Identification of Potential Impurities of Montelukast Imported to Syria by LC-MS and HPLC-UV

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ABSTRACT

Montelukast is a specific cysteiny1 leukotriene receptor antagonist, which could include in its raw material many impurities that ought to be separated and identified for drug development and quality control. These impurities were identified by two commonly used methods: high-performance liquid chromatography UV and LC-mass spectrometry. The HPLC-UV method employed a phenyl column while the LC-MS method employed a C18 column. HPLC-UV method differentiated montelukast from its cis-isomer impurity and indentified other pharmacopeial impurities, but The LC-MS couldn’t differentiated the cis-isomer impurity. On the other hand LC-MS is sufficient for impurity identification because it provides molecular weight information without the need for time-consuming isolation.

Keywords: Montelukast, Impurities, HPLC, LC-MS.

INTRODUCTION

Montelukast [R-(E):1-][1-[3-[2-(7-chloro-2 quinolinyl) ethenyl]phenyl]-3-[2-(1-hydroxy-1-methyl) phenyl] propyl] thio] methyl cyclopropanecarboxylic acid, monosodium salt (fig 1).1 (MO) is a leukotriene receptor antagonist (LTRA) used for the treatment of asthma and to relieve symptoms of seasonal allergies. MO is usually administered orally. MO is a cysteiny1 leukotriene receptor CysLT1 antagonist; belongs to quinoline series. MO blocks the action of leukotriene D4 on the CysLT1 in the lungs and bronchial tubes by binding to it. This reduces the bronchoconstriction caused by the leukotriene, and results in less inflammation.2,3

So, it is a successful therapeutic agent for the treatment of bronchial asthma and to relieve symptoms of seasonal allergies.2 MO is more than 99% bound to plasma proteins with bioavailability of 63% to 73% and half life of 2.7–5.5 h and extensively metabolized by liver and excreted by biliary.4,5 MO is the market leader in this pharmacologic class.6

MO could include in its raw material many impurities that ought to be separated and identified for drug development and quality control.7 These impurities could result from chemical instability of MO as well as the instability of the raw materials used for its chemical synthesis or non selectivity of chemical reaction or may be represented by residues of the raw material used especially solvents.8

And it is known to be prone to several types of degradation such as oxidation of the mercapto group to the sulfoxide, Isomerisation at the location of the double bond from trans to cis by the effect of light., dehydration at the location of tert. Alcohol, producing the corresponding olefin.9

Besides the impurities that come from the decomposition of the target substance, the API contains also specific impurities that can also be brought into the drug substance through the production process. Impurities of this type differ from the degradation impurities mainly by the fact that their content in the target substance does not grow any further.8

The presence of impurities or its related compounds in a drug substance can have a significant impact on the quality and safety of the drug product.3

So ICH recommended to isolate and characterize degradation products that generated during the storage or usability period of API.10

Few analytical methods were reported earlier for the identification of MO's impurities, and most of them focus on the process-related impurities and the kinetics of montelukast' photodegradation.3,10,11

So The aim of this study was to detect and identify impurities of MO imported to Syria using two different detectors such as ultra violet UV detector and mass spectroscopy MS detector.

![Figure 1: Chemical structure of Montelukast sodium](fig.jpg)

**MATERIALS AND METHODS**

**Samples and Chemicals**

Two Bulk drug substance of montelukast were obtained from (Inogent Laboratory private limited) Nacharam
Hyderabad, India. HPLC grade acetonitrile and methanol were purchased from Panreac Chemicals Corporation Ltd. Spain. Water used for the preparation of mobile phase was purchased from Chem – lab (Belgium). Tri fluoro acetic acid TFA was purchased from Sigma-Aldrich (USA) and ortho phosphoric acid was obtained from BDH Laboratory Supplies (Poole, UK).

**High Performance Liquid Chromatography (HPLC) With MS Detector**

An Agilent HPLC system and an Eclipse XDB C18 column (250x4.6 mm, 5 µm particle size) was used as stationary phase. A mobile phase consisting of solution A 0.1% ortho phosphoric & solution B: 80 % acetonitrile + 20 % water, with a time gradient program of T (min)/%B (v/v). Initial gradient of B starts with 60%, at 10 min it is 70%, at 15 min it is 90%. The ratio being increased to 100% at 20 min then decrease to 60 % at 32 min, and maintained 60% until 45 min with a flow rate of 0.4ml/min. The electrospray ionization and MS studies were performed on a triple quadruple mass spectrometer PE Sciex model API 3000.

The instrument parameters used are:
- Capillary: 2000 V
- Extractor: 2500v
- Source temperature: 350 C
- Gas flow: 10 L/min
- Scan range: 100-800 mu
- Injection volume: 20 µl

**High Performance Liquid Chromatography (HPLC) With UV Detector**

A Hitachi HPLC system containing a L 2130 pump plus degasser, L 2200 auto-sampler, L 2420 UV/VIS Detector.

A SHIMADZU L11 column (150x4.6 mm, 5 µm particle size) was used as stationary phase. A mobile phase consisting of solution A: 0.15% trifluoroacetic acid (TFA) in water & solution B: 0.15% TFA in acetonitrile in ratio of 40% of solution B from 0 – 3 min in isocratic elution to increase to 58.6% at 25 min in a liner gradient. The mobile phase was pumped at a flow rate of 1.2 ml / min the eluents were monitored at 238 nm. 13

**Sample Preparation for HPLC-UV**

About 50 mg of bulk MO was weighed accurately and transferred to 50 ml volumetric flask. The volume was made up to mark with the diluent (Methanol and water 9:1) to obtain a concentration of 1mg/ml.

**Sample Preparation for LC-MS**

About 50 mg of bulk MO was weighed accurately and transferred to 50 ml volumetric flask. The volume was made up to mark with the diluent (acetonitrile and water 8:2) to obtain a concentration of 1mg/ml.

**RESULTS AND DISCUSSION**

**Results**

MO impurities (related to synthesis as well as degradation processes) were identified using LC-MS and HPLC-UV. Bulk drug substances of MO were analyzed by LC-MS using scan mode from 100-800 mu to produce spectra of molecular weight due to different types of impurities, reported in tables (1) and (2).

<table>
<thead>
<tr>
<th>Compound</th>
<th>Molecular weight</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sulfoxide impurity</td>
<td>602</td>
</tr>
<tr>
<td>methylketone impurity</td>
<td>570</td>
</tr>
<tr>
<td>Methylstyrene impurity</td>
<td>568</td>
</tr>
<tr>
<td>Dihydro impurity</td>
<td>588</td>
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<td>588</td>
</tr>
<tr>
<td>Michael Adducts 1c and 2d</td>
<td>732</td>
</tr>
<tr>
<td>Unknown impurity</td>
<td>772</td>
</tr>
</tbody>
</table>

The same bulk drug substances of MO were analyzed by HPLC-UV to produce chromatograms of MO with different types of impurities, reported in tables (3) and (4).

**Table 3: Impurities of MO by UV in sample (1)**

<table>
<thead>
<tr>
<th>Compound</th>
<th>RT</th>
<th>RRT</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sulfoxide impurity</td>
<td>5.55</td>
<td>0.5</td>
</tr>
<tr>
<td>Cis-isomer impurity</td>
<td>9.51</td>
<td>0.88</td>
</tr>
<tr>
<td>Methylketone impurity</td>
<td>11.65</td>
<td>1.1</td>
</tr>
<tr>
<td>Un known impurity1</td>
<td>13.71</td>
<td>1.27</td>
</tr>
<tr>
<td>Un known impurity2</td>
<td>17.25</td>
<td>1.6</td>
</tr>
</tbody>
</table>

**Table 4: Impurities of MO by UV in sample (2)**

<table>
<thead>
<tr>
<th>Compound</th>
<th>RT</th>
<th>RRT</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sulfoxide impurity</td>
<td>5.33</td>
<td>0.5</td>
</tr>
<tr>
<td>Cis-isomer impurity</td>
<td>9.20</td>
<td>0.88</td>
</tr>
<tr>
<td>Un known impurity1</td>
<td>17.05</td>
<td>1.63</td>
</tr>
<tr>
<td>Un known impurity2</td>
<td>17.57</td>
<td>1.67</td>
</tr>
</tbody>
</table>

A typical chromatogram showing the MO separated from its related impurities of the drug is given in Fig.2.
Montelukast in Human Plasma by HPLC Coupled with ESI-MS/MS: Application to a Bioequivalence Study, Scientific Pharmedaceutica, 30, 2010, 411-422.

REFERENCES
2. Balasekhar Reddy Challa, Bahlul Z. Awen, Babu Rao Chandu, Method Development and Validation of

DISCUSSION
More MO impurities were detected and identified by using MS detector, on the other hand less impurities were detected and identified by using UV detector.

Identification of impurities in MS was done by using molecular weight, while in UV it was done by using relative retention time RRT.

Michael Adducts 1c and 2d impurities were detected by MS only, because it is very hard to be resolve them from MO peak.

A major degradation product "cis-isomer impurity" which rises very fast by exposure to ambient light affecting substance quality14, was identified by UV only, because cis-isomer impurity has the same molecular weight as MO that make identifying it by MS detector not possible, while it is separated using the HPLC-UV method with a RRT of 0.88

CONCLUSION
The use of LC-MS is sufficient for impurity identification and is a common approach within the early stages of pharmaceutical development but isn’t sufficient in the case of drugs which have isomers impurities such as MO with cis-isomer impurity. We should integrate data from two detectors (MS, UV) to possess full information for identification of MO impurities.

Figure 2: Typical chromatogram of MO and its impurities in sample (1)

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