



# Chemical Composition and Antioxidant Activity of Schizophyllum commune

L. Sushila Devi<sup>1, 4</sup>, Adhiraj Dasgupta<sup>2</sup>, Mayukh Chakraborty<sup>3</sup>, S.K. Borthakur<sup>1</sup>, N. Irabanta Singh<sup>4\*</sup>

<sup>1</sup> Botany Department, Guwahati University, Guwahati – 781014, India.

<sup>2</sup> Department of Botany, University of Calcutta, Kolkata – 700019, India.

<sup>3</sup> Department of Biotechnology, St. Xavier's College, Kolkata - 700016, India.

<sup>4</sup> Centre of Advanced Study in Life Sciences, Manipur University, Canchipur, Imphal – 795003, India.

\*Corresponding author's E-mail: <u>irabanta.singh@gmail.com</u>

### Accepted on: 18-05-2014; Finalized on: 31-07-2014.

#### ABSTRACT

This study was conducted with ethanolic extract of edible mushroom, *Schizophyllum commune*, which was tested for total phenol, flavonoid,  $\beta$ -carotene, lycopene and ascorbic acid and *in vitro* antioxidant activity in terms of DPPH radical scavenging activity and reducing power and total antioxidant capacity were done with varying concentrations. Findings showed that most of the EC<sub>50</sub> values were more than 2 mg /ml except DPPH radical scavenging test. The extract exhibited 50% DPPH radical scavenging activity at only 1.5 mg /ml concentration. Estimated putative antioxidant components was in order of phenol > ascorbic acid > flavonoids >  $\beta$ -carotene > lycopene. Result implies that *S. commune* can be a potential source of natural antioxidants which may be used as a food supplement to treat various oxidative stress related diseases.

Keywords: Antioxidant, DPPH, Flavonoid, Schizophyllum commune, Phenol, Reducing power.

## **INTRODUCTION**

ifferent biochemical reactions in our body generate free radicals and under different pathophysiological conditions, the balance between the generation and elimination of reactive oxygen species (ROS) is broken, as a result a wide range of essential biomolecules are damaged by this ROS mediated oxidative stress, leading to various diseases<sup>1</sup>. Every organism is equipped with defence system against oxidative damage but they often fall short, thus making the search for external antioxidants increasingly important. Antioxidants can protect human body from damages caused by reactive oxygen species. Although synthetic antioxidants are available in the market, their use is being restricted due to reported carcinogenicity and hepatotoxicity. Thus, in recent times anti-oxidants from natural sources are being sought extensively<sup>2</sup>.

Among different natural sources mushrooms are now becoming more attractive because of its strong nutritional value and therapeutic potentiality. Mushrooms are rich in different biologically active compounds like phenolics, tocopherol, lycopene, βcarotene etc.<sup>3</sup>. Thus, mushrooms in recent times are gradually rising to an elite position as a source of bioactive compounds. Because of the geoclimatic variation India becomes a harbour for a large number of edible mushrooms, many of which have been evaluated for their therapeutic potentiality and established to have anticancer<sup>4,5</sup>, antidiabetic<sup>6</sup>, ective<sup>8-11</sup>, cardioprotective<sup>12</sup>, antiulcer<sup>7</sup>. effective hepatoprotective<sup>8-11</sup>, immune modulatory<sup>13-15</sup>, antimicrobial<sup>16-19</sup> and apoptogenic<sup>20,21</sup> activities. In the present investigation, we report the antioxidant properties of Schizophyllum commune based on in vitro antioxidant assay systems.

# MATERIALS AND METHODS

### **Preparation of extract**

Ethanolic extract of the sample was prepared following Dasgupta et al (2013)<sup>22</sup>. The sample was dried, powdered and extracted with ethanol at 25°C for 2 days. After filtration, the residue was then re-extracted with ethanol, as described above. The supernatant was concentrated under reduced pressure in a rotary evaporator. Then, this ethanolic extract of *Schizophyllum commune* was stored at 4°C for further analysis. The percentage yield extract was calculated based on dry weight as follows:-

Where W1 = weight of extract after solvent evaporation; W2 = Weight of the minced mushroom.

### Determination of total phenolic content

Total phenols were estimated following Singleton and Rossi  $(1965)^{23}$ . To the extract sample  $(100 \ \mu l)$ , 1 ml of 1N Folin-Ciocalteu reagent was added. After 4 min, a saturated sodium carbonate solution (approximately 35 g /100ml, 1 ml) was added to it. The absorbance of the reaction mixture was measured at 725 nm after incubation for 1 hr 30 min at room temperature. Gallic acid was used as a standard and the results were expressed as milligram gallic acid equivalent (mg GAE) /g of extract.

### **Determination of flavonoids**

100  $\mu$ l of the sample extract was added to 80% ethanol containing 0.1 ml of 10% aluminium nitrate and 0.1 ml of 1M Potassium acetate. The mixture was incubated at room temperature for 40 min and its absorbance was measured at 415 nm<sup>24</sup>. Quercetin was used as standard.



Available online at www.globalresearchonline.net

## Determination of β-carotene and lycopene

For  $\beta$ -carotene and lycopene determination, 100  $\mu$ l of the sample extract was vigorously shaken with 10 ml of an acetone–hexane mixture (4:6) for 1 min and filtered through Whatman No. 4 filter paper. The absorbance of the filtrate was measured at 453, 505, and 663 nm<sup>25</sup>.  $\beta$ -carotene and lycopene content were calculated according to the following equations:-

Lycopene (mg /100 ml) =  $-0.0458 \times A663 + 0.372 \times A505 - 0.0806 \times A453$ 

 $\beta$ -carotene (mg /100 ml) = 0.216×A663 0.304×A505 + 0.452×A453

The results are expressed as mg of carotenoid/g of extract.

## Ascorbic acid content determination

Ascorbic acid content was determined following Rekha et al  $(2012)^{26}$  with a little modification. Standard ascorbic acid (100 µg /ml) was taken in a conical flask and made up to 10 ml by 0.6% oxalic acid. It was titrated with a dye, 2, 6-dichlorophenol indophenol. The amount of dye consumed (V1 ml) is equivalent to the amount of ascorbic acid. The sample (w µg /ml) was similarly titrated with the dye (V2 ml). The amount of ascorbic acid was calculated using the formula:-

Ascorbic acid (µg /mg) = [{(10 µg /V1ml) × V2 ml} × w µg] × 1000

# **Reducing power**

Reducing power of the sample was determined following Oyaizu (1986)<sup>27</sup>. Varied concentrations of the sample (0.5 - 2 mg /ml) were added to 2.5 ml of 0.2 M phosphate buffer (pH 6.6) and 2.5 ml of 1% potassium ferricyanate. 2.5 ml of 10% trichloro acetic acid was added to the mixture after an incubation of 20 min at 50°C. It was then centrifuged for 10 min at 12000 rpm. 2.5 ml of the supernatant was mixed with distilled water and 0.5 ml of 0.1% ferric chloride. Its absorbance of the reaction mixture was interpreted as an increase in reducing power of the sample. Antioxidant has ability of donation of electron and causes conversion of the oxidation form of iron (Fe<sup>+3</sup>) in ferric chloride to ferrous (Fe<sup>+2</sup>). Resulting Perl's Prussian blue is measured at 700 nm and higher absorbance indicates higher reducing power.

# **DPPH radical scavenging assay**

The model of scavenging DPPH radical is a widely used method to evaluate antioxidant activities in a relatively short time compared with other methods. Effect of antioxidants on DPPH free radicals is due to their hydrogen donating ability. Upon reduction, solution of DPPH fades from purple to yellow. Thus, a lower absorbance at 517 nm indicates a higher radical scavenging activity of extract. The DPPH radical scavenging ability of the sample extract was measured according to Shimada et al (1992)<sup>28</sup>. 2 ml of reaction

mixture was prepared using different concentrations of sample (1 - 2.5 mg /ml) and methanol solution of DPPH (0.004 %) (w /v). The absorbance was read against a methanol blank at 517 nm after 30 min incubation at room temperature in dark. The degree of scavenging was calculated by the following equation:-

Scavenging effect (%) = {( $\alpha 0$ - $\alpha 1$ ) /  $\alpha 0$ } × 100

where  $\alpha 0$  and  $\alpha 1$  were the absorbance of control and in presence of sample.

# Determination of total antioxidant capacity (TAC)

The TAC was determined on the basis of reduction of Mo (VI) to Mo (V) by the antioxidant compound and the formation of green phosphate /Mo (V) complex at acidic pH. Total antioxidant capacity of the sample was investigated and compared against ascorbic acid. The TAC of the sample was determined by the assay prescribed by Preito et al  $(1999)^{29}$  with little modifications. A reaction mixture was prepared consisting of 0.3 ml of reagent solution (0.6 M H<sub>2</sub>SO<sub>4</sub>, 28 mM Na<sub>2</sub>SO<sub>4</sub>, 4 mM NH<sub>4</sub>Mo). Absorbance was measured at 695 nm after heating tubes at 95°C for 90 min. Ascorbic Acid was used to draw a standard curve and TAC was expressed as the equivalent of ascorbic Acid.

## Statistical analysis

Statistical analyses were done using MS Excel (Microsoft Office 2010 Professional).

# **RESULTS AND DISCUSSION**

### Extractive value and estimation of phytochemicals

The studied fraction had an extractive value of 5.58%. Phenol was present in the highest amount i.e.  $12.5 \pm 1.4 \mu g$  /mg of the sample. Flavonoid ( $1.78 \pm 0.36 \mu g$  /mg) and ascorbic acid ( $2.18 \pm 0.12 \mu g$  /mg) were present in moderate amounts whereas  $\beta$  carotene ( $0.0136 \pm 0.001\mu g$ /mg) and lycopene ( $0.011 \pm 0.007\mu g$  /mg) were present in trace amounts.

Phenolic compounds possess scavenging ability due to their hydroxyl groups and are known to be powerful antioxidant. In the present study, the total phenolic content of *S. commune* was found higher than *Pleurotus ostreatus*<sup>30</sup> and *Pleurotus citrinopileatus*<sup>31</sup> which were reported to be 5.49 µg/mg and 8.62 µg/mg respectively. The phenolic content was also found to be comparatively higher than that of *Russula albonigra* (9.53 µg/mg) as reported ealier<sup>32</sup>.

### **Reducing power**

Reducing power of a compound indicates its potential antioxidant activity. The reducers (i.e., antioxidants) reduces  $Fe^{3+}$  /ferricyanide complex to ferrous form. The yellow colour of the test solution is changed to various shades of green and blue, depending on the reducing power of the sample. *S. commune* was found to be a potential reducing agent, having an EC<sub>50</sub> value of 0.825 mg/ml (Fig 1). Compared to previously reported studies,



© Copyright protected. Unauthorised republication, reproduction, distribution, dissemination and copying of this document in whole or in part is strictly prohibited.

*Hypsizigus marmoreus*<sup>33</sup>, *Calocybe gambosa*<sup>34</sup>, *Tricholoma giganteum*<sup>35</sup>, *Russula albonigra*<sup>32</sup> *Amanita vaginata*<sup>36</sup> had slightly better reducing ability than *S. commune*. Thus, the sample has excellent reducing ability.

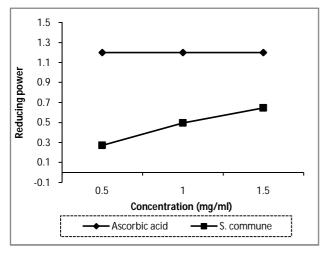
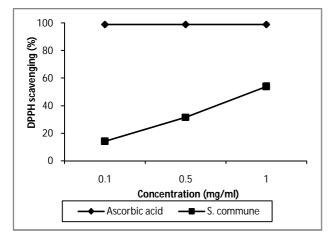


Figure 1: Reducing ability of ethanolic extract of *S. commune.* 

(Ascorbic acid was used as the standard. Results are mean  $\pm$  standard deviation of three separate experiments each in triplicate).

# DPPH radical scavenging assay

DPPH is a stable free radical that shows a characteristic absorbance at 517 nm, which decreases significantly when exposed to radical scavengers by providing hydrogen atom or electron to be a stable diamagnetic molecule. Ethanolic fraction of *S. commune* had an EC<sub>50</sub> value of 0.883 mg /ml (Fig 2) which indicates the potent DPPH radical scavenging potential of the extract. In comparison, the DPPH radical scavenging activities of *Russula albonigra*, *Pleurotus flabellatus* and *Amanita vaginata* were much lower than that of S. Commune. Expressed in the descending order, *S. commune* >*P. flabellatus*<sup>22</sup>> *A. vaginata*<sup>37</sup>>*R. albonigra*<sup>33</sup>. So, it is clear that the ethanolic extract of the sample has significant DPPH radical scavenging activity.



**Figure 2:** Reducing ability of ethanolic extract of *S. commune.* 

(Ascorbic acid was used as the standard. Results are mean  $\pm$  standard deviation of three separate experiments each in triplicate).

### Total antioxidant capacity

Total antioxidant capacity of ethanolic fraction of *S. commune* was determined by the formation of green phosphomolybdenum complex. The phosphomolybdenum method is based on the reduction of Mo (VI) to Mo (V) by the antioxidant compound and the formation of green phosphate/Mo (V) complex with the maximal absorption at 695 nm. Total antioxidant activity of the extract was estimated, using ascorbic acid as standard. On analysis, it was found that 0.1 mg of *S. commune* is as functional as approximately 67 ± 8 µg of ascorbic acid, expressed as 67 µg ascorbic acid equivalent (AAE).

## CONCLUSION

The data recorded in the above experiments showed that the ethanolic fraction of *Schizophyllum commune* has good antioxidant property and free radical scavenging power in different *in vitro* assays including Ferrous iron chelating, ferric iron reducing, DPPH free radical scavenging and total antioxidant activity, having a good amount of phenolics, flavonoids and ascorbic acid. Thus, it can be a good source of natural antioxidants for use as food additives.

Acknowledgement: The corresponding author (Prof. N. Irabanta Singh) wishes to thank Department of Biotechnology, Government of India for funding a coordinated project "Developing a digital database on Bioresources of N.E. India" (BT/29/NE/2011 dt 28 Nov. 2011) during this tenure the work was conducted.

## REFERENCES

- 1. Khatua S, Paul S, Acharya K, Mushroom as the Potential Source of New Generation of Antioxidant: A Review, Research Journal of Pharmacy and Technology, 6(5), 2013, 496-505.
- Ferreira ICFR, Barros L, Abreu RMV, Antioxidants in Wild Mushrooms, Current Medical Chemistry, 16, 2009, 1543-1560.
- Asatiani MD , Elisashvili V, Songulashvili G, Reznick AZ, Wasser SP, Higher Basidiomycetes Mushrooms as a Source of Antioxidants, Progress in Mycology, 2010, 311-326.
- Biswas G, Acharya K, Chemopreventive Activity of the Ethanolic Extract of Astraeus hygrometricus (Pers) Morg. on Ehrlich's Ascites Carcinoma Cell, Digest Journal of Nanomaterials and Biostructures, 7, 2012, 185-191.
- 5. Chatterjee S, Biswas G, Chandra S, Saha GK, Acharya K, Apoptogenetic Effects of *Tricholoma giganteum* on Ehrlich's Ascites Aarcinoma Cell, Bioprocess and Biosystems Engineering, 36, 2013, 101-107.
- Biswas G, Acharya K, Hypoglycemic Activity of Ethanolic Extract of Astraeus hygrometricus (Pers) Morg. In Alloxan-Induced Diabetic Mice, International Journal of Pharmacy and Pharmaceutical Sciences, 5(I1), 2013, 391-394.



Available online at www.globalresearchonline.net

© Copyright protected. Unauthorised republication, reproduction, distribution, dissemination and copying of this document in whole or in part is strictly prohibited.

- Chatterjee A, Khatua S, Chatterjee S, Paloi S, Mukherjee S, Mukherjee A, Acharya K, Bandyopadhyay SK, Polysaccharide-rich Fraction of *Termitomyces eurhizus* Accelerate Healing of Indomethacin Induced Gastric Ulcer in Mice, Glycoconjugate Journal, 30, 2013, 759-768.
- Chatterjee S, Dey A, Datta R, Dey S, Acharya K, Hepatoprotective Effect of the Ethanolic Extract of *Calocybe indica* on Mice with CCl<sub>4</sub> Hepatic Intoxication, International Journal of PharmTech Research, 3, 2011,2162-2168.
- 9. Biswas G, Sarkar S, Acharya K, Hepatoprotective Activity of the Ethanolic Extract of *Astraeus hygrometricus* (Pers.) Morg., Digest Journal of Nanomaterials and Biostructures, 6,2011,637-641.
- Acharya K, Chatterjee S, Biswas G, Chatterjee A, Saha GK, Hepatoprotective Effect of a Wild Edible Mushroom on Carbon Tetrachloride-Induced Hepatotoxicity in Mice, International Journal of Pharmacy and Pharmaceutical Sciences. 4 (3), 2012, 285-288.
- 11. Chatterjee S, Datta R, Dey A, Pradhan P, Acharya K, *In vivo* Hepatoprotective Activity of Ethanolic Extract of *Russula albonigra* against Carbon Tetrachloride-Induced Hepatotoxicity in Mice, Research Journal of Pharmacy and Technology, 5(8), 2012, 1034-1038.
- 12. Biswas G, Rana S, Sarkar S, Acharya K, Cardioprotective Activity of Ethanolic Extract of *Astraeus hygrometricus* (Pers.) Morg., Pharmacologyonline, 2, 2011,808-817.
- 13. Patra P, Bhanja SK, Sen IK, Nandi AK, Samanta S, Das D, SanjanaK, Devi P, Maiti TK, Acharya K, Islam SS, Structural and Immunological Studies of Hetero Polysaccharide Isolated from the Alkaline Extract of *Tricholoma crassum* (Berk.) Sacc., Carbohydrate Research, 362, 2012, 1-7.
- Nandi AK, Sen IK, Samanta S, Maity K, Sanjana K, Devi P, Mukherjee S, Maiti TK, Acharya K, Islam SS, Glucan from Hot Aqueous Extract of an Ectomycorrhizal edible Mushroom, *Russula albonigra* (Krombh.) Fr.: structural characterization and study of immunoenhancing properties, Carbohydrate Research, 363, 2012, 43-50.
- Nandi AK, Samanta S, Sen IK, Sanjana K, Devi P, Maiti TK, Acharya K, Islam SS, Structural Elucidation of an Immunoenhancing Heteroglycan Isolated from *Russula albonigra* (Krombh.) Fr., Carbohydrate Polymers, 94, 2013, 918–926.
- Lai TK,Biswas G, Chatterjee S, Dutta A, Pal C, Banerji J, Bhuvanesh N, Reibenspies JH, Acharya K, Leishmanicidal and Anticandidal Activity of Constituents of Indian Edible Mushroom Astraeus hygrometricus, Chemistry and Biodiversity, 9, 2012,1517-1524.
- 17. Giri S, Biswas G, Pradhan P, Mandal SC, Acharya K, Antimicrobial Activities of Basidiocarps of Wild Edible Mushrooms of West Bengal, India, International Journal of PharmTech Research,4(4), 2012, 1554-1560.
- Rai M, Sen S, Acharya K, Antimicrobial Activity of Four Wild Edible Mushrooms from Darjeeling Hills, West Bengal, India, International Journal of PharmTech Research, 5(3), 2013,949-956.
- 19. Mallick S, Dutta A, Dey S, Ghosh J, Mukherjee D, Sultana SS, Mandal S, Paloi S, Khatua S, Acharya K, Pal C, Selective Inhibition of *Leishmania donovani* by Active Extracts of

Wild Mushrooms Used by the Tribal Population of India: An *in vitro* Exploration for New Leads Against Parasitic Protozoans, Experimental Parasitology, 138, 2014, 9-17.

- Biswas G, Chatterjee S, Acharya K, Chemopreventive Activity of the Ethanolic Extract of Astraeus hygrometricus (Pers.) Morg. On Ehrlich's Ascites Carcinoma Cell, Digest Journal of Nanomaterials and Biostructures, 7, 2012,185-191.
- 21. Chatterjee S, Biswas G, Chandra S, Saha GK, Acharya K, Apoptogenic Effects of *Tricholoma giganteum* on Ehrlich's Ascites Carcinoma Cell, Bioprocess and Biosystems Engineering, 36, 2013, 101–107.
- 22. Dasgupta A, Rai M, Acharya K, Chemical Composition and Antioxidant Activity of a Wild Edible Mushroom *Pleurotus flabellatus*, International Journal of PharmTech Research, 5(4), 2013, 1655-1663.
- 23. Singleton VL, Rossi JA, Colorimetry of Total Phenolics with Phosphomolybdicphosphotungstic Acid Reagents, American Journal of Enology and Viticulture, 16, 1965, 144-158.
- Adebayo EA, Oloke JK, Ayandele AA, Adegunlola CO, Phytochemical, Antioxidant and Antimicrobial Assay of Mushroom Metabolite from *Pleurotus pulmonarius* –LAU 09 (JF736658), Journal of Microbiology and Biotechnology Research, 2(2), 2012, 366-374.
- 25. Nagata M, Yamashita I, Simple Method for Simultaneous Determination of Chlorophyll and Carotenoids in Tomato Fruit, Nippon Shokuhin Kogyo Gakkaishi, 39(10), 1992, 925-928.
- Rekha C, Poornima, G, Manasa M, Abhipsa V, Pavithra DJ, Vijay KHT, Prashith KTR, Ascorbic Acid, Total Phenol Content and Antioxidant Activity of Fresh Juices of Four Ripe and Unripe Citrus Fruits, Chemical Science Transactions, 1(2), 2012, 303-310.
- 27. Oyaizu M, Studies on Product of Browning Reaction Prepared from Glucose Amine, Japanese Journal of Nutrition, 44, 1986, 307-315.
- Shimada K, Fujikawa K, Yahara K, Nakamura T, Antioxidative Properties of Xanthan on the Autoxidation of Soybean Oil in Cyclodextrin Emulsion, Journal of Agricultural and Food Chemistry, 40(6), 1992, 945-948.
- 29. Prieto P, Pineda M, Aguilar M, Spectrophotometric quantitation of Antioxidant Capacity Through the Formation of a Phosphor Molybdenum Complex: Specific Application to the Determination of Vitamin E, Analytical Biochemistry, 269, 1999, 337-341.
- Jayakumar T, Thomas PA, Geraldine P, *In-vitro* Antioxidant Activities of an Ethanolic Extract of the Oyster Mushroom, *Pleurotus ostreatus*, Innovative Food Science and Emerging Technologies, 10(2), 2009, 228-234.
- Pal J, Ganguly S, Tahsin KS, Acharya K, *In vitro* Free Radical Scavenging Activity of Wild Edible Mushroom, *Pleurotus* squarrosulus (Mont.) Singer., Indian Journal of Experimental Biology, 48(12), 2010, 1210-1218.
- 32. Dasgupta A, Ray D, Chatterjee A, Roy A, Acharya K, *In vitro* Antioxidative Behaviour of Ethanolic Extract of *Russula albonigra*, Journal of Chemical and Pharmaceutical Research, 6(3), 2014, 1366-1372.



176

- 33. Lee YL, Yen MT, Mau JL, Antioxidant Properties of Various Extracts from *Hypsizigus marmoreus*, Food Chemistry,104, 2007, 1-9.
- Vaz AJ, Barros L, Martins A, Santos-Buelga C, Vasconcelos HM, Ferreira ICFR, Chemical Composition of Wild Edible Mushrooms and Antioxidant Properties of Their Water Soluble Polysaccharidic and Ethanolic Fractions, Food Chemistry, 126, 2011, 610-616.
- 35. Chatterjee S, Saha Gk, Acharya K, Antioxidant Activities of Extracts Obtained by Different Fractionation from *Tricholoma giganteum* Basidiocarps, Pharmacologyonline, 3, 2011, 88-97.
- Paloi S, Acharya K, Evaluation of Antioxidative Activity and Chemical Composition of Ethanolic Extract from *Amanita vaginata* (Bull.) Lam.: an *in vitro* Study, Asian Journal of Pharmaceutical and Clinical Research, 7(2), 2014, 88-92.

## Source of Support: Nil, Conflict of Interest: None.

