2 (COX-2), and inducible nitric oxide synthase (iNOS). Curcumin, resveratrol, quercetin, and EGCG have each been demonstrated to suppress NF- κ B activation through distinct upstream mechanisms: curcumin inhibits the I κ B kinase (IKK) complex, resveratrol suppresses I κ B α phosphorylation, and quercetin inhibits Src kinase-mediated activation of the NF- κ B pathway. COX-2 inhibition by polyphenols, which reduces prostaglandin E2 (PGE2) synthesis, is relevant to both their anti-inflammatory and putative anticancer activities, as prostaglandins are established promoters of tumour angiogenesis and immune evasion¹³.

6.3 Anticancer Properties

The anticancer activities of polyphenols encompass a range of mechanisms operating at distinct stages of carcinogenesis. EGCG from green tea has been demonstrated to inhibit multiple receptor tyrosine kinases, including the epidermal growth factor receptor (EGFR) and vascular endothelial growth factor receptor (VEGFR), thereby suppressing downstream PI3K/Akt/mTOR and Ras/Raf/MEK/ERK signalling cascades that drive tumour cell proliferation and survival. Resveratrol has been shown to activate the pro-apoptotic protein Bax, downregulate the anti-apoptotic proteins Bcl-2 and survivin, and induce G1-S and G2-M cell cycle arrest through upregulation of p21 and p53, collectively promoting apoptosis in diverse human cancer cell lines. Quercetin has demonstrated synergistic anticancer activity with conventional chemotherapeutic agents, including cisplatin and doxorubicin, in cell and animal models, suggesting potential clinical utility as a chemosensitizer. Pomegranate-derived punicalagins and ellagic acid have been shown to inhibit NF- κ B, suppress VEGF-mediated angiogenesis, and reduce tumour cell invasion in preclinical models¹⁴.

6.4 Antimicrobial Activity

The antimicrobial properties of polyphenols were recognized empirically long before their chemical basis was understood, as the use of tannin-rich plant preparations for wound treatment and food preservation predates modern microbiology. Mechanistically, polyphenols exert antimicrobial effects through disruption of the microbial cell membrane and cell wall integrity, inhibition of extracellular and intracellular microbial enzymes, chelation of essential metal cofactors, and interference with nucleic acid synthesis. Tannins and condensed procyanidins exhibit particularly potent antibacterial activity against both Gram-

positive organisms (e.g., *Staphylococcus aureus*, *Streptococcus mutans*) and Gram-negative bacteria (e.g., *Escherichia coli*, *Pseudomonas aeruginosa*), with minimum inhibitory concentrations (MICs) in the range of 0.1–1.0 mg/mL reported for high-purity extracts. Importantly, EGCG has demonstrated activity against methicillin-resistant *Staphylococcus aureus* (MRSA) strains, suggesting potential utility in the context of antimicrobial resistance¹⁵.

6.5 Cardioprotective and Neuroprotective Roles

Cardiovascular protection by polyphenols operates through multiple interdependent mechanisms. Improvement of endothelial function is among the most consistently reported cardiovascular effects, mediated primarily through polyphenol-stimulated activation of endothelial nitric oxide synthase (eNOS) and consequent elevation of vascular nitric oxide (NO) bioavailability, which promotes vasodilation, reduces platelet aggregation, and inhibits monocyte adhesion. Flavanols from cocoa and green tea have been shown in randomized controlled trials to significantly reduce brachial artery blood pressure and improve flow-mediated dilation. Polyphenols also reduce low-density lipoprotein (LDL) oxidation, a critical initiating event in atherosclerotic plaque formation, through their antioxidant activity and their ability to chelate the copper ions that catalyse LDL oxidation in the sub-endothelial space.

With respect to neuroprotection, polyphenols have attracted growing interest as candidate agents for the prevention and management of Alzheimer's disease (AD) and Parkinson's disease (PD). In cellular and rodent models of AD, EGCG has been shown to inhibit the aggregation of amyloid- β (A β) peptides into toxic oligomers and fibrils, to activate the α -secretase pathway, and to reduce tau hyperphosphorylation through inhibition of glycogen synthase kinase-3 β (GSK-3 β). Resveratrol activates SIRT1-mediated deacetylation of PGC-1 α , enhancing mitochondrial biogenesis and reducing the mitochondrial oxidative stress that contributes to dopaminergic neuron loss in PD. Quercetin and fisetin have demonstrated neurogenesis-promoting effects in the hippocampus in animal models, a finding of potential relevance to cognitive ageing and depression¹⁶.

7. MECHANISM OF ACTION

7.1 ROS Scavenging and Redox Signalling

Beyond their direct free radical scavenging activity, polyphenols modulate cellular redox homeostasis through the regulation of antioxidant enzyme expression via the Nrf2-Keap1 axis. Under basal conditions, the transcription factor Nrf2 is sequestered in the cytoplasm through its interaction with the adaptor protein Keap1, which facilitates its ubiquitination and proteasomal degradation. Electrophilic or oxidative modification of critical cysteine residues (notably Cys151, Cys273, and Cys288) within Keap1 disrupts this interaction, allowing Nrf2 to translocate to the nucleus, where it binds the antioxidant response element (ARE) in the promoters of cytoprotective genes. Several polyphenols, including sulforaphane (from Brassica



vegetables), curcumin, quercetin, and caffeic acid phenethyl ester (CAPE), have been demonstrated to activate Nrf2 through either direct Keap1 modification or through indirect activation of upstream kinases, including PI3K/Akt and MAPK pathways¹⁷.

7.2 Interaction with Enzymes and Transcription Factors

Polyphenols exhibit broad enzyme-inhibitory activity with specificity profiles that reflect their structural features, particularly the planarity of the aromatic ring system and the spatial arrangement of hydroxyl groups. EGCG inhibits the 67-kDa laminin receptor (67LR), through which it triggers apoptosis signaling in cancer cells independently of receptor tyrosine kinase pathways. Quercetin functions as an ATP-competitive inhibitor of casein kinase 2 (CK2), a pleiotropic kinase implicated in cell survival, DNA repair, and circadian rhythm regulation. At the transcriptional level, polyphenols modulate the activity of epigenetic regulators: curcumin inhibits histone acetyltransferases (HATs), including p300/CBP, EGCG inhibits DNA methyltransferase 1 (DNMT1), and resveratrol activates the class III histone deacetylase SIRT1, collectively producing alterations in chromatin organization that influence the expression of genes involved in differentiation, senescence, and apoptosis¹⁸.

8. APPLICATIONS IN BIOTECHNOLOGY

8.1 Pharmaceutical Applications

The pharmaceutical sector has pursued polyphenol-based drug development along two complementary strategies: the direct use of purified polyphenolic compounds as active pharmaceutical ingredients, and the exploitation of polyphenol scaffolds as lead compounds for medicinal chemistry optimisation. Resveratrol has been evaluated in clinical trials for conditions ranging from type 2 diabetes and metabolic syndrome to Alzheimer's disease and multiple myeloma, with a mixed efficacy profile attributed largely to its rapid metabolic clearance and poor systemic bioavailability. Several pharmaceutical companies have consequently developed prodrugs and structural analogues of resveratrol, including pterostilbene and SRT501 (a micronised resveratrol formulation), with improved pharmacokinetic profiles. Quercetin has been formulated as a pharmaceutical-grade preparation in combination with bromelain for the management of prostatitis and as an adjunctive agent in chemotherapy protocols. Polyphenol-derived scaffolds have additionally contributed to the development of flavone acetic acid analogues as antitumour vascular-disrupting agents¹⁹.

8.2 Nutraceuticals and Functional Foods

The nutraceutical industry represents the most commercially advanced application of polyphenol science. Standardised green tea extracts providing defined EGCG

content, grape seed proanthocyanidin extracts (GSPE), and pomegranate polyphenol concentrates are among the most commercially significant polyphenol nutraceuticals. The development of functional foods — conventional food products to which specific bioactive compounds have been added to confer health benefits beyond basic nutrition — has incorporated polyphenol enrichment strategies including the fortification of dairy products with green tea catechins, the addition of resveratrol to fruit beverages, and the encapsulation of curcumin in emulsified food matrices to improve its stability and bioaccessibility during gastrointestinal processing²⁰.

8.3 Food Preservation

The antimicrobial and antioxidant properties of polyphenols have been exploited in food science for the extension of product shelf-life and the reduction of synthetic preservative usage. Rosemary extract, standardized to carnosic acid and rosmarinic acid content, is widely used as a natural antioxidant in meat products, fish oils, and snack foods. Grape pomace extract, a cost-effective byproduct of wine production with high procyanidin content, has been demonstrated to retard lipid oxidation in comminuted meat systems as effectively as synthetic antioxidants such as butylated hydroxytoluene (BHT) at equivalent concentrations. The incorporation of polyphenol-rich plant extracts into active food packaging materials, through their inclusion in biopolymer films based on chitosan, zein, or cellulose derivatives, represents an emerging strategy in which the polyphenol functions simultaneously as a bioactive component and as a structural crosslinker that enhances the mechanical and barrier properties of the packaging film²¹.

8.4 Cosmetic Industry

The cosmetic and dermatological industry has incorporated polyphenols extensively into topical formulations targeting photoprotection, anti-ageing, hyperpigmentation, and inflammatory skin conditions. Green tea extract is among the most widely used polyphenol ingredients in cosmeceuticals, exploited for its capacity to absorb UV radiation, scavenge UV-induced ROS, suppress UV-activated NF-κB-mediated inflammatory responses in keratinocytes, and inhibit matrix metalloproteinases (MMPs) that degrade structural collagen and elastin. Resveratrol has been incorporated into anti-ageing serums and creams based on its ability to activate SIRT1 in dermal fibroblasts, promoting mitochondrial biogenesis and enhancing collagen synthesis while reducing the UV-induced expression of collagenase (MMP-1). Polyphenols from pomegranate, particularly the ellagitannin metabolite urolithin A produced by gut bacteria from ellagic acid, have demonstrated potent inhibitory activity against tyrosinase, the rate-limiting enzyme of melanin biosynthesis, positioning pomegranate-derived preparations as functional skin-lightening agents²².



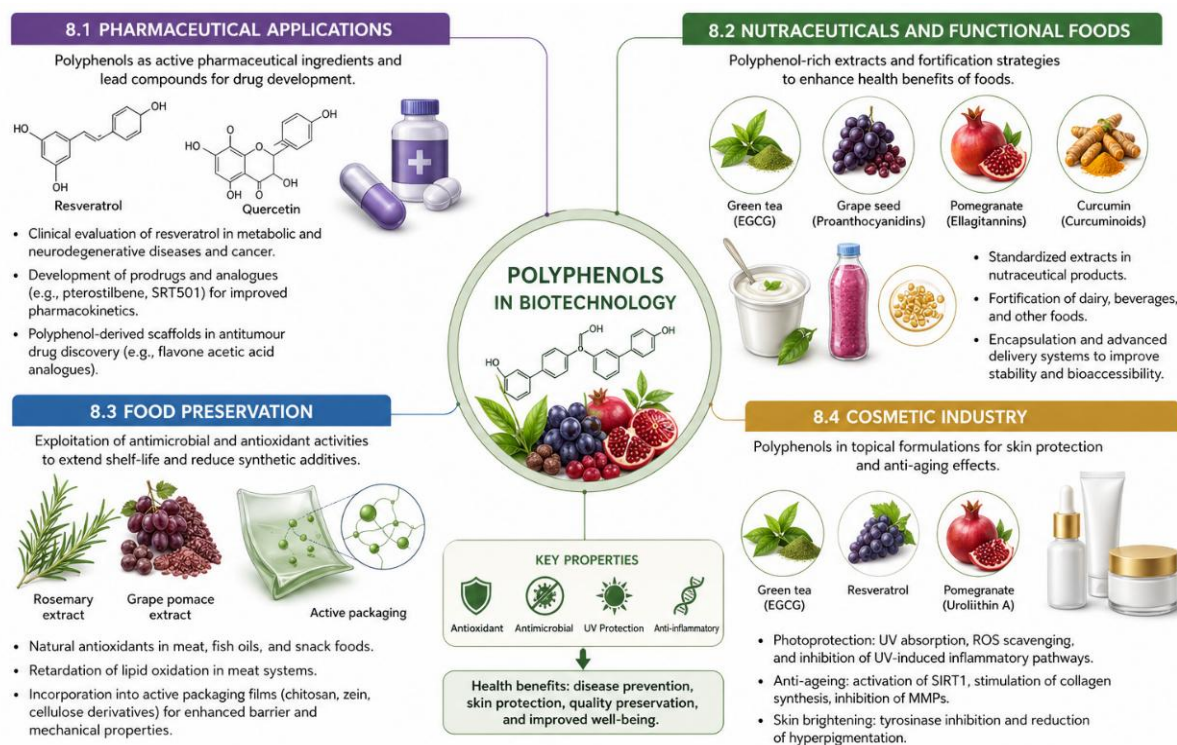


Figure 3: Polyphenol: multifunctional bioactives driving innovation in pharmaceuticals, nutrition, food technology, and cosmetics.

9. EXTRACTION AND ANALYSIS TECHNIQUES

9.1 Conventional Extraction Methods

The isolation of polyphenols from plant matrices has been accomplished conventionally through solid-liquid extraction techniques that exploit the differential solubility of polyphenol classes in solvents of varying polarity. Maceration, the simplest approach, involves soaking comminuted plant material in an aqueous-organic solvent (typically 50–80% methanol, ethanol, or acetone in water) at ambient temperature for a defined period, with periodic agitation to promote mass transfer. The Soxhlet extraction apparatus facilitates continuous solvent cycling through the sample matrix in a heated thimble, enabling exhaustive extraction with relatively small solvent volumes; however, the thermal conditions may promote degradation of heat-labile polyphenols such as anthocyanins. Aqueous extraction at elevated temperatures (decoction) is particularly effective for hydrophilic polyphenols, including phenolic acids and flavonoid glycosides. The selection of the extraction solvent is critical: pure water favours the recovery of glycosylated and highly polar phenolics, methanol provides broad-spectrum recovery with good stability, and acetone-water mixtures are preferred for procyanidins and condensed tannins²³.

9.2 Advanced Analytical Techniques

High-performance liquid chromatography (HPLC) coupled with ultraviolet-diode array detection (DAD) has been the workhorse analytical platform for polyphenol quantification and profiling over the past three decades. Reversed-phase

HPLC on C₁₈ stationary phases, using gradient elution with acidified aqueous-acetonitrile mobile phases, achieves effective separation of the major flavonoid and phenolic acid classes within analytical run times of 30–60 minutes. The hyphenation of HPLC with electrospray ionisation mass spectrometry (LC-ESI-MS/MS) enables simultaneous identification and quantification of polyphenols at sub-nanomolar concentrations based on their characteristic fragmentation patterns, eliminating the need for authentic reference standards for structural assignment. Liquid chromatography coupled with high-resolution mass spectrometry (LC-HRMS), using Orbitrap or quadrupole time-of-flight (Q-TOF) instruments, enables untargeted metabolomics approaches in which hundreds of individual polyphenols can be detected and tentatively identified in a single analytical run. Nuclear magnetic resonance (NMR) spectroscopy remains indispensable for the unambiguous structural characterisation of novel polyphenol isolates, providing information on substitution patterns, stereochemistry at chiral centres, and glycosidic linkages that cannot be inferred from mass spectrometric data alone²⁴.

10. BIOAVAILABILITY AND METABOLISM

10.1 Absorption and Systemic Metabolism

The bioavailability of dietary polyphenols is characteristically low and highly variable, with plasma concentrations typically in the nanomolar to low micromolar range following consumption of polyphenol-rich foods or supplements. Polyphenol aglycones of moderate polarity, such as quercetin and resveratrol, can be

absorbed directly in the small intestine by passive diffusion across the enterocyte brush border membrane, assisted in some cases by active transport via sodium-dependent glucose transporters (SGLTs) for certain glycoside forms. The majority of polyphenol glycosides, however, cannot traverse the intestinal epithelium without prior hydrolysis of the glycosidic bond, a transformation effected by mucosal lactase-phlorizin hydrolase (LPH) for glucose conjugates or by cytosolic β -glucosidase (CBG) following cellular uptake.

Within enterocytes and hepatocytes, polyphenol aglycones undergo Phase I oxidative metabolism by cytochrome P450 enzymes (predominantly CYP1A1, CYP1A2, and CYP3A4) and Phase II conjugation reactions including glucuronidation by UDP-glucuronosyltransferases (UGTs), sulfation by sulfotransferases (SULTs), and methylation by catechol-O-methyltransferase (COMT). The resulting glucuronide, sulfate, and methyl conjugates are the predominant circulating forms of dietary polyphenols in human plasma and represent the physiologically relevant species at target tissues. Efflux transporters of the ABC family, particularly MRP2 (ABCC2), actively export polyphenol conjugates back into the intestinal lumen or into bile, contributing to the enterohepatic recirculation that prolongs polyphenol residence in the body but also limits net absorption²⁵.

10.2 Role of the Gut Microbiome

Polyphenols that escape absorption in the small intestine, estimated to constitute 90–95% of ingested flavonoids and condensed tannins, pass into the large intestine, where they encounter the diverse and metabolically active gut microbiota. Microbial catabolism of polyphenols generates a range of bioactive low-molecular-weight phenolic acids and lactones that are more readily absorbed from the colon than their parent structures. The most extensively characterized examples of microbiome-dependent polyphenol bioactivation include the conversion of ellagitannins and ellagic acid to urolithins (urolithin A, B, and C) by *Gordonibacter* and *Ellagibacter* species, the hydrolysis of daidzein to equol by specific *Lactobacillus* and *Clostridia* species, and the cleavage of procyanidins and flavonol glycosides to simple phenolic acids by *Bacteroides*, *Bifidobacterium*, and *Clostridium* species. The capacity for urolithin production and equol conversion exhibits substantial inter-individual variation linked to gut microbiota composition, providing a biological explanation for the highly variable response to polyphenol supplementation observed in clinical trials²⁶.

11. LIMITATIONS AND CHALLENGES

Despite the compelling body of preclinical evidence supporting the pharmacological activities of polyphenols, their translation into validated clinical applications has been impeded by a constellation of technical, biological, and methodological challenges. The low systemic bioavailability fundamentally constrains the tissue concentrations achievable through oral supplementation, and it remains uncertain whether the plasma concentrations attained following physiological dietary intake are sufficient to exert

the biological effects observed in cell culture systems where polyphenols are applied at concentrations of 10–100 $\mu\text{mol/L}$. Polyphenols are chemically labile under conditions of elevated temperature, alkaline pH, and oxidative processing, leading to significant degradation during food preparation, gastrointestinal digestion, and product storage; anthocyanins are particularly susceptible to hydrolysis and oxidative degradation, and curcumin undergoes rapid aqueous decomposition at neutral-alkaline pH to yield ferulic acid and other breakdown products. The design and interpretation of human intervention studies are complicated by the difficulty of accurately quantifying polyphenol intake using dietary assessment instruments, by the extensive inter-individual variability in polyphenol absorption and metabolism, and by the absence of well-validated biomarkers of polyphenol status and biological effect. At very high supplementation doses, quercetin inhibits CYP3A4 and P-glycoprotein, potentially altering the pharmacokinetics of co-administered drugs, and high-dose resveratrol has been reported to paradoxically impair exercise-induced cardiovascular adaptations in healthy subjects²⁷.

12. FUTURE PERSPECTIVES

The most significant emerging direction in polyphenol biotechnology is the development of nanoformulation strategies designed to overcome the bioavailability limitations inherent to this compound class. Lipid-based nanocarriers, including nanoemulsions, solid lipid nanoparticles (SLNs), and nanostructured lipid carriers (NLCs), have been shown to improve the oral bioavailability of curcumin by 10–20-fold relative to the free compound, by enhancing solubilization in the intestinal lumen, facilitating lymphatic absorption, and reducing first-pass hepatic metabolism. Polymeric nanoparticles based on poly (lactic-co-glycolic acid) (PLGA) and chitosan provide a further platform for controlled-release delivery of polyphenols at specific sites in the gastrointestinal tract, enabling targeted delivery to the colonic mucosa for local anti-inflammatory and chemoprevention applications. Cyclodextrin inclusion complexes and phospholipid complexes (phytosomes) represent simpler excipient-based approaches that have achieved regulatory approval and commercial implementation for curcumin and silymarin formulations.

The integration of polyphenol research with precision medicine and systems biology represents a transformative future direction. The recognition that individual responses to polyphenol supplementation are partly determined by genetic polymorphisms in metabolising enzymes (e.g., COMT Val158Met, UGT1A polymorphisms) and by gut microbiota composition has stimulated the concept of precision nutrition. The application of multi-omics approaches — integrating genomics, transcriptomics, metabolomics, and microbiomics — to human polyphenol intervention studies is beginning to generate the mechanistic data necessary to advance this concept toward clinical implementation. Metabolic engineering of plant



biosynthetic pathways, enabled by CRISPR-Cas9 genome editing and synthetic biology tools, offers the prospect of producing polyphenols of pharmaceutical interest in microbial fermentation systems at scale and cost that are unachievable through plant extraction²⁸.

CONCLUSION

This review has presented a comprehensive account of polyphenol science spanning fundamental chemistry, biosynthetic biology, nutritional science, pharmacology, and applied biotechnology. Polyphenols are chemically distinguished by their structurally diverse hydroxylated aromatic frameworks, which underpin a remarkable range of biological activities mediated through antioxidant, anti-inflammatory, antimicrobial, anticancer, cardioprotective, and neuroprotective mechanisms. Their biosynthesis through the shikimic acid and phenylpropanoid pathways is tightly regulated in response to environmental and developmental signals, and the enzymatic machinery responsible for the structural diversification of the polyphenol scaffold has been substantially elucidated at the molecular level. The dietary abundance of polyphenols across fruits, vegetables, beverages, and grains positions them as quantitatively significant components of the human food environment, and epidemiological evidence consistently associates high polyphenol intake with favourable health outcomes across multiple chronic disease endpoints.

The principal challenge confronting the clinical translation of polyphenol science remains the gap between the concentrations demonstrated to be biologically active in cell culture and the systemic concentrations achievable through dietary intake or oral supplementation. Advances in nanoencapsulation technology, structural analogue development, and microbiome-informed delivery strategies are progressively narrowing this gap. The emergence of precision nutrition as a clinical discipline, supported by multi-omics characterization of individual metabolic phenotypes, holds particular promise for optimising the deployment of polyphenol-based interventions in preventive medicine and personalised therapeutics. For researchers operating at the interface of plant biochemistry, pharmacognosy, and biomedical engineering, polyphenols represent a scientifically rich and practically consequential domain that will continue to generate significant advances in both fundamental understanding and applied biotechnology for decades to come.

JOINT FIRST AUTHORSHIP

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REFERENCES

1. Quideau S, Deffieux D, Douat-Casassus C, Pouységu L. Plant polyphenols: chemical properties, biological activities, and synthesis. *Angew Chem Int Ed Engl*. 2011;50(3):586-621.
2. Manach C, Scalbert A, Morand C, Rémésy C, Jiménez L. Polyphenols: food sources and bioavailability. *Am J Clin Nutr*. 2004;79(5):727-747.
3. Scalbert A, Johnson IT, Saltmarsh M. Polyphenols: antioxidants and beyond. *Am J Clin Nutr*. 2005;81(1 Suppl):215S-217S.
4. Cory H, Passarelli S, Szeto J, Tamez M, Mattei J. The role of polyphenols in human health and food systems: a mini-review. *Front Nutr*. 2018;5:87.
5. Tsao R. Chemistry and biochemistry of dietary polyphenols. *Nutrients*. 2010;2(12):1231-1246.
6. Adlercreutz H. Lignans and human health. *Crit Rev Clin Lab Sci*. 2007;44(5-6):483-525.
7. Vogt T. Phenylpropanoid biosynthesis. *Mol Plant*. 2010;3(1):2-20.
8. Koes R, Verweij W, Quattrocchio F. Flavonoids: a colorful model for the regulation and evolution of biochemical pathways. *Trends Plant Sci*. 2005;10(5):236-242.
9. Pandey KB, Rizvi SI. Plant polyphenols as dietary antioxidants in human health and disease. *Oxid Med Cell Longev*. 2009;2(5):270-278.
10. Balasundram N, Sundram K, Samman S. Phenolic compounds in plants and agri-industrial by-products: antioxidant activity, occurrence, and potential uses. *Food Chem*. 2006;99(1):191-203.
11. Rice-Evans CA, Miller NJ, Paganga G. Structure-antioxidant activity relationships of flavonoids and phenolic acids. *Free Radic Biol Med*. 1996;20(7):933-956.
12. Williamson G. The role of polyphenols in modern nutrition. *Nutr Bull*. 2017;42(3):226-235.
13. Pan MH, Lai CS, Ho CT. Anti-inflammatory activity of natural dietary flavonoids. *Food Funct*. 2010;1(1):15-31.
14. Ramos S. Effects of dietary flavonoids on apoptotic pathways related to cancer chemoprevention. *J Nutr Biochem*. 2007;18(7):427-442.
15. Daglia M. Polyphenols as antimicrobial agents. *Curr Opin Biotechnol*. 2012;23(2):174-181.
16. Vauzour D. Dietary polyphenols as modulators of brain functions: biological actions and molecular mechanisms. *Nutr Res Rev*. 2012;25(2):238-250.
17. Surh YJ. Cancer chemoprevention with dietary phytochemicals. *Nat Rev Cancer*. 2003;3(10):768-780.
18. Link A, Balaguer F, Goel A. Cancer chemoprevention by dietary polyphenols: promising role for epigenetics. *Biochem Pharmacol*. 2010;80(12):1771-1792.
19. Baur JA, Sinclair DA. Therapeutic potential of resveratrol: the in vivo evidence. *Nat Rev Drug Discov*. 2006;5(6):493-506.
20. Granato D, Shahidi F, Wrolstad R, et al. Antioxidant activity, total phenolics and flavonoids contents: should we ban in vitro screening methods? *Food Chem*. 2018;264:471-475.



21. Shahidi F, Ambigaipalan P. Phenolics and polyphenolics in foods, beverages and spices: antioxidant activity and health effects. *J Funct Foods*. 2015;18:820-897.
22. Nichols JA, Katiyar SK. Skin photoprotection by natural polyphenols: anti-inflammatory, antioxidant and DNA repair mechanisms. *Arch Dermatol Res*. 2010;302(2):71-83.
23. Dai J, Mumper RJ. Plant phenolics: extraction, analysis and their antioxidant and anticancer properties. *Molecules*. 2010;15(10):7313-7352.
24. Ignat I, Volf I, Popa VI. A critical review of methods for characterisation of polyphenolic compounds in fruits and vegetables. *Food Chem*. 2011;126(4):1821-1835.
25. Del Rio D, Rodriguez-Mateos A, Spencer JPE, Tognolini M, Borges G, Crozier A. Dietary polyphenolics in human health: structures, bioavailability, and evidence of protective effects. *Antioxid Redox Signal*. 2013;18(14):1818-1892.
26. Selma MV, Espín JC, Tomás-Barberán FA. Interaction between phenolics and gut microbiota: role in human health. *J Agric Food Chem*. 2009;57(15):6485-6501.
27. D'Archivio M, Filesi C, Vari R, Scazzocchio B, Masella R. Bioavailability of the polyphenols: status and controversies. *Int J Mol Sci*. 2010;11(4):1321-1342.
28. Saini RK, Keum YS. Microbial platforms to produce polyphenols: applications and perspectives. *Trends Biotechnol*. 2019;37(6):605-617.

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