Research Article



Potential Role of Curcumin and Garlic Acid against Diazinon and Propoxur Hepatotoxicity

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ABSTRACT

Diazinon, an organophosphorus insecticide and propoxur, a carbamate insecticide have been used in agriculture and public health for several years. The aim of the present study was to evaluate the oxidative stress caused by the two insecticides and biochemical changes in adult male Wistar rats and to evaluate the protective effect of curcumin and garlic. Diazinon (10 mg/kg per day in corn oil), propoxur (10 mg/kg per day in corn oil) alone or with curcumin (100 mg/kg per day in corn oil) and/or garlic (20 mg/kg per day in distilled water) were given to rats (n=11) orally through gavage for four weeks. Biochemical parameters in serum [total protein, albumin, triglyceride (TG), cholesterol, uric acid, urea, creatinine, γ-glutamyl transpeptidase (γ-GT), glutamic pyruvic transaminase (GPT), glutamic oxaloacetic transaminase (GOT), lactate dehydrogenase (LDH), catalase (CAT) and total antioxidant capacity (TAC)], superoxide dismutase (SOD) and glutathion reduced (GSH) were determined in liver homogenate and malondialdehyde (MAD) was determined in RBCs. All ultrastructural changes were investigated at the end of 4th week. Results obtained showed that albumin, total protein, GOT, GPT, LDH, creatinine, urea, GSH and SOD were statistically high significance (p<0.001), γ-GT, uric acid, CAT, TAC and cholesterol were statistically significance (p<0.01) when diazinon or propoxur-treated groups compared to control group. On the other hand, GOT, GPT, LDH, GSH and SOD were statistically high significance (p<0.001), albumin, total protein, γ-GT, urea, CAT and TAC were statistically significance (p<0.01) when curcumin and/or garlic + diazinon or propoxur-treated groups compared to diazinon or propoxur-treated groups. We conclude that curcumin and garlic decreases diazinon and propoxur oxidative toxic effects and hepatotoxicity.

Keywords: curcumin, garlic acid, diazinon, propoxur, hepatotoxicity

INTRODUCTION

he agricultural chemicals commonly labeled, as pesticides are perhaps the largest group of poisonous substances being disseminated throughout our environment¹. The contamination of food, water and air with these pollutants has become imminent, and consequently adverse health effects are inevitable in humans, animals, wildlife and fish².

Carbamates compounds such as propoxur have a common mechanism of action toward insect pests and unintended toxicity to non target organisms including humans, that is, acetvlcholinesterase (AChE) inhibition by carbamylating the serine hydroxyl group in the active site of the enzyme in the nervous system, leading to the persistent action of the neurotransmitter, acetylcholine, on cholinergic postsynaptic receptors³⁻⁵. Propoxur (2isopropoxyphenyl N-methyl carbamate) is a widely used broad spectrum insecticide. In addition to the control of cockroaches, mosquitoes, bugs, fleas, ants, millipedes, this insecticide is also used against pests in food stores, open areas, and households. Propoxur exhibits a toxic effect characterized by the inhibition of the enzyme cholinesterase⁶. Although mildly toxic to humans and domestic animals (class II), propoxur is highly toxic to birds and fish and cannot be used in the later. Propoxur is highly toxic to honeybees⁷. Several cases of suicidal and occupational poisoning have also been reported⁸. Hence, propoxur has generated considerable concern regarding its subtle health effects. Possible effect of persistent exposure to propoxur on oxidative stress and functional integrity of the immune system has heightened interest in these parameters as additional indices to analyze potential long term health effects⁹. Recent studies on animals and human from our laboratory suggest that propoxur can cause oxidative stress and immunotoxicity¹⁰.

(*p* -diethyl-*o*-[2-isopropyl-6-methyl-4-Diazinon pyrimidinyl] phosphoro-thioate) is an organophosphorus pesticide insecticide with a broad range of activities which inhibit acetyl-cholinesterase activity. It has been widely and effectively used throughout the world with applications in agriculture and horticulture for controlling insects in crops, ornamentals, lawns, fruit, vegetables and other food products¹¹. Some reports have been published with respect to Diazinon and its effects on haematological and biochemical parameters of rat, rabbits and mice¹². Many insecticides such as Diazinon are hydrophobic molecules, which bind extensively to biological membranes, especially to the phospholipids bilayers. Diazinon may interfere with lipid metabolism in mammalian animals that included the levels of total cholesterol, high-density lipoprotein cholesterol, lowdensity lipoprotein cholesterol, triglycerides and phospholipids and its effect was dose-dependent¹³. Diazinon's mutagenicity studies, its ability to cause genetic damage, showed that Diazinon in fact can



damage DNA in human blood cells, in cells from laboratory animals, and bacteria¹⁴. Diazinon exposure was found to increase the occurrence of a type of genetic damage called micronuclei. Micronuclei may be induced by strand breaks in DNA due to oxidative stress¹⁵.

Plant products are known to exert their protective effects by scavenging free radicals and modulating antioxidant defense system. Curcumin (diferuloymethane), a yellow orange dye derived from the rhizomes of Curcuma **b**

in , is used as a spice and food-coloring agent in cooking^{16,17}. Curcumin represents a class of antiinflammatory and antioxidants reported to be a potent inhibitor of reactive oxygen species formation^{18,19}. Reddy and Lokesh indicated that Curcumin is a potent scavenger of a variety of reactive oxygen species including superoxide radicals and hydroxyl radicals²⁰.

Garlic, **B** , is a member of the lily family that has been cultivated by humans as a food plant over 10,000 years. Ancient Egyptian records mentioned that use of garlic as a remedy for a variety of diseases²¹. Recently, it has been found that the sulfur-containing compounds of garlic have anti-mutagenesis and anticarcinogenesis effects. In vivo studies show that garlic and its associated sulfur compound. So, the aim of this study was to evaluate the oxidative toxic effects of the organophosphorus pesticide diazinon and the carbamate pesticide propoxur and to investigate the protective effect of curcumin and garlic.

MATERIALS AND METHODS

Animals

Male Wistar albino rats, weighing 100-150 gm were obtained from the animal house of the Faculty of Pharmaceutical Science of Mansoura University. The animals were fed a standard laboratory diet and water dbn . Rats were kept 12 h light 12 h dark cycles at a room temperature of 25°C at least two days prior testing. All animal experiments were approved by the Animal House of Biochemistry branch of Chemistry Department

Chemicals

Animal Treatment Schedule

Rats were divided into 13 groups (n=11). The compounds were administered in the morning (between 10:00 and 12:00 AM) to non-fasted rats. All rats were treated for 4 weeks.

Group 1: control group

Rats fed a standard laboratory diet and tap water \boldsymbol{a}

Group 2: corn oil group

Rats received corn oil through gavages daily.

of Faculty of Science of Mansoura University.

Group 3: diazinon-treated group

Rats received Diazinon at dose of (10 mg/kg bw/day) in

corn oil through gavages.

Group 4: diazinon+curcumin-treated group

Rats received Diazinon at dose of (10 mg/kg bw/day) in corn oil and Curcumin (100 mg/kg bw/day) in corn oil both through gavages.

Group 5: diazinon+garlic-treated group

Rats received Diazinon at dose of (10 mg/kg bw/day) in corn oil and Garlic (20 mg/kg bw/day) in distilled water both through gavages.

Group 6: diazinon+curcumin+garlic-treated group

Rats received Diazinon at dose of (10 mg/kg bw/day) in corn oil, Curcumin (100 mg/kg bw/day) in corn oil and Garlic (20 mg/kg bw/day) in distilled water all through gavages.

Group 7: propoxur-treated group

Rats received Propoxur at dose of (10 mg/kg bw/day) in corn oil through gavages.

Group 8: propoxur+curcumin-treated group

Rats received Propoxur at dose of (10 mg/kg bw/day) in corn oil and Curcumin (100 mg/kg bw/day) in corn oil both through gavages.

Group 9: propoxur+garlic-treated group

Rats received Propoxur at dose of (10 mg/kg bw/day) in corn oil and Garlic (20 mg/kg bw/day) in distilled water both through gavages.

Group 10: propoxur+curcumin+garlic-treated group

Rats received Garlic at dose of (20 mg/kg bw/day) in distilled water through gavages.

Group 11: garlic-treated group

Rats received Curcumin at dose of (100 mg/kg bw/day) in corn oil through gavages.

Group 12: curcumin+garlic-treated group

Rats received Curcumin at dose of (100 mg/kg bw/day) in corn oil and Garlic (20 mg/kg bw/day) in distilled water both through gavages.

Collection of Samples

Collection of Blood Samples

Blood samples were collected from all groups by cardiac puncture into heparin-coated and dry tubes.

The blood samples collected were centrifuged at 3000 rpm for 10 min for the separation of plasma and sera.

The lower erythrocyte layer in the heparinised tubes was washed three times with phosphate buffered saline and diluted with an equal volume of the indicated solution.

Next the erythrocytes were hemolysed with ice-cold distilled water (1:5).



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Collection of Liver Samples

Livers were excised, washed with deionized water for the removal of blood, and later fatty parts were removed. Livers were rinsed with isotonic saline and dried by blotting between 2 pieces of filter paper. Livers were stored at -20°C in plastic vials containing a 0.5 ml of ice-cold sterile isotonic saline.

Preparation of Homogenates

An accurately weighed piece of liver tissue was homogenized in an ice-cold phosphate buffer solution with a PH value adjusted to 7.4 using a Teflon Pestle connected to a Braun Homogenizer motor (25 strokes per minute at 1000 rev/min), then the liver homogenate was diluted to yield a 5% (w/v) liver homogenate. The homogenate was centrifuged at 5000 rpm for 30 minutes at 4°C to remove cell debris and nuclei. The resulting supernatant was used for biochemical analysis.

Biochemical and Specific Biomarker Evaluation

Biodiagnostic label kits were used for the determination of serum total protein, albumin, triglyceride (TG), cholesterol, uric acid, urea, creatinine, γ -glutamyl transpeptidase (γ -GT), glutamic pyruvic transaminase (GPT), glutamic oxaloacetic transaminase (GOT), lactate dehydrogenase (LDH), catalase (CAT) and total antioxidant capacity (TAC). Superoxide dismutase (SOD) and glutathion reduced (GSH) were determined in liver homogenate. Malondialdehyde (MAD) was determined in RBCs²². were analyzed by SPSS 19 to calculate the significance. P<0.01 value between study groups was taken as statistically significant and P<0.001 value was taken as statistically high significant.

RESULTS

Table 1 showed significant decreased levels of serum albumin, total protein and urea in rats treated with either diazinon or propoxur than that non-treated. However, levels increased significantly in rats treated with curcumin and/or garlic before pesticides treatment. On the other hand, serum γ -glutamyl transpeptidase, glutamic pyruvic transaminase (GPT), glutamic oxaloacetic transaminase (GOT) and lactate dehydrogenase (LDH) levels showed significant increase in rats treated with either diazinon or propoxur than normal rats, but these levels became like normal by curcumin and/or garlic pretreatment. Uric acid and creatinine showed elevated levels in pesticides treated rats, but with the curcumin and/or garlic pretreatment the levels became near levels of normal rats.

Table 2 showed significant increased levels in serum (TG) and cholesterol trialycerides and RBCs malondialdehyde (MAD) in rats treated with pesticides only compared to normal rats, but these levels decreased in rats pretreated with curcumin and/or garlic compared to pesticides treated rats. On the other way, table 2 showed significant decreased levels of tissue glutathione (GSH) and superoxide dismutase and serum catalase (CAT) and total antioxidant capacity (TAC) in rats treated with diazinon or propoxur. These levels showed significant elevations in rats treated with curcumin and/or garlic before pesticides treatment.

Statistical Analysis

The mean ± SEM were determined for each group. Data

Table 1: Effect of diazinon, diazinon + curcumin and/or garlic, propoxur and propoxur + curcumin and/or garlic on liver and kidney function enzymes.

Parameter	Albumin g/dl	Protein g/dl	γ-GT IU/L	GOT IU/L	GPT IU/L	LDH IU/L	Uric acid mg/dl	Creatinine mg/dl	Urea mg/dl
Control	3.0±0.1	7.2±0.2	38.9±3.8	18.1±0.4	15.0±0.6	272.2±14.3	3.9±0.4	0.7 ± 0.03	39.8 ± 2.04
Corn oil	2.9±0.2	7.2±0.3	29.9±3.1	32.8±0.9	26.0±0.5ii	272.0±12.1	4.1±0.5	0.7±0.03	33.0±4.05
Diazinon (DZN) 10mg/kg	1.9±0.2ii	5.2±0.3ii	70.1±7.3i	58.2±2.5ii	44.7±0.5ii	811.5±10.2ii	5.4±0.4i	1.1±0.07ii	16.0±1.9ii
Curcumin+D ZN	2.7±0.2*	6.4±0.4*	34.9±6.3*	43.1±1.0ii**	33.9±1.2ii**	455.9±13.9ii**	4.2±0.4	0.9±0.1*	23.4±2.5ii* *
Garlic+DZN	2.5±0.1i*	6.8±0.1**	45.9±2.2*	42.0±0.7ii**	34.0±1.0ii**	418.4±17.9ii**	4.2±0.5	0.9±0.1*	23.7±1.5ii*
Curcumin+ga rlic+DZN	2.2±0.1ii	7.0±0.4*	28.3±3.2*	42.4±1.1ii**	34.8±1.2ii**	457.7±18.6ii**	4.7±0.3	0.9±0.1	27.3±4.9i*
Propoxur (PPX) 10mg/kg	2.1±0.1ii	5.0±0.2ii	60.9±4.1i	75.1±1.2ii	53.0±0.6ii	918.9±20.9ii	5.7±0.5i	1.1±0.06ii	22.9±2.08ii
Curcumin+P PX	2.9±0.3*	6.5±0.3*	36.4±3.4**	45.8±0.4ii**	40.4±0.3ii**	647.4±17.8ii**	5.1±0.4i	0.9±0.1*	23.5±1.7ii
Garlic+PPX	3.0±0.2**	6.2±0.4i*	29.3±2.3**	45.0±1.1ii**	39.0±1.8ii**	589.6±19.2ii**	3.4±0.3*	1.03±0.1i	22.8±3.0ii
Curcumin+ga rlic+PPX	2.8±0.2*	4.9±0.5ii	34.3±3.1**	46.6±1.0ii**	41.2±1.5ii**	585.4±21.9ii**	4.1±0.4*	0.9±0.11i	21.9±3.8ii
Garlic	2.9±0.2	7.2±0.4	33.1±2.3	31.5±0.9i	23.2±0.8ii	597.6±15.6ii	4.2±0.2	0.8±0.04	24.7±1.7ii
Curcumin	2.8±0.2	5.3±0.3ii	26.1±0.95i	33.1±0.6i	28.0±0.8ii	628.0±15.4ii	4.4±0.3	0.8±0.02	23.8±1.8ii
Garlic + Curcumin	3.2±0.4	6.2±0.7	27.5±0.95i	42.9±0.6i	29.4±0.8ii	538.0±8.4ii	4.9±0.5	0.8±0.05	23.3±3.8i

Data showed as mean \pm standard error of the mean (SEM), i= Significant and ii= highly significant compared with those of the control and *= Significant and **= highly significant compared with those of pesticide treated only.



Available online at www.globalresearchonline.net © Copyright protected. Unauthorised republication, reproduction, distribution, dissemination and copying of this document in whole or in part is strictly prohibited. **Table 2:** Effect of diazinon, diazinon + curcumin and/or garlic, propoxur and propoxur + curcumin and/or garlic on biochemical indices levels in rats.

Parameter	TG mg/dl	Cholesterol mg/dl	GSH mg/gm tissue	SOD IU/gm tissue	CAT IU/L	TAC mmol/L	MAD (× 10 ⁻⁵) mmol/1ml packed cells
Control	76.9±10.6	68.1 ± 5.67	68.4±3.7	682.7±11.0	686.9±83.3	0.54±0.02	0.68±0.16
Corn oil	66.7±7.6	76.9±11.91	64.1±5.5	691.0±13.0	629.9±87.1	0.53±0.02	0.48±0.08
Diazinon (DZN) 10mg/kg	90.6±7.0	94.0±7.0i	42.8±0.9ii	393.8±9.1ii	474.9±54.4i	0.30±0.09i	1.10±0.21
Curcumin+DZN	85.7±3.1	101.1±15.3	52.9±0.3ii**	630.7±7.0i**	789.0±68.8*	0.52±0.02*	0.45±0.11*
Garlic+DZN	85.5±6.8	75.2±9.04	58.2±0.4i**	611.3±4.9ii**	710.2±79.0*	0.52±0.03*	0.82±0.08
Curcumin+garlic+DZN	95.4±9.0	49.2±5.7**	52.3±0.3i**	576.5±8.6ii**	482.1±65.6	0.34±0.03ii	0.76±0.21
Propoxur (PPX) 10mg/kg	94.0±5.4	95.7±5.6i	35.0±0.8ii	448.3±4.9ii	488.0±53.7	0.22±0.05ii	1.08±0.22
Curcumin+PPX	85.5±3.7	83.8±7.3	52.4±0.6ii**	687.9±8.4**	688.3±75.9*	0.47±0.05*	0.54±0.07*
Garlic+PPX	89.7±2.6	89.7±7.4	56.1±0.5i**	618.8±7.3ii**	612.0±82.5	0.36±0.07i	0.22±0.04i*
Curcumin+garlic+PPX	89.2±9.0	70.8±7.8*	55.4±0.3i**	596.3±6.9ii**	535.6±76.7	0.38±0.06i	0.83±0.07
Garlic	92.3±2.9	99.3±6.0i	61.0±0.4i	721.8±6.7i	609.6±22.2	0.44±0.03i	0.52±0.08
Curcumin	105.8±4.5i	94.2±6.1i	59.7±0.6i	718.8±8.5i	411.9±49.9i	0.52±0.03	0.53±0.11
Garlic + Curcumin	90.4±6.1	94.2±6.1i	61.3±0.7	722.3±8.5i	617.8 ±53.0	0.50±0.03	0.72±0.06

Data showed as mean \pm standard error of the mean (SEM), i= Significant and ii= highly significant compared with those of the control and *= Significant and **= highly significant compared with those of pesticide treated only.

DISCUSSION

At present there is considerable interest in free radicalmediated damage in biological systems following pesticide exposure.

However, there is no consensus with regard to the best quantitative indices of pesticide-induced oxidative stress and the effect of antioxidant interventions.

Diazinon not only used in control of vegetables and fruits but also in ectoparasiticide formulations for sheep and cattle and in collars and washes for external asitic control.

Diazinon not only has toxic effect orally, it also has toxic effects by dermal and inhalation way¹¹.

On the other hand, carbamate insecticides such as propoxur cause oxidative stress through the generation of free radicals and changes in antioxidant enzymes and oxygen-free radical scavengers. Lipid peroxidation is known to be one of the molecular mechanisms of carbamate-induced toxicity¹⁰.

There is evidence that curcumin enhances liver detoxification by increasing the activity of glutathione S-transferase, an enzyme which conjugates glutathione with a wide variety of toxins to facilitate their removal from body²³. It has the protective effect against pesticide induced biochemical alterations and oxidative damage in the various organs of rats. This protective effect is due to its free radical scavenging activity and increased antioxidant enzymes in rats²⁴. Garlic has ability to reduce free radical-induced oxidative damage in the liver²⁵, garlic extract has been shown to decrease liver enzymes in serum and prevent liver damage of rats with liver fibrosis²⁶.

Treated rats with diazinon or propoxur showed significant increase in yGT, GOT, GPT and LDH and significant decrease in total protein and albumin. All these biochemical indices including yGT, GOT, GPT, LDH, total protein and albumin are mainly monitored for the elevation of liver damage. Albumin is the main protein in blood and is made by the liver. Changes in serum albumin content indicated either increased albumin synthesis by the tissues or decreased albumin degradation within each tissue. The probability of albumin synthesis by the various rat tissues contributing to an increase in serum albumin content can be considered as more realistic. Hepatotoxicity leads to decrease in albumin production²⁷. Nevertheless, decreased content of protein in serum had also been reported²⁸. The estimation of total proteins in the body is helpful in differentiating between a normal and damaged liver function as the majority of plasma proteins like albumins and globulins are produced in the liver²⁷. y-Glutamyl transferase (yGT) is an enzyme which is found in liver, kidney and pancreatic tissues, the enzyme concentration being low in liver as compared to kidney²⁹. y-Glutamyl transferase catalyzed the transfer of a glutamyl moiety between peptide donors and amino acid/peptide acceptors³⁰. γ-Glutamyl transferase was also involved in the transfer of amino acid across the cell membrane. Further, y-glutamyl transferase had a role in glutathione metabolism, transferring the glutamyl moiety to various acceptor molecule including water, L-amino acids and peptides. Such a process results in the retention of the cysteinyl glycine that was considered to preserve intracellular homoeostasis during oxidative stress³¹. Glutamic pyruvic transaminase (GPT) is an enzyme that helps metabolize protein. When the liver is damaged, GPT is increased in liver and released in the bloodstream, so



high level of this enzyme is observed. The estimation of this enzyme is a more specific test for detecting liver abnormalities since it is primarily found in the liver³²⁻³⁴, also this enzyme showed elevated levels during hepatocellular necrosis. Glutamic oxaloacetate transaminase (GOT) is another liver enzyme that aids in producing proteins. It catalyzes the reductive transfer of an amino group from aspartate to α -ketoglutarate to yield oxaloacetate and glutamate. Glutamic oxaloacetic transaminase is the mitochondrial enzvme, predominantly found in the liver, skeletal muscles and kidneys. Injury to any of these tissues can cause an elevated blood level³⁵. It also helps in detecting hepatocellular necrosis but is considered a less specific biomarker enzyme for hepatocellular injury²⁹ as it can also signify abnormalities in heart, muscle, brain or kidney^{33,34}. The ratio of serum GOT to GPT can be used to differentiate liver damage from other organ damage³⁵. Lactate dehydrogenase (LDH) can be used as an indicator for cellular damage and cytotoxicity of toxic agents. In fact, elevation in lactate dehydrogenase (LDH) activity indicates cell lysis and death as well as the switching over of anaerobic glycolysis to aerobic respiration³⁶. The change in lactate dehydrogenase (LDH) activity resulted from overproduction of superoxide anions and hydroxyl radicals, which cause oxidative damage to the cell membrane³⁷ and increase in membrane permeability³⁸. Elevated level of this enzyme is released from damaged cells in many areas of the body, including the liver. It also helps in detecting hepatocellular necrosis²⁷.

Rats treated with either diazinon or propoxur showed slight increase of creatinine and uric acid in blood than normal rats. So the results of creatinine and uric acid showed no significantly disorder in kidney. On other hand, Urea levels were observed to be significantly decreased in rats treated with diazinon or propoxur than non-treated rats. Rats treated with curcumin and/or garlic before pesticides treatment show slight decrease in serum urea levels than normal rats. Low levels of urea in blood showed that there is disorder in urea cycle in which liver is responsible for. Low blood urea nitrogen levels are not usually a cause for concern. They may be seen in severe liver disease and over hydrated, but blood urea nitrogen levels is not usually used to diagnose for liver disease or over degredation. However, Rats treated with diazinon or propoxur had high levels of glutamic pyruvic transaminase (GPT), glutamic oxaloacetic transaminase (GOT), y-Glutamyl transferase (yGT) and Lactate dehydrogenase (LDH) specific activities and low levels of albumin and total protein. These results emphasizes that the depletion of urea blood nitrogen levels in these rats is due to disorder in urea cycle, which in turn due to liver damage or liver dysfunction. Blood urea results of rats treated with curcumin and/or garlic indicate the protective effect of curcumin and garlic against toxic effects of diazinon and propoxur as organophophorus and carbamate pesticide respectively.

Superoxide dismutase (SOD) enzyme activity in liver tissue showed decreased levels in rats treated with diazinon or propoxur than non-treated rats. Also, liver glutathione reduced (GSH) showed the same decreased levels in pesticides treated rats. Catalase and total antioxidant capacity (TAC) showed decreased levels in rats treated with diazinon or propoxur than normal rats. All these results indicate the potential of diazinon and propoxur to induce oxidative stress. Oxidative stress can cause irreversible cellular damage because intracellular defense mechanisms are depleted and therefore cannot protect cells against reactive oxygen species (ROS), which are a group of highly reactive zmolecules that are produced through sequential reductions of O_2^{39} . There is a balance between ROS generation and ROS degredation with antioxidants in the cells. Both excess of ROS or antioxidants can result in an abnormal oxidative stress state⁴⁰. Most chemicals such as hydrogen peroxide (H_2O_2) damage liver cells mainly by including lipid peroxidation and oxidative stress in liver⁴¹. The cells have different mechanism to alleviate and repaired damaged macromolecules. The primary defense is offered by enzymatic and non enzymatic antioxidants which have been shown to scavenge free radicals and reactive oxygen species (ROS). The antioxidant enzymes, SOD and CAT have been shown to be significantly affected by pesticides⁴². Increased levels of serum GOT, GPT, y-GT and LDH and decreased levels in albumin and total protein of treated rats with pesticides prove that the livers of rats are damaged. However, it is not evidence that liver damage is due to oxidative stress caused by pesticides. So, we needed other specific markers to prove the oxidative stress mechanism in liver damage after pesticides treatment. Hepatic levels GSH and SOD estimated in liver homogenate to confirm that the oxidative stress is the main cause of the liver dysfunction. If we looked to the rest of hepatic GSH and SOD results, we would find the role of the two used antioxidants, curcumin and garlic, in liver protection. All these results emphasize that diazinon and propoxur induce oxidative stress in rats' body. As the liver is the first defense organ in body which protects the body from toxicants through hepatic biotransformation, involve phase I and phase II, it is the first organ to be affected.

Result data showed increased levels of cholesterol and triglycerides in rats treated with diazinon or propoxur. Cholesterol and triglycerides levels were considered as valuable indicator of drug-induced disruption of lipid metabolism. Increase of cholesterol and triglycerides levels in rats suggest increased synthesis and accumulation of cholesterol and trialycerides. Accumulation of pesticides in the liver is reported to disrupt lipid metabolism and increase serum cholesterol and triglycerides⁴³. This disruption may be due to decreased lipoprotein lipase activity in adipose tissue and increased the levels of total serum cholesterol and triglycerides in the affected rats⁴⁴.



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Red blood cells are highly susceptible to oxidative damage due to the high cell concentration of oxygen and hemoglobin, a powerful promoter of the oxidative process. They are one of the first cells to be affected by adverse conditions. RBCs have a plasma membrane rich in polyunsaturated fatty acid chains. Thus they are highly susceptible to oxidation. Lipids are considered crucial in the maintenance of the red blood cells shape. Even minimal changes in the surface area may lead to morphological and functional abnormalities. Red cell membrane fluidity is also important for proper red cell function. Reactive oxygen species attack causes lipid peroxidation and formation of an array of unwanted products⁴⁵. Malondialdehyde (MDA) is a major lipid peroxidation product. Several hypotheses in relation to formation of MDA have been proposed. It was the *ii*ø proposed that oxidized lipids are able to produce MDA as a decomposition product and the mechanism is thought formation involve of prostaglandins, like endoperoxides from poly unsaturated fatty acid with two or more double bonds⁴⁶. Also, it was suggested an alternative mechanism for the generation of MDA, based on successive hydroperoxide formation and β -cleavage of poly unsaturated fatty acid which is the main source of MDA generation in

However other minor sources of MDA formation also exist such as byproducts of free radical generation by ionizing radiation and biosynthesis of prostaglandins⁴⁸.

Curcumin and garlic results showed protective effect against pesticides oxidative stress. Curcumin was suggested to exert protective effect by modulating the biochemical marker enzymes, lipid peroxidation and augmenting antioxidant defense system⁴⁹. A protective effect of curcumin has also been reported in *kip* experiments against cadmium and lead induced lipid peroxidation in rat brain homogenates⁵⁰. Aqueous garlic extract effectively reduces the combination pesticide induced oxidative damage as shown by a decrease in lipid peroxidation and enhanced the antioxidant levels in liver of mice⁵¹. Several plant products, including garlic *bn* components have been reported to modulate the levels of lipid peroxides and antioxidants⁵².

CONCLUSION

From the present results, it can be concluded that exposure of animals to diazinon or propoxur are capable of including marked hazardous alterations. The generation of excessive levels of free radicals is one of the basic underlying mechanisms of these changes. Changes observed in oxidative stress markers further support this. The result also demonstrated the protective effect of curcumin and garlic against pesticide oxidative stress toxicity.

REFERENCES

- 1. Ecobichon DJ, and Joy RM, Pesticides and Neurological Diseases, in CRC Press, Boston, 1994, p. 1-23.
- 2. Gupta RC, Carbofuran toxicity, J. Toxicol. Environ. Health, 43(4), 1994, 383–418.
- Baron RL, Carbamate insecticides. In "Handbook of Pesticide Toxicology" Academic Press, New York, 1991, 1125–1189.
- Ecobichon DJ, Toxic Effects of Pesticides Chapter 18. In "Casarett and Doull's Toxicology: The Basic Science of Poisons" (M. O. Amdur, J. Doull, and C. D. Klaassen, eds.) Pergamon Press, New York, NY, 4th ed, 1991, 565–623.
- Knaak JB, Dary CC, Okino MS, Power FW, Zhang X, Thompson CB, Tornero-Velez R, Blancato J, Parameters for carbamate pesticide QSAR and PBPK/PD models for human risk assessment. Rev. Environ. Contam. Toxicol., 193, 2008, 53–210.
- Shukla Y, Baqar SM, Mehrotra NK, Carcinogenicity and cocarcinogenicity studies on propoxur in mouse skin, Food Chem. Toxicol., 36, 1998, 1125-1130.
- 7. Kaya S, Insektisitler. In: Veteriner Hekimliginde Toksikoloji. Medisan Yayinevi, Ankara, 2002, 401-454.
- 8. Banerjee BD, Pasha ST, Hussain QZ, Koner BC, Ray A, A comparative evaluation of immunotoxicity of Malathion after subchronic exposure in experimental animals. Indian J Exp Biol., 36(3), 1998, 273-282.
- Banerjee BD, The influence of various factors on immune toxicity assessment of pesticide chemicals, Toxicol. Lett., 107(1-3), 1999, 21-31.
- Seth V, Banerjee BD, Chakravorty AK, Lipid Peroxidation, Free Radical Scavenging Enzymes, and Glutathione Redox System in Blood of Rats Exposed to Propoxur. Pesticide Biochemistry and Physiology, 71, 2001, 133-139.
- 11. Grafitt SJ, Jones K, Mason HJ, Cocker J, Exposure to the organophosphate diazinon: data from a human volunteer study with oral and dermal doses, Toxicol Lett., 134(1-3), 2002, 105-113.
- Kalender S, Ogutcu A, Uzunhisarcikli M, Açikgoz F, Durak D, Ulusoy Y, Kalender Y, Diazinon-induced hepatotoxicity and protective effect of vitamin E on some biochemical indices and ultrastructural changes, Toxicology, 211(3), 2005, 197-206.
- 13. Ibrahim NA, EI-Gamal BA, Effect of diazinon, an organophosphate insecticide, on plasma lipid constituents in experimental animals, J. Biochem Mol Biol., 36(5), 2003, 499-504.
- Grover P, Danadevi K, Mahboob M, Rozati R, Banu BS, Rahman MF, Evaluation of genetic damage in workers employed in pesticide production utilizing the Comet assay, Mutagenesis, 18(2), 2003, 201-205.
- Fenech M, The cytokinesis-block micronucleus technique: a detailed description of the method and its application to genotoxicity studies in human populations, Mutat. Res., 285(1), 1993, 35-44.



Available online at www.globalresearchonline.net

- 16. Joe B, Vijaykumar M, Lokesh BR, Biological properties of curcumin-cellular and molecular mechanisms of action, Crit. Rev. Food Sci. Nutr., 44(2), 2004, 97-111.
- 17. Maheshwari RK, Singh AK, Gaddipati J, Srimal RC, Multiple biological activities of curcumin: a short review, Life Sci., 78(18), 2006, 2081-2087.
- Biswas SK, McClure D, Jimenez LA, Megson IL, Rahman I, Curcumin induces glutathione biosynthesis and inhibits NFkappaB activation and interleukin-8 release in alveolar epithelial cells: mechanism of free radical scavenging activity, Antioxid Redox Signal., 7(1-2), 2005, 32-41.
- 19. Venkatesan N, Punithavathi D, Arumugam V, Curcumin prevents adriamycin nephrotoxicity in rats, Br. J. Pharmacol., 129(2), 2000, 231-234.
- 20. Reddy AC, Lokesh BR, Studies on the inhibitory effects of curcumin and eugenol on the formation of reactive oxygen species and the oxidation of ferrous iron, Mol. Cell Biochem., 137(1), 1994, 1-8.
- 21. Block E, The chemistry of garlic and onions, Sci. Am., 252(3), 1985, 114-119.
- 22. Stocks J, Donnandy T, The autoxidation of human red cell lipids induced by hydrogen peroxide, Br. J. Haematol., 20, 1971, 95-111.
- 23. Piper JT, Singhal SS, Salameh MS, Torman RT, Awasthi YC, Awasthi S, Mechanism of anticarcinogenic properties of curcumin on glutathione linked detoxification enzymes in rat liver, Int. J. Biochem. Cell Biol., 30, 1998, 445-456.
- 24. Sankar P, Telang AG, Manimaran A, Protective effect of curcumin on cypermethrin-induced oxidative stress in Wistar rats, Exp. Toxicol. Pathol., 64(5), 2012, 487-493.
- 25. Gedik N, Kabasakal L, Sehirli O, Ercan F, Sirvanci S, Keyer-Uysal M, Sener G, Long-term administration of aqueous garlic extract (AGE) alleviates liver fibrosis and oxidative damage induced by biliary obstruction in rats, Life Sci., 76(22), 2005, 2593-2606.
- 26. Nakagawat S, Kasuga S, Matsuura H, Prevention of liver damage by aged garlic extract and its components in mice, Phytotherapy Research, 3(2), 1989, 50-53.
- 27. Thapa BR, Walia A, Liver function tests and their interpretation, Indian J. Pediatr., 74, 2007, 663-671.
- 28. Lakkawar AW, Chattopadhyay SK, Somvanshi R, Experimental cypermethrin toxicity in rabbits-a clinical and pathoanatomical study, Folia. Veterinaria, 48, 2004, 3-8.
- 29. Ozer J, Ratner M, Shaw M, Bailey W, Schomaker S, et al., The current state of serum biomarkers of hepatotoxicity, Toxicology, 245, 2008, 194-205.
- 30. Meister A, Tate SS, Ross LL, Membrane Bound Gamma Glutamyl Transpeptidase: In the Enzyme of Biological Membranes, Martonosi A: Plenum Press., 1973, 315–347.
- 31. Shukla Y, Antonym M, Mehrota NK, Studies on c-Glutamyl transpeptidase in rodents exposed to benomyl, Bull. Environ. Contam. Toxicol., 42, 1989, 301-306.
- 32. Amacher DE, A toxicologist's guide to biomarkers of hepatic response, Hum. Exp. Toxicol., 21, 2002, 253-262.

- 33. Dufour DR, Lott JA, Nolte FS, Gretch DR, Koff RS, et al., Diagnosis and monitoring of hepatic injury: I. Performance characteristics of laboratory tests, Clin. Chem., 46, 2000, 2027-2049.
- Dufour DR, Lott JA, Nolte FS, Gretch DR, Koff RS, et al., Diagnosis and monitoring of hepatic injury: II. Recommendations for use of laboratory tests in screening, diagnosis and monitoring, Clin. Chem., 46, 2000, 2050-2068.
- 35. Nathwani RA, Pais S, Reynolds TB, Kaplowitz N, Serum alaninen-aminotransferase in skeletal muscle diseases, Hepatology, 41, 2005, 380-382.
- Bagchi D, Bagchi M, Hassoun EA, Stohs SJ, *Ib* and *i*n
 ib generation of reactive oxygen species, DNA damage and lactate dehydrogenase leakage by selected pesticides, Toxicology, 104, 1995, 129–140.
- 37. Yadav P, Sarkar S, Rhatnagar D, Action of Capparis deciduas against alloxan- induced oxidative stress and diabetes in rat tissues, Pharmaceut Res., 36, 1997, 221–228.
- Kaczor JJ, Ziolkowski W, Popinigis J, Tarnopolsky M, Anaerobic and aerobic enzyme activities in human skeletal muscle from children and adults, Pediatr Res., 57, 2005, 331–335.
- Rani MS, Emmanuel S, Sreekanth MR, Ignacimuthu S, Evaluation of *inip* antioxidant and hepatoprotective activity of Cassia occidentalis Linn. against paracetamol induced liver toxicity in rats, Int. J. Pharmacy Pharmaceut. Sci., 2, 2010, 67-70.
- 40. Akinrinmade JF, Akinrinde AS, Effect of oral administration of methanolic extract of Ocimum gratissimum on intestinal Ischemia-reperfusion injury in rats, Eur. J. Med. Plants, 3, 2013, 591-602.
- 41. Zhou T, Zong R, Zhang Z, Zhu C, Pan F, SERPINA3K protects against oxidative stress via moduling ROS generation/degradation and KEAP1-NRF2 pathway in the corneal epithelium. Invest. Ophthalmol, Visual Sci., 53, 2012, 5033-5043.
- Bebe FN, Panemangalore M, Pesticides and essential minerals modify endogenous antioxidants and cytochrome P450 in tissues of rats, J. Environ. Sci. Health, B40, 2005, 769-784.
- Kalender S, Ogutcu A, Uzunhisarcikli M, Acikgoz F, Durak D, Ulusoy Y, Kalender Y, Diazinon-induced hepatotoxicity and protective effect of vitamin E on some biochemical indices and ultrastructural changes, Toxicol., 211, 2005, 197-206.
- 44. Sakr SA, Abel-Samie, HA, Apoptosis related protein Bax in liver of metalaxyl fungicide –treated mice: The effect of antox. Ozean, J. Appl. Science, 1(1), 2008, 17-27.
- 45. Pandey KB, Rizvi SI, Markers of oxidative stress in erythrocytes and plasma during aging in humans, Oxid. Med. Cell Longev., 3, 2010, 2-12.
- Pryor WA, Stanley JP, A suggested mechanism for the production of malonaldehyde during the autoxidation of polyunsaturated fatty acids, Nonenzymatic production of prostaglandin endoperoxides during autoxidation, J. Org. Chem., 40, 1975, 3615–3617.



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- 47. Esterbauer H, Cheeseman KH, Determination of aldehydic lipid peroxidation products: Malondialdehyde and 4hydroxynonenal, Methods Enzymol., 186, 1990, 407-413.
- 48. Lykkesfeldt J, Malondialdehyde as biomarker of oxidative damage to lipids caused by smoking, Clin. Chim. Acta, 380, 2007, 50-58.
- 49. Kalpana C, Menon VP, Curcumin ameliorates oxidative stress during nicotine-induced lung toxicity in Wistar rats, Ital. J. Biochem., 53(2), 2004, 82-86.
- 50. Daniel S, Limson JL, Dairam A, Watkins GM, Daya S, Through metal binding, curcumin protects against leadand cadmium-induced lipid peroxidation in rat brain homogenates and against lead-induced tissue damage in rat brain, J. Inorg. Biochem., 98(2), 2004, 266-275.
- 51. Borek C, Antioxidant health effects of aged garlic extract, J. Nutr., 131, 2001, 1010 S-1015 S.
- 52. Egen-schwind C, Eckard R, Kemper FH, Metabolism of garlic constituents in the isolated perfused rat liver, Planta. Medica., 58, 1992, 301-305.

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