Research Article



Antimicrobial Efficacy of Silver Nanoparticles Synthesized from *Withania somnifera* – An Important Ethnomedicinal Herb of Kurnool District, Andhra Pradesh, India

Venkata Subbaiah KP*, Savithramma N

Department of Botany, Sri Venkateswara University, Tirupati, A.P. India. *Corresponding author's E-mail: subbupandu2@gmail.com

Accepted on: 01-07-2013; Finalized on: 31-08-2013.

ABSTRACT

Withania somnifera roots are extensively using by the ethnic groups of Kurnool district, Andhra Pradesh, India to cure leucoderma. Biological synthesis of silver nanoparticles was carried out from root aqueous extract of Withania somnifera 10 ml root extract was mixed to 90 ml of 1 mM aqueous of $Ag(NO_3)_2$ and was heated at $60-80^{\circ}$ C for 20 min. The colour change of aqueous solution into dark brown colour. For characterization using UV-Vis spectrophotometer and AFM. AFM, UV-Vis spectrophotometer showed the formation of silver nanoparticles with spherical shape and average size 25.02 nm. SNPs have good antimicrobial activity against different microorganisms.

Keywords: Anti microbial efficacy, Atomic Force Microscope (AFM), Inhibition zone, Medicinal plant, Secondary metabolites, Silver nanoparticles.

INTRODUCTION

ithania somnifera L. is an important medicinal plant, the roots of which have been employed in Indian traditional systems of medicine, Ayurveda and Unani. The plant has been found useful in the treatment of burns, wounds and dermatological disorders. W. somnifera reduces tumor cell proliferation and enhances the effectiveness of radiation therapy while potentially mitigating undesirable side effects. India is one of the twelth mega biodiversity hotspot in the world. Eastern Ghats are one among them, which are characterized by different wild medicinal flora. Nallamalai hills as a part of Eastern Ghats in Kurnool District of Andhra Pradesh, India. Mainly four ethnic groups (Chenchu, Sugali, Yerukala and Yanadi) are inhabited in this region.

Leucoderma is a skin disorder in humen for a number of reasons depigmentation occurs due to auto immune disorder³ or lacking of pigments due to absence of melanocytes.⁴ Skin diseases are commonly occurring among the rural masses due to poor hygienic conditions, poor sanitation facilities and contaminated water etc., the traditional healers of these ethnic groups are extensively using *W. somnifera* roots to treat leucoderma.

W. somnifera root contains a variety of important chemical compounds. Withaferin⁵ and reported to have immunosuppressive.⁶ Withanolid A, Withanone and Withanolid D are present in the medicinal plant are reported to have antioxidant, immunomodulatory and other activities.⁷⁻⁹ The most important of these phytochemicals are alkaloids, tannins, flavonoids and phenolic compounds.¹⁰

Nanoparticles are gaining much importance especially in medicinal field. Synthesis of metal nanoparticles receives great attention due to their unusual optical, chemical, phytochemical and electronic properties.¹¹ Silver a nobel metal is known to improve the immunity since ancient times, Ag(NO₃)₂ was using for biosynthesis of nanoparticles by using root aqueous extract of *W. somnifera*. The possibility of using plant materials for the synthesis of nanoscale metals was initially reported by Gradea-Torresdey.¹²⁻¹³ SNPs have particular properties that may perhaps have numerous applications in the field of dentistry, clothing, catalysis, mirrors, optics, phytography, electronics and food industry.¹⁴

At present extensive work has been done to develop new drug from natural products because of the resistance of microorganisms to the existing drug. The pathogens like *E. coli, Bacillus, Salmonella typhi* and *Staphylococcus aureus.* ¹⁵

The present study is aimed to study the qualitative analysis of phytochemical constituents and biological synthesis of silver nanoparticles by using root extract of *W. somnifera* L. and also screening of SNPs for microbial efficacy.

MATERIALS AND METHODS

Plant material

The fresh roots of *W. somnifera* L. was collected in January 2012 from Srisailam reserve Forest, Kurnool District of Andhra Pradesh, India. The root was cleaned, cut into small pieces (1-2 cm) dried at room temperature and ground to fine powder.

Preparation of extract

25 g of root powder of *W. somnifera* L. was taken into 250 ml conical flask and added 100 ml of sterile distilled water and boiled for 10 min at 100°C on water bath. Then plant material extracts were collected in separate conical flask by standard filtration method and stored in refrigerator for further use.



Phytochemical screening

10 ml root extract was used for preliminary phytochemical screening. The qualitative analysis of secondary metabolites was carried out by using the methods for flavonoids¹⁶; steroids, alkaloids and phenols¹⁷; triterpenoids and glycosides¹⁸; for tannins, anthraquinons, leucoanthocyanins and emodins¹⁹; saponins²⁰ and reducing sugars and anthocyanins.²¹

Preparation of 1 mM Silver nitrate solution

1 molar silver nitrate stock solution was prepared by 1.7 g of $AgNO_3$ was dissolved in 10 ml distilled water. 1 mM solution was prepared by 1 ml of 1 M solution was made up to 100 ml with 99 ml of distilled water. This solution was stored in amber colored bottle for further use.

Synthesis of silver nanoparticles

SNPs were synthesized by using root powder extract of *Plumbago zeylanica*. The reduction of pure Ag²⁺ ions were monitored by measuring the UV-Vis spectrum of the reduction media at 5th h after diluting a small aliquot of the sample in distilled water by using Systronic 118 UV-Vis Spectrophotometer. The size and shape of SNPs were confirmed with AFM.

UV-Vis spectra analysis

The reduction of pure silver ions was monitored by measuring the UV-Vis spectrum of the reaction medium at 3 hrs. after diluting a small aliquot of the sample into distilled water. UV-Vis spectral analysis was carried out by using UV-Vis spectrophotometer (Systronics type 118).

AFM analysis

The silver nanoparticles extracted by the above protocol were visualized with an Atomic Force Microscope (AFM). A thin film of the sample was prepared on a glass slide by dropping 100 μl of the sample on the slide and was allowed to dry for 5 min, the slides were then scanned with the AFM (Nano surf $^{\circ}$ AG, Switzerland, Product: BTO2089, 3RO). Nanosurf $^{\circ}$ Easyscan-2 software was used for the AFM Analysis (VIT, Vellore, Tamil Nadu).

Microorganisms

Pure cultures of *Escherichia coli*, *Bacillus subtilis*, *Staphylococcus aureus*, *Salmonella typhi* species of bacteria and *Paecilomyces varioti*, *Pencillium rubrum* and *Aspergilus flavus* species of fungi were procured from the Department of Microbiology of Sri Venkateswara Institute of Medical Sciences (SVIMS), Tirupati, Andhra Pradesh, India.

Antimicrobial activity

The antimicrobial activities of SNPs were carried out with paper disc diffusion method using nutrient agar medium and potato dextrose agar medium for bacterial and fungal cultures respectively. Zones of inhibition for control, SNPs and silver nitrate were measured after 24 h and 7 days and compared with standard drugs Gentamycin and Nystatin for bacterial and fungal growth respectively. The

experiments were repeated thrice and mean values of inhibition zone diameter were presented.

RESULTS AND DISCUSSION

The ethnic groups (Chenchu, Sugali, Yanadi and Yerukala) of Kurnool District, Andhra Pradesh, India. The traditional healers of these ethnic groups have staunch confidence to treat leucoderma. The fresh roots of *W. somnifera* L. ground in to paste is applied to depigmented parts of skin to treat leucoderma. This information was cross checked with Ayurvedic Physicians Sri Venkateswara Ayurvedic College, Tirupati, Andhra Pradesh, India for authentication. In this regard *W. somnifera* L. roots are extensively used in Ayurveda to treat leucoderma.

The phytochemical study of W. somnifera L. showed that the root is rich in alkaloids, antraquinones, flavonoids, glycosides, phenols, reducing sugars, saponins, steroids, tannis and triterpenoids and lacking antrocyanins, coumarins, emodins, lignins and laucoanthocyanins (Table 1). The secondary metabolites like flavonoids and phenolic compounds are medically used as antistomatic, diarrhoea. anti-inflammatory, anticancer antioxidative. The biological function of flavonoids include protection against allergies; inflammations, platelets aggregation, microbes, ulcer and tumors. 22-23 It also known to possess antiviral and anti fungal²⁴⁻²⁵ and antimicrobial properties.²⁶ The presence of bioactive compounds indicates the medicinal values of the plant. Steroids and triterpenoids possess anti bacterial activity.²⁷ possess astringent, anti-inflammatory, antidiarrhoeal, antioxidant and antimicrobial activities. 28 Similar chemical constituents were also found in Shorea tumbuggaia²⁹, Thespesia populnea³⁰ and in Curcuma longa.3

Table 1: Secondary metabolites of root extract of *W. somnifera* L.

Root
++
-
+
-
-
-
++
++
-
-
++
+
+
++
+
++

Note: '+' indicates presence, '++' indicates presence of more amounts, '-' indicates absence



In the present study SNPs were synthesized by using root aqueous extract of *W. somnifera* rapidly within 10 min of incubation period dark brown color was developed by addition of Ag(NO₃)₂ (Figure 1a). The appearance of dark-brown color in the reaction vessels indicates the formation of SNPs. The colour change in aqueous solution is due to the surface- Plasmon resonance (SPR) phenomina. The reason could be that the quantitative variation in the formation of SNPs (or) availability of H⁺ ions to reduce the silver. It is well known that SNPs exhibit dark brown color in aqueous solution due to excitation of surface plasmon vibrations in silver nanoparticles. By using medicinal plants Ag(NO₃)₂ can be reduced into SNPs at the fast rate. The biomolecules found in plants induce in reduction of Ag⁺ ions into silver nanoparticles.

The synthesis of SNPs had been confirmed by measuring the UV-Vis spectrum of the reaction media. The UV-Vis

spectrum of colloidal solutions of SNPs synthesized from root extract of W. somnifera have the characteristic absorbance peaks at 260 nm and 440 nm (Figure 1b) and the broadening of peak indicated that the particles are poly-dispersed. This peak illustrates that the presence of homogenous distribution of hydrosol nanoparticles after stirring.34 It is generally recognized that UV-Vis spectroscopy could be used to examine size and shape of nanoparticles in aqueous suspension.³⁵ The weak absorption peak at shorter wavelengths due to the presence of several organic compounds which are known to interact with silver ions same results observed in *Boswellia ovalifoliolata* stem bark.³⁶ Silver nanoparticles have free electrons, which give rise to an SPR absorption bonds³⁷, due to the combined vibration of electrons of metal nanoparticles in resonance with the light waves.³⁸-³⁹ The secondary metabolites present in plant systems may be responsible for the reduction of silver and synthesis of nanoparticles.

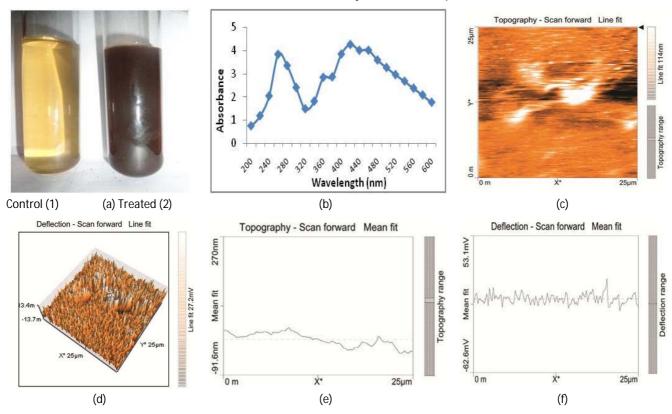


Figure 1: The color change of root extract of *Withania somnifera* (1) blank extract without silver nitrate (2) root extract with 1 mM silver nitrate; b) UV-Vis spectroscopy of synthesized of silver nanoparticles, (c, e & f) AFM of Topography of SNPs (d) Three dimensional structure of SNPs.

The size and shape of SNPs was detected by using AFM (Atomic Force Microscope). Size of SNPs was 25.02 nm spherical in shape (Figure 1d). The results obtained in this study are interesting because it can serve as a foundation in terms of identification of potential medicinal plants for the synthesis of SNPs. The plant species pay a vital role to cure skin diseases. Biological synthesis of metal nanoparticles in a traditional method and the use of plants extract have a new awareness for the control of diseases, besides being safe, eco-friendly and no phytotoxic effects.

The SNPs of root extract of w. somnifera showed highest percentage of bacterial inhibition towards gram-negative bacterias Salmonella typhi (11.6±0.23) and E. coli (11.2±0.38) followed by gram-positive bacterias Staphyllococcus aureus (10.68±0.31) and Bacillus (10.61±0.83) and antifungal activity against Pencillium rubrum (8.8±0.66) followed by Aspergillus flavus (8.5±0.50) and Paecilomyces varioti (7.3±0.49) (Table 2), (Figure 3) and (Figure 2). The maximum toxicity was observed in SNPs than $Ag(NO_3)_2$ and plant extract. The reason could be that the smaller size of the particles



which leads to increased membrane permeability and cell destruction. The results were compared to that of standard antibiotics Gentamycin / Nystatin anti bacterial and antifungal respectively. Standard drugs (Gentamycin / Nystatin), showed higher inhibition zones, because these are highly purified forms which may be cost and leads to side effects in high dosage, whereas the SNPs are biologically synthesized form with less in cost, ecofriendly, safe and pollutant free with less or no side effects.

In general, gram-positive bacteria appeared to be more tolerant to silver than gram-negative cells. The cell wall of

gram-positive bacteria contains multiple layers of peptidoglycon compared to the cell wall of gram-negative bacteria. Thus, gram-positive bacteria may allow less Ag⁺ to reach the cytoplasmic membrane than gram-negative bacteria⁴⁰ and may therefore be less susceptible. The SNPs are also reported to be nontoxic to human and most effective against bacteria, viruses and other eukaryotic micro-organisms at very low concentrations and without any side effects Jeong et al., SNPs synthesized from Svensonia hyderobadensis⁴², Shorea tumbuggaia and Boswellia ovalifoliolata^{43, 29} and Thespesia populnea³⁰, Curcuma longa exhibit the antibacterial activity.

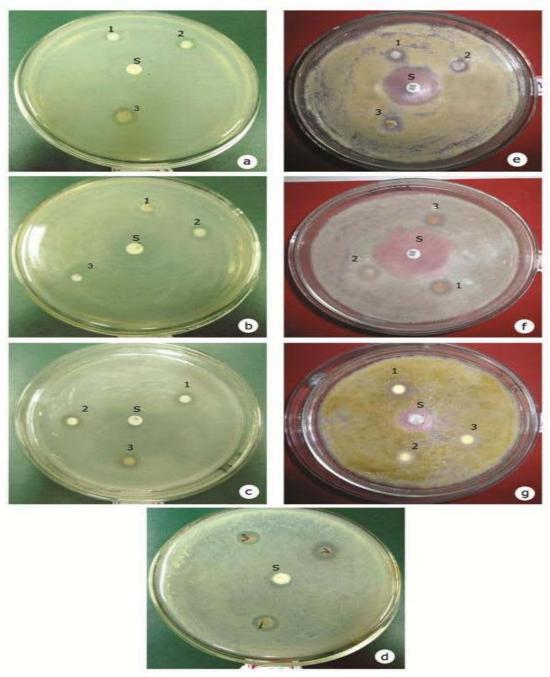


Figure 2: Antimicrobial activity of root extract of Withania somnifera

a) Staphylococcus aureus, b) Salmonella typhi, c) E.coli, d) Bacillus, e) Paecilomyces varioti, f) Pencillium rubrum and g) Aspergillus flavus. 1) $Ag(No_3)_2$, 2) Plant extract control, 3) SNPs and S) Standard(Gentamycin/Nystanin)



Inhibition zone in mm Microorganisms Plant extract control **SNPs** Standard: Gentamycin/ Nystatin $Ag(NO_3)_2$ **Bacterial species** 8.0±0.40 9.3 + 0.7710.68±0.31 15.8±0.22 Staphylococcus aureus Salmonella typhi 9.6±0.16 8.9±0.23 11.60±0.23 13.7±0.16 8.1±0.24 5.6±0.21 11.2±0.31 11.8±0.35 E.coli Bacillus 8.2±0.33 9.2±0.38 10.70±0.38 12.6±0.26 **Fungal species** Paecilomyces varioti 7.0±0.40 6.8±0.55 7.3±0.49 11.7±0.43 Pencillium rubrum 7.2±0.68 7.5±0.47 8.8±0.66 13.7±0.54 Aspergillus flavus 7.6±0.47 6.4±0.54 8.5±0.50 11.8±0.72

Table 2: Antimicrobial activity of SNPs isolated from root extract of Withania somnifera

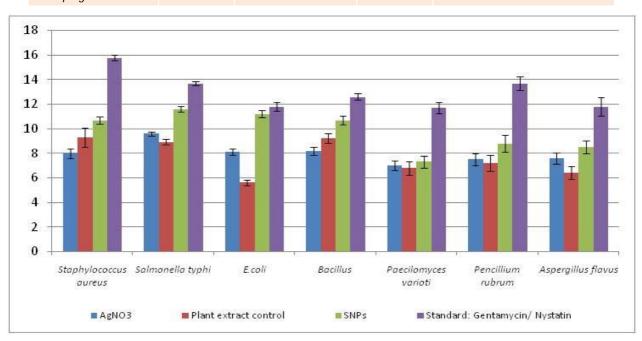


Figure 3: Antimicrobial activity of SNPs isolated from root extract of Withania somnifera

CONCLUSION

The present study includes the treatment of leucoderma using w. somnifera by the ethnic groups. Phytochemical screening indicates that the plant part is a good source for bio active principle for pharmacognostic and pharmaceutical industries. The SNPs prepared by using the aqueous root extract of w. somnifera. The aqueous silver ions exposed to the extracts, the synthesis of SNPs were confirmed by the change of color of plant extracts. These environmentally benign SNPs were further confirmed by using UV-Vis spectroscopy finally the size and shape of the SNPs was characterized by AFM analysis. The results indicated that SNPs have good antimicrobial activity against different microorganisms due to the cumulative effect of secondary metabolites or active molecules present in the plant extract of selected medicinal plant used by ethnic groups of Kurnool district of Andhra Pradesh, India to cure skin diseases. It is confirmed that SNPs of w. somnifera are capable of rendering antimicrobial efficacy and hence has a great potential in the preparation of drugs used against bacterial and fungal diseases.

Acknowledgements: Authors are highly thankful to Ethnic groups of Kurnool district for providing valuable ethnic information and to VIT University, Tamil Nadu for AFM studies.

REFERENCES

- Grierson DS, Afolayan AJ, Antibacterial activity of some indigenous plants used for the treatment of wound in the Eastern Cape, S Afr J Ethnopharmacol, 66, 1999a, 103-106.
- Kaur K, Rani G, Widodo, Evaluation of the anti-proliferative and anti-oxidative activities of leaf extract from *In vivo* and *In vitro* raised aswagandha, Food chem Toxicol, 42, 2004, 2015-2020.
- Lerner AB, Nordland JJ, Vitiligo: What is it? Is it important? AMA, 239, 1978, 1183-1187.
- Aaron B, Lerner MD, Tetsuo shiohara, A mouse model for vitilago, The Journal of Investigative dermatology, 87(3), 1986, 299-304.



- Sethi PD, Thiagrajan AR, Subramanian S, Studies on the anti-inflammatory and anti-arthritic activity of Withaferin-A, Indian journal of pharmacology, 2, 1970, 165.
- 6. Jayaprakashan B, Nair MG, Cyclooxygenase-2 enzyme inhibitory Withanolides from *Withania somnifera* leaves, Trtrahedron, 59, 2003, 841-849.
- 7. Furmanowa M, *In vitro* propagation of *Withania somnifera* and isolation of Withanolides with immunosuppressive activity, Planta Med, 67, 2001, 146-149.
- 8. Bhattacharya SK, Satyan KS, Ghosal S, Antioxidant activity of glycowithanolides from *Withania somnifera*, Indian J Experimental Biology, 35, 1997, 236-239.
- Zhao J, Nakamura N, Hattori M, Kuboyama T, Tohda C, Komatsu K, Withanolide derivatives from roots of Withania somnifera. And their neuriten outgrowth activities, Chem Pharm Bull, 50, 2002, 760-765.
- Nathiya S, Santhi N, Kalaiselvi S, A comparative study on ontogenic expression of antioxidants and secondary metabolites in *Withania somnifera*. Int Re J Pharmacy, 3(1), 2012, 210-216.
- 11. Mohanpuria P, Rana NK, Yadav SK, Biosynthesis of nanoparticles: technological concepts and future applications, J Nanopart Res, 10, 2008, 507-517.
- 12. Gradea-Torresdey JL, Parsons JG, Dokken K, Formation and growth of Au nanoparticles inside live alfalfa plants, Nanolet, 2002, 397-401.
- 13. Gradea-Torresdey JL, Gomez E, Paralta-videa JR, Parsons JG, Troiani H, Jose-Yacaman M, Alfalfa sprouts: A natural source for the synthesis of silver nanoparticles, Longmuir, 19, 2003, 1357-1361.
- Rai M Yadav A, Gade A, Silver nanoparticles as a new generation of antimicrobials, Biotechnol Adv, 27, 2009, 76-83.
- 15. Patil RS, Kokate MR, Kolekar SS, Bio-inspired synthesis of highly stabilized silver nanoparticles using *Ocimum tenuiflorum* leaf extract and their antibacterial activity, Spectro Act Part A 91, 2012, 234-238.
- 16. Peach K, Tracey MV, Modern Methods of Plant Analysis, Springer Verlag, Berlin, 3, 1956, 5-8.
- 17. Gibbs RD, Chemotaxonomy of Flowering Plants, Mc Gill Queen's University Press, Montreal, ISBN. 0773500987, London, 1, 1974, 22-36.
- 18. Ayoola GA, Coker HAB, Adesegun SA, Adepoju Bello AA, Obaweya K, Ezennia EC, Atangbayila TO, Phytochemical screening and antioxidant activities of some selected medicinal plants used for malaria therapy in South Western Nigeria, Trop J Pharm Res, 7, 2008, 1019-1024.
- 19. Treare GE, Evans WC, Pharmacognosy 13th ed. ELBS/Bailliere Tindal, London UK, 1982, 61-67.
- Kumar A, Ilavarasn R, Jayachandran T, Decaraman M, Aravindhan P, Padmanaban N, Krishnan MRV, Phytochemical investigation on a tropical plant, Pakistan Journal of Nutrition, 8, 2009, 83-85.
- 21. Paris R, Moyse H, Precis de matiere medicinal, Paris: Masson, 1969.

- 22. Okwu DE, Okwu ME, Chemical composition of *Spondias mombin* plants, J Sustain Agri Environ, 6, 2004, 140-147.
- 23. Farquar JN, Plants sterols, their biological effects on humans, Handbook of lipids in human nutrition, BOCA Rotan Hr CRC Press, 1996, 101-105.
- 24. Fairbarin JW, The pharmacology of plant phenolics, Academic Press, Newyork, 1959, 45-47.
- 25. Tripati VD, Rastogi RP, Flavonoids in biology and medicine, J Sci Indian Res, 40, 1981, 116-124.
- Takahasi T, Kokubo R, Sakaino M, Antimicrobial activity of Eucalyptus leaf extracts and flavonoids from *Eucalyptus* maculate, Lett Appl Microbiol, 39, 2004, 60-64.
- 27. Reddy V, Ravindra K, Srinivasulu M, Goud T, Reddy M, Kumar D, Rao T, Murthy U, Venkateswarlu Y, Two new macrocyclic diaryl ether heptanoids from *Boswellia ovalifoliolata*, Chem Pharm Bull, 51, 2003, 1081-1084.
- 28. Killender SG, More HN, Estimation of tannins in different parts of *Memecylon umbellatum*, Burn J Phar Res, 3, 2010, 554-556.
- Ankanna S, Savithramma N, Biological syntheses of silver nanoparticles by using stem of *Shorea tumbaggaia* Roxb and its antimicrobial efficacy, Asian J Pharm Clin Res, 4(2), 2011, 137-141.
- Bhumi G, Lingarao M, Savithramma N, Biological synthesis of silver nanoparticles from stembark of *Thespesia* populnea (L.) Soland, Indian Steams Research J, 3(3), 2013, 1-7
- Venkata Subbaiah KP, Ramanjaneyulu G, Savithramma N, Synthesis of silver Nanoparticles and validation from Rhizome powder of *Curcuma longa* L.- An Ethnobotanical plant for skin disease, Indian steam Res Journal, 3(5), 2013, 1-7
- 32. Shankar SS, Rai A, Ahmad A, Sastry MJ, Rapid synthesis of Au, Ag and Bimetallic Aushell nanoparticles using Neem, J Colloid Inter Sci, 275, 2004, 496-502.
- 33. Thirumurugan A, Tomy NA, Jai Ganesh R, Gobikrishnan S, Biological reduction of silver nanoparticles using plant leaf extracts and its effect an increased antimicrobial activity against clinically isolated organism, De Phar Chem, 2, 2010, 279-284.
- 34. Shameli K, Ahmed MB, Jazayeri SD, Synthesis and characterization of polyethylene glycol mediated silver nanoparticles by the green method, Int J Mol Sci, 13, 2012, 6639-6650.
- Wiley BJ, Im SH, Li ZY, McLellan J, Siekkinen A, Xia Y, Maneuvering the Surface Plasmon Resonance of Silver Nanostructures through Shape-Controlled Synthesis, J Phys Chem, 110, 2006, 15666-15675.
- 36. Ankanna S, Prasad TNVKV, Elumalai EK, Savithramma N, Production of Biogenic silver nanoparticles using *Boswellia ovalifoliolata* stem bark, Digest J Nano Biostruct, 5, 2010, 369-372.
- 37. Noginov MA, Zhu G, Bahoura M, Adegoka J, Small C, Ritzo BA, Drachev VP, Shalaev VM, The effect of gain and absorption on surface Plasmon in metal nanoparticles, Appl Phy B, 86, 2006, 455-460.



- 38. Nath SS, Chakdar D, Gope G, Synthesis of CdS and ZnS quantum dots and their applications in electronics, Nano J Nanotech App, 2, 2007, 1-5.
- 39. Dubey M, Bhadauria S, Kushwah BS, Green synthesis of nanosilver particles from extract of *Eucalyptus hybrid* (Safeda) leaf, Dig J Nano Biostr, 4, 2009, 537-543.
- Kawahara K, Tsuruda K, Morishita M, Uchida M, Antibacterial effect of silver-zeolite on oral bacteria under anaerobic conditions, Dent Mater, 16, 2000, 452–455.
- 41. Jeong SH, Yeo SY, Yi SC, The effect of filler particle size on the antibacterial properties of compounded polymer/silver fibers, J Materials Sci, 40(20), 2005, 5407-5411.
- 42. Lingarao M, Savithramma N, Biological synthesis of silver nanoparticles using *Svensonia hyderobadensis* leaf extract and evaluation of their antimicrobial efficacy, J Pharm Sci Res, 3, 2011, 1117-1121.
- 43. Savithramma N, Lingarao M, Suvarnalatha Devi P, Evaluation of antibacterial efficacy of biologically synthesized silver nanoparticles using stem barks of *Boswellia ovalifoliolata* Bal. and Henry and *Shorea tumbuggaia* Roxb, J Biol Sci 11, 2011, 39-45.

Source of Support: Nil, Conflict of Interest: None.

