



A Review: Traditional, Ethnomedicinal Utilization, Pharmacological Properties and Phytochemistry of *Barleria prionitis* Linn.

Ajeet Singh*, Navneet

¹Department of Botany and Microbiology, GurukulKangri University, Haridwar -249404, Uttarakhand, India.

*Corresponding author's E-mail: ajeetchoudharykv@gmail.com

Received: 05-03-2017; Revised: 06-04-2017; Accepted: 10-06-2017.

ABSTRACT

Barleria prionitis Linn. is a widely dispersed indigenous plant throughout the Indian subcontinent. In the Ayurvedic medicine of India, it has a significant place due to its biological and pharmacological activities. The various parts of *B. prionitis* are widely used to cure an array of ailments by different ethnic communities. The whole plant or its parts like leaf, root, stem, bark and flower has been widely utilized for the cure of catarrhal affections, swellings, whooping cough, inflammations, toothache, glandular swellings, urinary infection, fever, gastrointestinal infections, diuretic and also in the treatment of dental infections. A lot of efforts have been made by several researchers to substantiate the effectiveness of plant in the course of precise biological and pharmacological activities to cure of various diseases. The examination of scientific literature revealed the outstanding biological activities of this plant such as antidiabetic, antibacterial, antifungal, analgesic, anti-inflammatory, hepatoprotective, antioxidative property etc. The excellent biological activity of the *B. prionitis* is due to presence of a wide range of phytochemicals like balarenone, pipataline, prionisides, barlerino side, verbascoside, barlerin, acetyl barlerin, lupulinoside, scutellarin that are responsible for a group of biological and medicinal activities. This review summarizes the current knowledge of the *B. prionitis* with a comprehensive insight, especially focusing on their traditional, ethanobotanical properties, pharmacognostic, phytochemical and pharmacological activity.

Keywords: *Barleria prionitis* Linn, Traditional uses and ethanobotanical aspects, Phytochemistry, Pharmacological properties.

INTRODUCTION

Plants have been used as traditional medicine for several thousands of years. Since the beginning of this century, ethno-botanical and traditional uses of natural compounds, mainly of plant origin established much interest as they are well tested for their efficacy and generally believed to be safe for human use. Plant derived medicine is still a basis of about 70-80% of the world's population as they are effortlessly accessible source for healthcare purposes in rural and tribal areas. India being the largest producer of medicinal plants therefore it is perfectly recognized as botanical garden of the world. Plants are the backbone of all life on the Earth and indispensable resource for human welfare as raw medicine, food and fuel. According to WHO more than 80% of world's population relies on traditional medicine for their health care needs¹⁻³. Traditional plant derived medicines have been used in most parts of the world and their use in combating microbial diseases is attractive the focus of a number of studies^{4-5,3}. Plant derived substances have recently become of great interest owing to their resourceful applications. It has been estimated that 14-28 % of higher plant species are used in the medicinal purposes and that 74% of pharmacologically active phytochemicals components were revealed after following up on ethno medicinal exploit of the plants⁶. In the last couple of decade, a new progress in the research and promotion of plants based drugs has become increasingly towards the herbal medicines⁷⁻⁸. At the present time multiple drug resistance has developed due

to indiscriminate exploitation of commercial antimicrobial drugs that frequently used in the treatment of infectious diseases⁹.

Distribution

B. prionitis is distributed throughout the hotter parts of India and commonly grown in gardens as a hedge plant¹⁰⁻¹¹. In India it is commonly found in Andaman and Nicobar Islands, Andhra Pradesh, Assam, Bihar, Chhattisgarh, Delhi, Diu and Daman, Goa, Gujarat, Jharkhand, Karnataka, Kerala, Laccadive and Maldiv Islands, Madhya Pradesh, Maharashtra, Orissa, Pudhucherry, Rajasthan, Tamil Nadu, Uttarakhand, Uttar Pradesh and West Bengal⁵.

Habitat

Barleria prionitis Linn. (Acanthaceae) is well known perennial ayurvedic herb dispersed all over Africa, India, Sri Lanka and tropical Asia. In ayurveda it is known by various names like kuranta, kurantaka, kuranda, kurandaka, sahachara, shairiya. In folk medicine it is popularly known as piyaabasaa, jhinti and ketsariyaa. It is also known as 'vajradanti' because of its anti dotalgic property^{10,12}.

Taxonomy

Kingdom – Plantae
Division – Magnoliophyta
Class – Magnoliopsida
Order – Scrophulariales
Family – Acanthaceae



Genus – *Baleria*

Species – *prionitis*

Morphology

B. prionitis Linn. is erect, bushy shrub grows up to 1-2 m in height. They possess 2-4 sharp long axillary spines which about 11 mm long. The stems are erected, glabrous, much branched with cylindrical and tapering branchlet¹³⁻¹⁴. *B. prionitis* is a shrub and flowers are yellow in colour. Flowering occurs during August – October¹⁵⁻¹⁶.

The flowers are sessile, and often solitary in lower axils and spicate in the upper axils. Flowers are equally broad as well as tubular and about 3-4 cm in length. The fruits are ovoid and capsular. The leaf are elliptic containing 5-20 mm long spines is about 3-10 cm long and 1.5-4 cm broad. The stems are light tan or gray coloured stiff, round, cylindrical and glabrous¹⁷⁻¹⁸. Leaves are smooth, opposite, ovate-elliptic to obovate, acuminate, tapering to base, bristle-tipped and about 6-15 cm long and 4-6 cm wide. The petioles are about 0.5-3 cm long¹¹. The fruit capsule is ovoid, 2 seeded and about 1.5-2 cm long and 0.6-0.8 cm wide. The seeds are oval-oblong, covered with silky copper-brown appressed hairs and measuring about 7.4-8.5x6-6.8 mm. The seeds of *B. prionitis* are flattened, covered with tangled hairs, about 8 mm long and 5 mm wide. Bracts are acute, linear-lanceolate, foliaceous, about 1-1.5 cm long and 0.2-0.8 cm wide with bristle tipped. The corolla is bright, golden yellow in colour with pubescent outside and glabrous inside and measuring 1.5 cm long. The stamens include 2 fertile stamens and 2 staminoid stamens. The fertile stamens are exerted away from the corolla tube while the staminoid stamens are very short. The filaments are hairy and about 2-2.5 cm long, glandular-pubescent and yellowish in colour. The anthers are yellow in colour with 3 mm long^{13-14, 11}.

Traditional and Ethno botanical uses

The whole plant, root, leaves and bark of the *B. prionitis* Linn. be present in a crucial place in the indigenous system of medicine (Ayurveda) in India for controlling the different types of ailments such as inflammations, swellings, boils, glandular etc¹⁹⁻²¹. The juice of *B. prionitis* has been reported to use for cure of whooping cough in Uttar Pradesh and Madhya Pradesh states of India, and leaves are use for the treatment of toothache, rheumatism, and root powder to cure fever²²⁻²⁴. The juice of leaves of *B. prionitis* is useful in fungal infections, ulcer and fever²⁵⁻²⁷. The decoction of aerial parts of *B. prionitis* is used in whooping cough, anti-respiratory syncytial virus, antiarthritic, anti-inflammatory and antifertility activities. It was also reported that 4 g of plant powder mixed with Nimbuka Swarasa and given twice in a day for 10 days to cure tonsillitis²⁸. Root extract is uses locally on skin to expel out spine from the skin and decoction is taken orally for the cure of snakebite²⁹⁻³⁰. *B. prionitis* is used in urinary infection, jaundice, hepatic obstruction and dropsy. Ash of the whole plant with honey is uses in bronchial asthma^{31-32, 19}.

In India, the aerial parts like leaves, stem, and flowers are used in catarrhal affections of children, glandular swellings, boils, fever, toothache, inflammation and gastrointestinal disorders. Bark is uses in whopping cough as an expectorant; the whole plant particularly the roots are used as tonic and diuretic^{34-34, 15, 35-37}. In medicobotanical survey of villages of Bulandshar district of Uttar Pradesh, (India), rural residents use *B. prionitis* in cases of asthma and whooping cough. Local peoples called it Kala Bansa or Piya-Bansa³⁸. *B. prionitis* is used in stiffness of limbs, enlargement of scrotum and sciatica²⁶⁻²⁷.

In Maharashtra (India), crushed leaves of *B. prionitis* are applied on the wound³⁹. It was revealed in an ethnomedicinal survey that pills prepared from *B. prionitis* are used for massage in combination with coconut oil and these pills give purity, rubefacient and blotch to body⁴⁰. The folk medicinal healers of Bangladesh use the *B. prionitis* for anti-inflammatory activity, and also for the treatment of cancer and tumour⁴¹. In a study it was reported that *B. prionitis* root with goat milk is given to treat rheumatic fever. Root, stem or leaves powder with cow milk is taken as remedy for dropsy and liver congestion⁴². In a ethnomedicinal survey conducted in Andhra Pradesh, India, revealed that local residents use *B. prionitis* to increase vitality by using seed extract daily once for fortnight. *B. prionitis* also being used in gout, ulcer of mouth and oedema⁴³. In Orissa (India) the *B. prionitis* have been used in cuts, wounds and malaria⁴⁴, and In Gujarat (India), leaf ash is being used for the management of leucoderma by applying with butter⁴⁵. The use of *B. prionitis* fresh leaf paste has also been reported against Scabies in Karnataka (India)⁴⁶.

Antimicrobial properties

Antibacterial activities

The antibacterial activity of different parts of *B. prionitis* has been reported. It was also reported that among the extracts, MeOH bark extract showed potential antibacterial activity against all the pathogens. Crude MeOH extract revealed good antibacterial activity against MDR (multidrug resistance) *E. coli* with 12 mm of inhibition zone^{21, 47}. Chetan et al., 2010 were reported the antibacterial activity of EtOH (ethanolic) leaf extract of *B. prionitis* against *S. aureus*, *B. subtilis*, *P. vulgaris*, *K. pneumoniae*, *E. coli* and *P. aeruginosa*. Antibacterial activity of H₂O, PET, CHCl₃ and ACE extracts *B. prionitis* were reported against *L. rhamnosus* (MTCC1408), *S. mutans* (MTCC 890), *S. aureus* MTCC 3408), *A. viscosus* (MTCC 7345), *S. epidermidis* (MTCC 3639), *E. coli* (MTCC 732) and *B. subtilis* (MTCC 3160)⁴⁸. Prominent inhibition of the four extracts was observed for bacterial species, *L. rhamnosus* and CHCl₃ extract was found to be more effective against the entire test microorganism⁴⁸. *In vitro* propagated shoot tips and leaves of *B. prionitis* with EtOH, ether and CHCl₃ extracts showed the antibacterial activity³¹. Antibacterial activities of *B. prionitis* bark and leaf MeOH extracts against *B. cereus* (22.66 mm) followed by PET leaf extract against *E. coli* (21.66 mm). Minimum



inhibition was showed by PET leaf extract against *A. faecalis* (4.66 mm) followed by MeOH bark extract against *A. faecalis* (5.33 mm)⁴⁹. Patel et al., 2015 reported the ethyl acetate of *B. prionitis* leaves extract showed inhibition zone on Gram positive *B. pumilus* (9.83 mm) and MeOH extract of *B. prionitis* stem showed inhibition zone on Gram negative *E. coli* (0.16 mm). PET extract did not show inhibition except, PET extract of *B. prionitis* stem on Gram positive *B. pumilus* (0.46 mm). MIC was showed by PET extract of *B. prionitis* leaves on Gram positive *B. pumilus* and Gram negative *P. aeruginosa* (1.0 mg/mL). Leaves and stem extract of *B. prionitis* showed difference in antibacterial activity. Aiswarya and Ravikumar, (2014)⁵⁰ reported the PET and EtOH extract of *B. prionitis* showed good antibacterial activity. The PET extract of *B. prionitis* was most effective against *P. putida* and *B. subtilis* with a zone of inhibition of 28 mm. Zone of inhibition for PET extracts of *B. prionitis* was compared with standard antibiotics. The EtOH extract of *B. prionitis* was most effective against *P. putida* with a zone of inhibition of 25 mm. Bacterial strains of *E. coli*, *P. beteli*, *P. fluouscense*, *S. paratyphi*, *S. aureus*, *B. subtilis*, *P. putida* were selected. The PET extract of *B. prionitis* was most effective against *P. putida* and *B. subtilis* while the EtOH extract of *B. prionitis* was most effective against *P. putida*⁵⁰. The antibacterial activity of rhizome of *B. prionitis* MeOH extract reported maximum inhibition zone (16.2 mm) against *E. coli* and minimum against *S. typhi*⁵¹. ACE, EtOH, MeOH extract of bark and ciprofloxacin showed significant activity against *S. mutans* (14.95±1, 11.94±1, 15.65±0.57 and 27.32±0.57 mm), *S. aureus* (14.31±0.57, 14.0±0, 16.32±0.57 and 34.66±0.57 mm), *Pseudomonas* sp. (18.32±0.57, 17.65±0.57, 19.32±0.57 and 33.66±0.57 mm) and *Bacillus* sp. (27.32±0.57, 23.97±1, 28.65±0.57 and 29.65±0.57 mm). The antibacterial activity of *B. prionitis* leaf extracts were reported against *S. typhi*, *V. cholerae*, *M. luteus*, *L. sporogens*, *Citrobacter*, *B. subtilis*, *B. cereus*, and *Providencia*⁵².

Antifungal activities

The antifungal activity of ACE, EtOH and MeOH bark extracts of *B. prionitis* against *S. cerevisiae*, *C. albicans* and MeOH extract was found more active against all the fungal strains^{31,21}. Antifungal activity of *B. prionitis* were reported against *C. neoformans*, *C. albicans*, *C. vaginitis*, *B. dermatidis* using CHCl₃, acetone and EtOH extract of stem, leaves and roots⁵³.

It was also revealed that PET, dichloromethane and EtOH stem and root extracts of *B. prionitis* showed fungistatic and fungicidal properties against *C. albicans*^{12,54}.

Anti-dental decay activity

Crude extract of *B. prionitis* Linn. reported good antimicrobial activity against dental decay pathogens. It was reported that MeOH extract of bark showed much more potent activity against oral pathogens like *S.*

mutans, *S. aureus*, *Pseudomonas* sp., *Bacillus* sp. and *C. albicans*, *S. cerevisiae*²¹.

Antiviral activities

Two iridoid glycosides (i.e., 6-O-trans-p-coumaroyl-8-O-acetylshanzhiside methyl ester and its cis isomer from *B. prionitis* were reported by Chen et al. (1998)⁵⁵. These bioactive phytochemicals revealed the potent antiviral activity against respiratory Syncytial virus (RSV) with EC₅₀ and IC₅₀ values of 2.46 and 42.2 µg mL⁻¹, respectively^{12,55}.

Pharmacological properties

Antioxidant activities

The MeOH extract of root leaves and stems showed potent antioxidant activity. EtOH extract of whole plant of *B. prionitis* showed significant antioxidant activities. It was reported that the antioxidant activity of MeOH extract of leaf and stem were showed IC₅₀ values 63.41±0.32, 81.69±0.40, respectively. Reducing power of the MeOH extract of *B. prionitis* was observed maximum^{56, 57, 58}. *In vitro* investigation showed that the EtOH and H₂O extract of whole plant possess considerable antioxidant activity⁵⁸. MeOH leaf extract showed significant high antioxidant activity (61.73) in 6000 ppm concentration followed by PET bark extract (59.11)⁴⁹. *In vitro* antioxidant activity of crude MeOH extract of *B. prionitis* was reported by Khobragade et al., (2012)⁵⁹.

MeOH extracts of roots, leaves and stems showed significant antioxidant potential⁵⁴. The leaves of *B. prionitis* showed high level of antioxidant activities as well as high amount of phenolic content as compared with flower and stem⁶⁰. Thabrew et al., (2001)⁶¹ was reported that effect of marketed preparation containing *B. prionitis* for antioxidant potential on rheumatoid arthritis patients. This investigation showed that three months treatment of preparation has high antioxidant potential.

Anti-diabetic activities

The alcoholic leaves extract of *B. prionitis* revealed antidiabetic potential. In a study it was reported that oral administration of alcoholic extract at dose concentration 200 mg kg⁻¹ body weight considerably decreased the blood sugar, glycosylated haemoglobin level and increased serum insulin and liver glycogen level in diabetic test organism (rats). The alcoholic extract of root of *B. prionitis* showed a moderate but insignificant antidiabetic activity in investigational animals⁶².

Anti-arthritis activities

The TAF fraction was showed antiarthritic activity in *M. tuberculosis* induced adjuvant arthritis rats model⁶³. It was observed that *B. prionitis* and isolated shanzhiside esters from the same plant can be strongly categorized under potential antiarthritic drugs since both were active in adjuvant induced arthritic model⁶⁴. From the bibliography search it was revealed that three reports have been known where *B. prionitis* showed anti-arthritis potential^{12, 63, 65, 66}.



Anti-fertility activities

The roots extract of *B. prionitis* showed the antifertility potential⁶⁷. Oral administration of MeOH root extract reduced the sperm formation in male albino rats^{67,68}. Root extract decreased the formation of round spermatids, sperm motility, spermatogonia, preleptotene spermatocytes population and mature leydig cells⁶⁷.

Anti-helminthic activity

Anti-helminthic activity of *B. prionitis* whole plant extract was reported in dose dependent manner. It was showed that *in vitro* EtOH and H₂O extracts were significantly paralyzed the *P.posthuma*, a worm at 50, 75 and 100mg/mL⁻¹ and also comprised with a standard drug albendazole. The extracts of *B. prionitis* caused death above 100 mg mL⁻¹⁶⁹.

Anti-diarrheal activity

Butanol fraction of *B. prionitis* leaves showed the anti-diarrheal activity. Iridoid rich fraction of butanol (BuOH or n(BuOH)) of leaf extract possess dose dependent anti-diarrhoeal activity at the concentration of 25-100 mg/kg in rats against castor oil induced diarrhoea⁷⁰⁻⁷¹.

Anti-inflammatory activities

Several reports demonstrated the usage of *B. prionitis* in the treatment of inflammations. The anti-inflammatory activity of *B. prionitis* was evaluated through *in vitro* enzyme based cyclooxygenase (COX-1 and COX-2) assays. It was found that the dichloromethane (DCM), PET and EtOH extracts of leaves, stems and roots exhibited significant inhibition of COX-1 and COX-2 with subsequent inhibition of prostaglandin synthesis that are involved in pain sensation⁵⁴. The H₂O fraction (TAF) of hydro-ethanolic extract of *B. prionitis* whole plant reported to have significant anti-inflammatory activity against the acute inflammation induced by carrageenan, histamine and dextran in rats. The anti-inflammatory activity of the 'TAF' may be due to the presence of iridoid glucosides, shanzhiside methyl ester, acetyl barlerin and barlein⁷². Another study revealed that the H₂O extract fractions (FR-III and FR-IV) of root significantly inhibited the caragennan induced rat paw edema³². The FR-III and FR-IV at oral dose concentration of 400 mg kg⁻¹ body weight inhibited the paw edema by 50.64 and 55.76%, respectively and the results were comprised with the reference standard drug (indomethacin) with a 60.25% of inhibition³². The EtOH extract of flowers also exhibited anti-inflammatory activity in rats⁷³. Oral administration of flower extract (200 mg kg⁻¹ body weight) showed significant dose-dependent reduction in carrageenin induced swelling and cotton pellet granuloma weight that were equivalent to 48.6 and 36.4% protection⁷³.

Cytoprotective activities/ mast cell migration activity

Mast cells play an important role in inflammatory responses and release histamine upon their degranulation to produce various allergic

reactions⁷⁴. Maji *et al.* (2011)⁷⁵ reported that the hydro-methanolic extract of *B. prionitis* (whole plant) showed dose-dependent mast cells and erythrocyte membrane protection activity in response to the toxic chemicals. The extract inhibited the compound 48/80 induced mast cells degranulation up to 64.91% at dose concentration 10 µg mL⁻¹ and the result was comprised with the reference standard (disodium cromoglycate) (10 µg mL⁻¹) with 19.32% protection⁷⁵. The extract (10 µg mL⁻¹) provided significant erythrocyte membrane protection (27.10%) against hypotonicity haemolysis and the result was comprised with reference standard (indomethacin) (10 µg mL⁻¹) with 61.29% protection⁷⁵.

Enzyme inhibitory effects

The extracts from different parts and isolated phytochemicals of *B. prionitis* reported to inhibit the clinically significant enzymes, Acetylcholinesterase (AChE) and glutathione S-transferase (GST). Kosmulalage *et al.* (2007)⁷⁶, Ata *et al.* (2007,2009)⁷⁷, Amoo *et al.* (2009)⁵⁴, reported that the MeOH extracts of leaf, stem and root of *B. prionitis* exhibited AChE inhibitory performance and the leaf and stem extracts exhibited higher potency of inhibition in compare the root extract. Several glycosides compounds showed different levels of AChE inhibitory activity. Prioniside B and prioniside C also showed GST inhibitory activity of which prioniside B and prioniside C were more potential GST inhibitors⁷⁶⁻⁷⁷.

Hepatoprotective activity

The iridoid glycosides enriched fraction from hydro-ethanolic extract of leaves and stems of *B. prionitis* was reported to show significant hepatoprotection against carbon tetrachloride, galactosamine and paracetamol induced hepatotoxicity in mice and rats⁷⁸. The oral administration of iridoid fraction significantly reduced the hepatotoxin induced elevated levels of serum alanine aminotransferase (ALT), aspartate transaminase (AST), alkaline phosphatase (ALP), bilirubin and triglycerides in a dose dependent manner. The fraction was also increased the hepatic glutathione content and reduced the hepatic lipid peroxidation in response to the hepatotoxicity in mice and rats⁷⁸.

Diuretic property

B. prionitis showed diuretic activity was reported by the extract of leaves and young inflorescence. Juice of leaves is used in urinary afflictions. The diuretic property may be due to the presence of high concentration of potassium⁷⁹⁻⁸⁰. The diuretic property of *B. prionitis* flower extract was performed by Musale *et al.* (2011)⁸¹. The oral administration of flower H₂O extract (200 mg kg⁻¹) was significantly increased the urination (dieresis) and sodium removal but not potassium in rats. The diuretic effect of flower extract (200 mg kg⁻¹) was comprised and statistically significant with drug furosemide (20 mg kg⁻¹)⁸¹.



Central nervous system depressant activity

Ethyl acetate portion (at dose concentration of 125 and 250 mg/kg) and diclofenac (4 mg/kg) treatment significantly increased fall off time of motor co-ordination in rota rod test⁶⁵. EtOH extract of *B. prionitis* leaves by using acto-photometer reported fluoxetine stimulant activity in mice as 91.93% while the test drug stimulated the animal only by 49.72%⁸².

Toxic effects

In a study it was reported that the alcoholic extract of roots and leaves of *B. prionitis* did not reported any toxic effects in adult albino rats⁶². In a study Dheer and Bhatnagar (2010) observed that the oral administration of alcoholic extract at the dose concentration up to 2.5 g kg⁻¹ body weight throughout the 14 days of study period without any mortality. Singh *et al.* (2005)⁷⁸ reported that the iridoidglucosides rich aqueous portion *B. prionitis* did not produced any signs of abnormalities or any mortality up to the single oral administration of 3000 mg kg⁻¹ dose in mice during the 15 days of study period. Nevertheless, the intra-peritoneal LD₅₀ was determined as 2530 mg kg⁻¹ for the aqueous portion in mice⁷⁸. In another study the acute oral toxicity of MeOH extract of *B. prionitis* was reported using Spargue – Dawley rats (n=5). The LD₅₀ was found to be more than 200 mg/kg, with no sign of abnormality or any mortality observed for 14 days after single dose administration⁶⁵.

Antinociceptive activity/analgesic activity

The analgesic activity of *B. prionitis* flowers extract was reported using an UgoBasile Analgesy meter induced artificial pain and acetic acid induced writhing models⁷³. *In vivo* study showed that the flower extract dose dependently provided a significant increase in the analgesio-meter-induced force and exhibited significant resistance against pain in mice⁷³. At a dose concentration of 50 mg kg⁻¹ body weight, the flower extract provided statistically significant reduction of writhing by 5.24%⁷³.

Anti-hypertensive property

The antihypertensive activity of MeOH extracts of leaves of *B. prionitis* using DOCA salt. *B. prionitis* showed significant anti-hypertensive effect in DOCA salt induced hypertensive rats in dose of 200 mg/b.w. and 400 mg/b.w.⁸³. DOCA salt induces reabsorption of salt and water leading to induced blood volume and hence increased blood pressure. SBP and DBP were increased persistently in DOCA salt treated nephrectomised rats as compared to normal Rat's *B. prionitis* extracts⁸³.

Anti-cataract activity

Atif *et al.*, (2015)⁸⁴ reported that the administration EtOH leaves extract of *B. prionitis* significantly restored the glutathione and malondialdehyde levels. SOD, catalase and glutathione S transferase levels were significantly restored to normal levels (p<0.05 and p<0.01

respectively). Oral administration of *B. prionitis* significantly late the onset and progression of cataract in selenite as well as galactose induced cataract. It can be said that *B. prionitis* significantly reversed the cataract parameters by virtue of its antioxidant property⁸⁴.

Gastro-protective activity

Maximum protections were found to be 66.26% and 59.42% by iridoid fraction (200 mg/kg) in PL induced ulcer and CRS-induced ulcer rat model. Iridoid fraction from leaves reduced ulcer index. In EtOH induced gastric ulcer rat model, MeOH extract of leaf (500 mg/kg bw) and ranitidine provided 67.7 and 75.5% inhibition of ulcer. Same dose of extract and drug displayed 70.3 and 62.2% inhibition in indomethacin induced gastric ulcers model. Extract also showed efficacy against indomethacin induced gastric mucosal damage and increased liver enzymes in EtOH induced ulcer rat model⁸⁵.

Anti-arthritis activity

Chaudhary *et al.*, (2014)⁶⁵ reported that the ethyl acetate fraction of *B. prionitis* leaves extract possesses antiarthritic activity in Sprague Dawlys rats following OECD 420 guidelines. Dose dependent and significant inhibition of oedema was observed in both acute as well as chronic models. The leaves extract of *B. prionitis* at dose 250 mg/kg showed most potent and significant (P 6 0.05–0.01) paw oedema inhibition which is supported by the results of body weight, biochemical parameters, and motor in coordination and nociceptive threshold in Freund's Complete Adjuvant-induced arthritis model⁶⁵. The effect of two different extracts namely MeOH (ME, prepared by maceration) and butanolic (BE, obtained after partitioning of ME) of *B. prionitis* and the isolated three major iridoids viz., acetylbarlerin (AB), barlerin (B) and shanzhiside methyl ester (SME) from the plant using chromatographic techniques were evaluated in a rat model of Complete Freund's Adjuvant (CFA) induced-arthritis at a single dose of 200 mg/kg for extracts and 1 mg/kg for pure compounds. The results were compared to untreated control and standard (indomethacin, INDO) treated groups. It was observed that on 21st day of experiment, the histopathological, and radiological and biochemical explanations were carried out along with rheumatoid factor. The serum level of cytokines (TNF- α and IL-1 β) were also determined using ELISA kits. The results indicate that *B. prionitis* protects rats against the bone loss, body weight changes and haematological perturbations induced by CFA. Further the histopathological and radiological studies also support the generated observations. Thus, the positive effect of the test samples in controlling the various parameters associated with the progression of arthritis demonstrated their pronounced antiarthritic effects, indicating that *B. prionitis* would be a potent candidate for treating arthritis⁶⁴.



Larvicidal activity

LC₅₀ values were found to be 34.756, 31.351 and 28.577 µg/mL in ACE, CHCl₃ and MeOH extract of leaf against *Culex tritaeniorhynchus*, respectively⁸⁶.

Phytochemical constituents

Preliminary phytochemical screening of *B. prionitis* hydro-methanol extract of whole plant revealed the occurrence of saponins, glycosides, tannins and flavonoids⁷⁵. Leaves and stem of *B. prionitis* showed the presence of alkaloids but absence of tannins and saponins were collected from Gujarat (India)⁸⁸. *B. prionitis* reported several phytochemicals such as balarenone, lupeol, prioniside A, prioniside B, prioniside C, pipataline were reported in EtOH extract⁷⁷. Bharat et al., (2006) isolated and identified few phytochemicals from *B. prionitis* like acbarlerin, barlerin, β-sitosterol, flavanol glycoside, iridoids and scutellarein-7-neohesperidoside and showed their anti-inflammatory activities⁸⁸. Some other bioactive phytochemicals like luteolin-7-O-β-D-glucoside, β-sitosterol, scutellarein 7-neohesperidoside, apigenin 7-O-glucoside, 13,14-seco-stigmasta-5,14-diene-3-a-ol are found in *B. prionitis*^{67,10,76,89,90}. Barlerinoside, 6-O-trans-p-coumaroyl-8-O-acetylshanzhiside methyl ester, 7-methoxydideroside, 13, and lupulinoside have been isolated from the aerial parts of *B. prionitis*⁷⁶⁻⁷⁷. *Barleria* genus is reported to have iridoids, anthraquinones, sterols, fatty acids and flavonoids. Iridoids comprise the major class of compounds isolated from *Barleria* and important bioactive iridoids are acetylbarlerin, barlerin and shanzside methyl ester⁹¹. For the first time 6-hydroxyflavones have been reported in the family Acanthaceae in the genus *Barleria*⁹⁰. The MeOH extract of *B. prionitis* showed the presence of phenols, flavonoids, glycosides, proanthocyanidins, alkaloids and tannins. Phenol and phenolic compounds like flavonoids have been shown to possess significant antioxidant properties⁹². The leaves and flowering tops of *B. prionitis* showed high amount of potassium salts¹⁰. *B. prionitis* extracts revealed the presence of alkaloids, flavonoids, steroids, saponins, tannins and phenolic compounds, because of these compounds the plants shows significant antihypertensive activity⁸³. Total phenolic and flavonoid content of the *B. prionitis* was 0.33±0.1 mgGAE/g and 0.9±0.5 mg of Quercetin equivalent per gram of dry extract respectively⁵⁹. The total polyphenols content in the EtOH and H₂O extract of *B. prionitis* Linn. was showed 43.71 and 35.58 GAE/mg, respectively⁵⁸. The total phenolic content of *B. prionitis* MeOH extract of leaf was found maximum (103.51±0.38mg/g) followed by ethyl acetate (44.31±0.45 mg/g), H₂O (32.82±0.31 mg/g) and n-Hexane (8.33±0.21 mg/g). Stem extract showed maximum with MeOH (94.37±0.18 mg/g) followed by ethyl acetate (44.31±0.45 mg/g), H₂O (32.82±0.31 mg/g) and n-Hexane (8.33±0.21 mg/g), respectively⁵⁷. It was reported that *B. prionitis* showed some antibacterial bioactive compound that include with balarenone, pipataline and 13, 14-seco-stigmasta-5, 14-diene-3-a-ol have been isolated from the

ethanolic extract. These phytochemicals showed potent antibacterial activity against *P. aeruginosa* and *B. cereus*⁷⁶.

CONCLUSION

B. prionitis Linn. occupy a significant place in the Ayurvedic medicine in all over, India, Sri Lanka including tropical Asia and Africa. *B. prionitis* Linn. depicted the piece of evidence that it is used as a cure for variety of ailments. It is fascinating to message that pure phytochemicals and crude extracts of leaves of *B. prionitis* Linn. have been screened for some pharmacological activities and found to have analgesic, anti-inflammatory, hepatoprotective activity and stem bark of the plant have antidiabetic activity, and juices are screened for hypocholesterolemic and antioxidant activity. The comprehensive survey information as provided in this review on *B. prionitis* traditional uses, ethanobotanical aspects, phytochemistry, pharmacology and toxicity of the extracts of different parts. All-embracing literature survey given away the promising pharmacological includes antimicrobial, anthelmintic, antifertility, antioxidant, antidiabetic, anti-inflammatory, anti-arthritis, cytoprotective, hepatoprotective, anti-diarrhoeal, enzyme inhibitory, diuretic and anti-nociceptive or analgesic activities of the extract and isolated bioactive compounds from *B. Prionitis* devoid of toxicity.

REFERENCES

- Gautam SS, Navneet, Kumar S. The antibacterial and phytochemical aspects of *Viola odorata* Linn. Extracts against respiratory tract pathogens. Proc. Natl. Acad. Sci., India B Biol. Sci. 82(4), 2012, 567-572.
- Singh A, Pathak VM Navneet. Screening of antimicrobial potential of *Barleria prionitis* Linn. aerial parts against common respiratory tract pathogens. Int. J. Curr. Microbiol. App. Sci., 5(7), 2016, 542-549.
- Singh A and Navneet, Evaluation of antimicrobial potential and phytochemical assessment of *Citrus maxima* Burm. Seeds extracts against respiratory tract pathogens. New. York. Sci. J. 9(9), (2016), 4-10.
- Bhavnani SM, Ballow CH. New agents for Gram-positive bacteria. Curr. Opin. Microbiol. 3, 2000, 528-534.
- Chariandy CM, Seaforth CE, Phelps RH. Screening of medicinal plants from Trinidad and Tobago for antimicrobial and insecticidal properties. J. Ethnopharmacol. 64, 1999, 265-270.
- Baroh M, Ahmed S, Das S. A comparative study of the antibacterial activity of the ethanolic extracts of *Vitex negunda* L., *Fragaria vesca* L., *Terminalia arjuna* and *Citrus smaxima* Burm. Asi J. Pharma. Biol. Res. 2(3), 2012, 183-187.
- Bisset NG. Herbal Drugs and Phyto pharmaceuticals. CRC Press, Boca Raton. 1994.
- Tyler VE. The Herbal Remedies Market. Chemtech. 27, 1997, 52-57.
- Naidu L, Kishore PV, Kumar K, Mohan Kumar C, Gunesh G, Narasimha Rao, M. Antimicrobial activity of *Achyranthes aspera*. Biosciences, Biotechnol Res Asia; 3(1), 2006.
- Khare CP. Indian medicinal plants: an illustrated dictionary. 1st Edn., Springer science, New York; 2007, 82-83.
- Shendage SM, Yadav SR. Revision of the Genus *Barleria* (Acanthaceae) in India. Rheedeia. 20, 2010, 81-130.
- Banerjee AK, Maji S, Mahapatra S, Banjeri P. *Barleria prionitis* Linn.: A Review of its Ttraditional uses, Phytochemistry, Pharmacology and Toxicity, Res. J. Phytochem. 6, 2012, 31-41.



13. Dassanayake MD. A Revised Handbook to the Flora of Ceylon. vol. 12, CRC Press, Boca Raton, USA; 87, 1998.
14. Kamble MY, Pal SR. Shendage SM, Dixit GB, Chavan PD, Yadav US, Yadav SR. Promising Indian Barlerias of Ornamental Potential. In: Underutilized and Underexploited Horticultural Crops, Peter, K. V. Ed. vol. 1. New India Publishing Agency, New Delhi; 144, 2007.
15. Kirtikar KR, Basu BD. Indian Medicinal Plants. Bishen Singh Mahendra Pal Singh, New Connaught Place, Dehradun; 1999, 1877-1878.
16. Kaushik P, Dhiman AK. Medicinal Plants and Raw Drugs of India. Bishen Singh Mahendra Pal Singh, Shiva Offset press. Dehradun, India; 2000, 412.
17. Sharma P, Shrivastava B, Sharma GN, Jadhav, HR. Phytochemical and Ethenomedicinal Values of *Barleria prionitis* L.: An overview. Jour Harmo Res Pharm. 2 (3), 2013, 190-9.
18. Jain S, Jain R, Singh R. Ethnobotanical survey of Sariska and Siliserh regions from Alwar district of Rajasthan, India. Ethnobotanical Leaflets; 1, 2009: 21.
19. Khare CP. Indian Herbal Remedies: Rational Western Therapy, Ayurvedic and Other Traditional Usage, Botany. 1stEdn., Springer, New York. 2004, 93-94.
20. Daniel M. Medicinal plants: chemistry and properties. 1stEdn., Science Publishers, USA, 2006, 78.
21. Aneja KR, Joshi R, Sharma C. Potency of *Barleria prionitis* L. Bark Extracts against Oral Diseases causing strains of Bacteria and Fungi of Clinical Origin. New York Sci. J. 3, 201, 5-12.
22. Mahajan SK. Traditional herbal remedies among the tribes of Bijargarh of West Nimar district, Madhya Pradesh. Indian J. Tradit. Knowle. 6(2), 2007, 375-377.
23. Singh AK, Raghubanshi AS, Singh JS. Medical Ethnobotany of the tribals of Sonaghati of Sonbhadra district, Uttar Pradesh, India. J. Ethnopharmacol. 81(1), 2002, 31-41.
24. Jadhav D. Ethnomedicinal plants used by Bhil tribe of Bibdod, Madhya Pradesh. Indian J. Tradit. Knowl. 5(2), 2006, 263-267.
25. Panwar HS, Nauriyal MM, Joshi HC. In vitro Screening of Certain Indigenous Plants for their Antimycotic Activity. Veterinary Res. Bul. 2.; 1979, 164-167.
26. Ambasta SP. The Useful Plants of India. New Delhi: CSIR; 1986.
27. Jain, SK, Defillips, RA. Medicinal plants of India. CSIR, New Delhi; 1991.
28. Radha J, Dilbag J, Shailaja U, Sudhakar P. Effects of saireyaka (*Barleria prionitis*) in tundikeri (tonsillitis): A Clinical Study. J. Biol. & Sci. Opin. 1(3), 2013, 168-72.
29. Karuppusamy S. Medicinal Plants Used by Paliyan Tribes of Sirumalai Hills of Southern India. Nat. Prod. Radi. 6(5), 2007, 436-42.
30. Jain A, Katewa S, Galav P, Nag A. Some Therapeutic Uses of Biodiversity among the Tribals of Rajasthan. Indian J. Tradit. Knowle. 7(2), 2008, 256-62.
31. Shukla P, Singh A, Gawri S, Alexander A, Sonwane S. *In vitro* propagation of *Barleria prionitis* Linn and its Antibacterial Activity. Int. J. Pharma Professional's Res. 2(1), 2011, 198-200.
32. Khadse CD, Kakde RB. Antinflammatory activity of aqueous extract fractions of *Barleria prionitis* L. roots. Asian J. Plant Sci. Res. 1, 2011, 63-68.
33. Chopra RN, Nayar S L, Chopra IC. *Barleria prionitis* Linn. In, Glossary of Indian Medicinal Plants. Council of Scientific and Industrial Research, New Delhi, 1956, 33-34.
34. Nadkarni AK. *Barleria prionitis* Linn. In Dr. KM. Nadkarni's. Indian Materia Medica, 3rd ed., Popular Book Depot. Bombay, 1994.
35. Parrotta JA. Healing plants of Peninsular India. CABI Publishing. New Delhi, India, 2001, 480-481.
36. Mohammed S, Kasera, PK, Shukla JK. Unexploited plants of potential medicinal value from the Indian Thar desert. Nat. Prod. Radiance. 3(2), 2004; 69-74.
37. Kala CP. Ethno medicinal botany of the Apatani in the Eastern Himalayan Region of India. J. Ethnobiol. & Ethnomed. 1, 2005, 11.
38. Alam MM, Anis, M. Ethnomedicinal uses of plants growing in the Bulandshahr district of Northern India. J. Ethnopharmacol. 1987, 19(1), 85-88.
39. Patil BS, Naikwade NS, Kondawar MS, Magdu CS, Awale VB. Traditional uses of plants for wound healing in the Sangli district, Maharashtra. Int. J. Pharma Tech Res. 1(3), 2009, 876-878.
40. Das HB, Majumdar K, Datta BK, Ray D. Ethnobotanical uses of some plants by Tripuri and Reang tribes of Tripura. Nat. Prod. Radiance. 8(2), 2009, 172-180.
41. Mollick MAH, Hossan MS, Paul AK, Rahman MTU. A comparative analysis of medicinal healers in three districts of Bangladesh and inquiry as to mode of selection of medicinal plants. Ethnobot Res. & Appl., 8, 2010, 195-218.
42. Bhuvaneshwar U, Praveen, Anil KD, Ashwani K. Ethnomedicinal and ethano pharmacostatistical studies of Eastern Rajasthan, Indian. J. of Ethnopharmacol. (1), 129, 2010, 64-86.
43. Reddy KN, Trimurthulu G, Reddy CS. Medicinal plants used by ethnic people of Medak district, Andhra Pradesh. Indian J. Tradit. Knowle. 9(1), 2010, 184-190.
44. Rout SD, Panda SK. Ethnomedicinal plant resources of Mayurbhanj district, Orissa. Indian J. Tradit. Knowle. 9(1), 2010; 68-72.
45. Brijesh S, Falguni S, and Minoo P. Documenting Grandmas' prescriptions for skin ailments in Valsad district, Gujrat. Indian J. Tradit. Knowle. 10(2), 2011, 372- 374.
46. Ghatapanadi SR, Johnson N, Rajasab AH. Documentation of folk knowledge on medicinal plants of Gulbarga district, Karnataka. Indian J. Tradit. Knowle. 10(2), 2011, 349-353.
47. Chetan CB, Shinde UV, Hogade M, Bhinge S. Screening of *in vitro* antibacterial assay of *Barleria prionitis* Linn. J. Herb. Med. Toxicol. 4, 2010, 197-200.
48. Diwan PD, Gadhikar YA. Assessment of Phytochemical Composition and Antibacterial Activity of Different Extracts of *Barleria prionitis* Leaves against Oral Microflora to Improve Dental Hygiene. Asi J. Pharma & Clin Res. 5(2), 2012, 182-184.
49. Kumar U, Ahmed F, Khanojia P, Kukreja K, Kumari S, Bhat RA. Exploration of Antioxidant and Antibacterial Activity of *Barleria prionitis* Linn. Int. J. Curr. Microbiol. App. Sci. 2(12), 2013, 585-591.
50. Aiswarya T, Ravikumar RA. Comparative study on physicochemical analysis, antibacterial activity and antioxidant activity of *Barleria prionitis* leaves extract of Petroleum ether and Ethanol extract. Int. J. Chem Tech Res. 6(5), 2014, 3025-3033.
51. Nidhi, Uttam K, Sumit K. Identification and Screening of Bioactive Compounds in *Barleria prionitis* Linn. Rhizome Exhibiting Antibacterial Activity. Int. J. Res. Biotech and Biochem. 3(1), 2013, 1-6.
52. Paul S, Saha D. Comparative study of the efficiency of *Barleria prionitis* leaf extracts against bacteria. Asi. J. Pharma Res. 2(3), 2012, 107-110.
53. Panchal P, Singh K. Antimicrobial activity of *Barleria prionitis* on Pathogenic Strains. 7(4), 2015, 73-75.
54. Amoo SO, Nalala AR, Finnie JF, Van Staden J. Antifungal, acetyl cholinesterase inhibition, antioxidant and phytochemical properties of three *Barleria* species. Sou. Afri. J. Bot. 77, 2011, 435-445.
55. Chen JL, Blanc P, Stoddart CA, Bogan M, Rozhon EJ. New iridoids from the medicinal plant *Barleria prionitis* with potent activity against respiratory syncytial virus. J. Nat. Prod. 61, 1998, 1295-1297.
56. Kapoor A, Shukla S, Kaur R, Kumar R. Preliminary phytochemical screening and antioxidant activity of whole plant of *Barleria prionitis* Linn. Int. J. Adv. Pharma, Biol. & Chem. 3(2), 2014, 410-419.
57. Sharma P, Sharma GN, Srivastava B, Jadhav HR. Evaluation of Antioxidant Potential of *Barleria prionitis* leaf and stem. Ame J. Phytomed Clin. Thera. 2(11), 2014, 1177-1186.



58. Chetan C, Suraj M, Maheshwari C, Rahul A, Priyanka P. "Screening of antioxidant activity and total phenolic content of whole plant of *Barleria prionitis*." Linn. Int. J. Res. Ayurveda Pharma. 2: 2011, 1313-1319.
59. Khobragade CN, Bhande RM. *In vitro* antibacterial, membrane damage, antioxidant and anti-inflammatory activities of *Barleria prionitis* Linn. extract on UTI causing multidrug resistant *E. coli*. Int. J. Cur. Pharma Res. 4(1), 2012, 64-69.
60. Jaiswal SK, Dubey MK, Das S, Verma AR, Vijyakumar M, Rao CV. Evaluation of iridoid glycosides flower of *Barleria prionitis* for anti-inflammatory and anti-nociceptive activity. Int. J. Pharm. Biol. Sci. 1, 2010, 1-10.
61. Thabrew MI, Senaratna L, Samarawickrema N, Munasinghe C. Antioxidant Potential of two Polyphenol Preparations used in Aurveda for the treatment of rheumatoid arthritis. J. Ethnopharmacol. 76(3), 2001, 285-281.
62. Dheer, R, Bhatnagar P. A study of the antidiabetic activity of *Barleria prionitis* Linn. Indian J. Pharmacol. 42(2), 2011, 70-73.
63. Singh BS, Bani DK, Chandan BK, Kaul A. Anti-inflammatory activity of TAF an active fraction from the plant *Barleria prionitis* Linn. J. Ethnopharmacol. 85, 2003, 187-193.
64. Kaur PK, Sharma S, Karan M. Evaluation of Anti-arthritis Activity of the Plant *Barleria prionitis* Linn. Int. J. Pharm. Sci. Res. 37(2), 2016, 1321.
65. Choudhary M, Kumar V, Gupta PK, Singh S. Anti-arthritis activity of *Barleria prionitis* Linn. Leaves in Acute and Chronic Models in Sprague Dawley rats. Bull. Faculty Pharma., Cairo University. 52(2), 2014, 199-209.
66. Majumder PM, Mondal A, Sasmal D. Evaluation of antiarthritic and immunomodulatory activity of *Barleria lupulina*. Asi. Paci. J. Trop. Biomed. 2(3), 2012; 1400-1406.
67. Gupta R, Kumar S, Dixit, VP, Dobhal MP. Antifertility studies of the root extract of the *Barleria prionitis* Linn. In male albino rats with special reference to testicular cell population. J. Ethano pharmacol. 70(2), 2000, 111-117.
68. Verma PK, Sharma A, Joshi SC, Gupta RS, Dixit VP. Effect of isolated fractions of *Barleria prionitis* root methanolic extract on reproductive function of ale rate: preliminary study. Fitoterapia. 76, 2005, 428-432.
69. Chavan CB, Shinde UV, Hogade M, and Bhide, S. Screening of *in vitro* antibacterial assay of *Barleria prionitis* Linn. J. Herb. Med. Toxicol. 4, 2010, 197-200.
70. Jaiswal SK, Dubey MK, Das S, Verma AR, Vijyakumar M, Rao CV. Evaluation of iridoid glycosides flower of *Barleria prionitis* for anti-inflammatory and anti-nociceptive activity. Int. J. Pharm. Biol. Sci. 1, 2010b; 1-10.
71. Sunil KJ, Mukesh KD, Ajay KV, Sanjib D, Vijaykumar M, Chandana VR. Evaluation of iridoid glycosides from leave of *Barleria prionitis* as an anti-diarrhoeal activity: An Ethno pharmacological study. Int. J. Pharma. Sci. & Res. 2(3), 2010, 680-686.
72. Singh BS, Bani DK, Gupta BK, Chandan and Kaul A. Antiinflammatory activity of TAF an active fraction from the plant *Barleria prionitis* Linn. J. Ethnopharmacol. 85, 2003, 187-193.
73. Jaiswal SK, Dubey MK, Das S, Verma AR, Vijyakumar M and Rao CV. Evaluation of flower of *Barleria prionitis* for anti-inflammatory and anti-nociceptive activity. Int. J. Pharm. Biol. Sci. 1, 2010, 1-10.
74. Manek RA, Sheth NR, Vaghasiya JD, Malaviya SV, Jiwari NP, Chavda JR. Study on herb-herb interaction potential of *Glycyrrhiza glabra* with *Solanum xanthocarpum* and *Adhatoda vasica* on mast cell stabilizing activity. Int. J. Pharmacol. 7, 2011; 589-598.
75. Maji AK, Mahapatra S, Banerji P and Banerjee D. Mast Cell Stabilization and Membrane Protection Activity of *Barleria prionitis* L. Pharmacog. J. 3, 2011, 67-71.
76. Kosmulalage KS, Zahid CS, Udenigwe S, Ahktar A, Ata R, Samarasekera. Glutathione S-transferase, acetyl cholinesterase inhibitory and antibacterial activities of chemical constituents of *Barleria prionitis*. Z. Naturforsch. 62b, 2007, 580-586.
77. Ata A, Bosch VDSA, Harwanik, DJ, Pidwinski, GE. Glutathione S-transferase and acetylcholinesterase-inhibiting natural products from medicinally important plants, Pure Appl. Chem. 79, 2007, 2269-2276.
78. Singh B, Chandan BK, Prabhakar A, Taneja SC, Singh J, Qazi GN. Chemistry and hepatoprotective activity of an active fraction from *Barleria prionitis* Linn. in experimental animals. Phtother. Res. 19, 2005, 391-404.
79. Bhavana BM, Bothara, SB. Investigation of Antihypertensive Activity of Leaves of *Barleria prionitis* in Doca Salt Induced Hypertensive Rat. Int. J. Pharma Sci. Res. 18(2), 2013, 17-19.
80. Ghule BV, Yeole PG. *In vitro* and *in vivo* immunomodulatory activities of iridoids fractions from *Barleria prionitis*. J. Ethano Pharma. 141(1), 424-431.
81. Musale SB, Jagtap VA, Patil MS, Chittam KP, Wagh, KP. Diuretic activity of *Barleria prionitis* Linn. Flower extract. Int. J. Drug Discovery Herbal Res. 1, 2011, 20-21.
82. Gangopadhyay A, Malakar J, Ghosh A, Pramanik G, Karmakar S. Comparative Antibacterial study of *Barleria prionitis* Linn. leaf extracts. Int. J. Pharma. & Biological Archive. 3(2), 2012, 391-3.
83. Marya BH, Bothara SB. Investigation of antihypertensive activity of leaves of *Barleria prionitis* in Doca salt induced hypertensive rats. Int. J. Pharm. Sci. Res. 18(2), 2013, 17-19.
84. Atif M, Rahman SA, Ahmed MI, Mahmood SB, Azharuddin M. Anticataract potential of *Barleria prionitis*: *in vivo* study. Int. J. Pharmacy & Pharma Sci. 7(2), 2015, 100-105.
85. Manjusha VK, Singh S. Gastroprotective activity of methanol leaves extract of *Barleria prionitis* Linn. on Ethanol and Indomethacin Induced Ulcer in Rats. Brit. J. Pharma. Res. 3 (4), 2013, 817.
86. Jeyasankar A, Premalatha S, Krishnappa K, Elumalai K. Larvicidal activity of *Barleria prionitis* L (Acanthaceae) against Japanese encephalitis vector, *Culex tritaeniorhynchus* Giles (Diptera: Culicidae). Int. J. Interdiscip Res. Rev. 1(2), 2013, 116-20.
87. Basaligappa LH, Chandravadan H. A survey of plant in Gujarat, India, for alkaloids, saponins and tannins. U.S.D.A. Forest service Research Paper Ne. 1971, 201.
88. Bharat BA, Haruyo I, Prachi G, Priya W, Gautam S, Indra DB, Manoj KP, Shishir S, Muraleedharan GN. From traditional Ayurvedic medicine to modern medicine, identification of therapeutic targets for suppression of inflammation and cancer. Expert opinion on therapeutic targets. 10(1), 2006, 87-118.
89. Harborne JB, Subramaniam SS, Nair AGR. Scutellarein 7-Rhamnosylglucoside from *Barleria prionitis*. Phytochem. 10, 1971, 2822-2823.
90. Gupta HM, Saxena VK. A new acylated luteolin-7-O- β -D-glucoside from the roots of *Barleria prionitis* (Linn.). Natl. Acad. Sci. Lett. 7, 1984, 187-189.
91. Taneja SC, Tiwari HP. Structures of two new iridoids from *Barleria prionitis* Linn. Tetrahedron Lett. 16, 1975, 1995-1998.
92. Van A, Van DB, Tromp DJ, Griffioen DHMN, Van BW, Vader V Bast A. Structural aspects of antioxidant activity of flavonoids. Free Radical Boil Med. 20, 1996, 331-342.

Source of Support: Nil, Conflict of Interest: None.

