

Research Article



Phytochemical Screening and Pharmacological Assessment of *Justicia aurea* Grown in Bangladesh

A. N. M. Shofi Uddin, Beton Chakma, Banani Mondal, Sarker Ramproshad, Md. Anisuzzman, Bishwajit Bokshi*, Samir Kumar Sadhu
Pharmacy Discipline, Life Science School, Khulna University, Khulna-9208, Bangladesh.

*Corresponding author's E-mail: bokshi06@pharm.ku.ac.bd

Received: 04-05-2022; Revised: 22-07-2022; Accepted: 29-07-2022; Published on: 15-08-2022.

ABSTRACT

Plants are the major source of therapeutic compounds and have huge applications in Pharma Industry. To identify new sources of therapeutic compounds, we studied acute toxicity, antihyperglycemic, anti-inflammatory, and anticoagulant activities of the polar and nonpolar fractions of *Justicia aurea* (*J. aurea*). Our initial phytochemical screening of *J. aurea* exhibited the presence of numerous secondary metabolites such as reducing sugars, glycosides, tannins, gums, alkaloids, phenolic compounds, and carbohydrates which might be primarily responsible for its medicinal properties. In the acute toxicity test, the result showed no toxicity up to 2000 mg/kg. In the antihyperglycemic activity test, administration of petroleum ether fraction and water fraction of *J. aurea* extract to glucose-loaded mice at 250 and 500 mg/kg body weight (BW) reduced the blood glucose levels compared to control mice. Between two fractions, the water fraction was found to exhibit better potentiality at 500 mg/kg BW. In the anti-inflammatory assay, at 250 and 500 mg/kg water fraction exhibited anti-inflammation by 20.00% and 31.67%, respectively and petroleum ether fraction by 13.33% and 18.00%, respectively. In anticoagulant activity, the test indicated that water fraction has minor anticoagulant potentiality compared to control. In conclusion, the polar water fraction shows better medicinal properties than the nonpolar petroleum ether fraction of *J. aurea*.

Keywords: *Justicia aurea*, Acanthaceae, acute toxicity, antihyperglycemic, anti-inflammatory, anticoagulant.

QUICK RESPONSE CODE →

DOI:
10.47583/ijpsrr.2022.v75i02.021



DOI link: <http://dx.doi.org/10.47583/ijpsrr.2022.v75i02.021>

MATERIALS AND METHODS

Plant collection and extraction

The whole plant of *J. aurea* was collected from Chalna, Khulna in March 2016. The collected plant was identified by the experts of Pharmacy Discipline, Khulna University, Bangladesh.

Plant extraction

Plant extraction is a method to extract certain chemical components present in the plants. The plant parts of *J. aurea* were isolated from unwanted materials and washed with fresh water. Next, the plant parts were air-dried which normally takes 7 days, and were powdered with a suitable grinder. The powder material of *J. aurea* was stored in an airtight glass container at room temperature in a dark, dry environment. Later, 460 g of powdered material was placed in a clean flat-bottomed glass container and soaked in 1600 ml 95.00% ethanol. Then the powdered materials were sealed for 14 days with continuous stirring. 14 days later, the whole mixture was filtered with clean white cotton followed by Whatman filter paper to remove unwanted debris. Finally, the solvent was evaporated to yield 2.83% w/w of the crude extract. Thereafter, the crude extract was separated into petroleum ether, and water fractions.

Phytochemical screening

The preliminary phytochemical studies revealed the existence of various chemical groups in test samples. Various tests on crude extracts were performed based on the established methods⁶⁻⁹.

INTRODUCTION

Natural products play a key role in drug discovery and over one-third of drugs approved by the Food and Drug Administration (FDA) was developed from natural products and their derivatives¹. Plants contain bioactive natural products and serve as raw materials to produce drugs². The genus *Justicia* (Acanthaceae) contains 600 species that are primarily habitat to tropical and pantropical regions. These species have been reported with various biological activities where antidiabetic, anti-inflammatory, and antiviral activities are the most noteworthy³⁻⁵. *J. aurea* which is also known as Yellow Jacobinia is an evergreen suburb with ovate leaves (10-30 cm). It is almost 10 feet tall and 2 to 3 feet wide. It has bright yellow flowers which are blooming from mid-summer to late summer and rains. This plant is best grown in well-drained soils and moist areas⁴. Upon literature survey, no work has been reported on the antihyperglycemic, anti-inflammatory, and anticoagulant activity of *J. aurea*. Therefore, this project aimed to evaluate the mentioned activities along with the phytochemical screening of polar and nonpolar fractions of crude extract and make a comparative feature.



Experimental animal

To experiment with the different fractions of plant extract, we chose Swiss-albino young mice aged 28-35 days (average weight of 20-30g). They were always kept in the animal house of Pharmacy Discipline, Khulna University, Khulna, Bangladesh at room temperature and relative humidity of 55-65% with 12 hours of daylight and 12 hours of darkness. These mice were fed with ICDDRB formulated food and water. The mice were cared for and handled following internationally accepted standard laboratory protocols and guidelines. The animals were regularly checked before experimentation.

Acute toxicity test

Acute toxicity test of *J. aurea* extract was carried out using the method of Devnath et al¹⁰. At first, 500 mg/kg BW was given orally using a feeding needle to three animals which were initially weighed and marked, and then 1000, 1500, and 2000 mg/kg BW were used¹¹. Then individual observations were started during the first half an hour and periodically during the first 24 hours. After 24 hours, if the animals were alive and does not show any adverse effects then the next dose was administered to another three animals.

Antihyperglycemic activity test

The antihyperglycemic activity was studied using the oral glucose tolerance test (OGTT)¹². Forty overnight fasted mice were distributed into eight groups comprising five in each group. The control group was administered tween-80 (2%) in water at 10 ml/kg BW. The standard group was administered glibenclamide at 5 mg/kg BW. The rest of the groups were administered the petroleum ether and water fraction at 250 and 500 mg/kg BW. The tail tips of mice were dissected and the blood sample was taken to measure blood glucose levels using a glucometer. The povidone-iodine ointment was put on to the injured area to prevent infection/inflammation on the dissected tail end. The control, standard, and fractions of extracts were fed using a feeding needle in the respective test group¹³. Half an hour later, glucose was fed at the dose of 2 g/kg BW to all test groups. After feeding, the levels of blood glucose were recorded at 60 minutes intervals from 30-150 minutes.

Anti-inflammatory activity test

The anti-inflammatory potentiality of *J. aurea* fractions was carried out by the method of Jahan et al with slight modification¹⁴. In this experiment, 30 young Swiss albino mice were randomly allocated into 6 groups having 5 in each group. Group I (control group) was given 0.9% sodium chloride solution and Group II (standard) was given ibuprofen (100 mg/kg) orally. Group III and group IV were fed water fractions and Group V and VI were administered petroleum ether fractions at the doses of 250 and 500 mg/kg BW orally. One hour later, each mouse got 20 µl of xylene on the anterior and posterior surfaces of the right ear lobe. The left ear was accounted for as a control.

Xylene-treated mice were sacrificed 60 minutes after administration. Circular sections were taken with the help of a cork borer having a 3 mm diameter and weighed. The level of percentage inhibition was measured following the equation:

$$\text{Inhibition (\%)} = (1 - Et/Ec) \times 100$$

Where, Et = average edema of the experimental group and

Ec = average edema of the control group

Anticoagulant test

0.2 ml of plasma, 0.1 ml of different crude fractions, and 0.3 ml of CaCl₂ (25mM) were put together in fusion test tubes. For negative control, 0.1 ml of 0.9% saline, 0.2 ml plasma, and 0.3 ml of CaCl₂ (25mM) were taken to the second test tube. For positive control, 0.1 ml warfarin, 0.2 ml plasma, and 0.3 ml CaCl₂ (25mM) were taken to another test tube. All test tubes were heated in a water bath at 37°C. The clotting time (prothrombin time) was determined with a stopwatch by tilting the test tubes every 5 seconds. Each of the tests was performed thrice and the average scavenging time was noted^{15, 16}.

RESULTS

Phytochemical tests

The phytochemical study of *J. aurea* crude extract revealed the existence of several natural compounds summarized in Table 1. These types of phytoconstituents might be contributors to different bioactivities¹⁷. Therefore, further biological studies were performed for assessing their antihyperglycemic, anti-inflammatory, and anticoagulant activities.

Table 1: Outcome of chemical group test of *J. aurea* crude extract

Phytochemical groups	Outcome
Reducing sugars	+
Glycosides	+
Tannins	+
Flavonoids	+
Saponins	-
Gums	+
Protein-xanthoproteins	+
Alkaloids	+
Acidic compounds	-
Carbohydrate	+
Phenolic compounds	+

+ = Presence; - = Absence

Acute toxicity test

The result showed that no mice died at 500, 1000, 1500, and 2000 mg/kg BW). The doses used in different tests to evaluate the pharmacological profile of *J. aurea* are a



minimum of 10 times less than the toxic dose as observed. So, it ensures that the toxic effect of the crude extract does not interfere with the result during pharmacological investigations.

Antihyperglycemic activity test

Administration of water and petroleum ether fractions of *J. aurea* extract to glucose-loaded mice at 250 and 500 mg/kg BW reduced minor blood glucose levels compared to control and the effect was comparable with the standard glibenclamide at 150 min. Petroleum ether fraction was found to exhibit a significant effect at 250 mg/kg whereas water fraction showed a significant role at 500 mg/kg (Table 2).

Anti-inflammatory activity test

The results of the test (Table 3) showed that at 250 and 500 mg/kg water fraction exhibits inhibition of inflammation by 20% and 31.67%, respectively, and petroleum ether

fraction shows inhibition of inflammation by 13.33% and 18%, respectively. Here, the standard drug ibuprofen inhibition was found to be 70% at 100 mg/kg BW.

So, it can be claimed that the anti-inflammatory activity of various fractions of *J. aurea* extract was significant compared to the negative control.

Anticoagulant test

Petroleum ether and water fractions of *J. aurea* were subjected at the dose of 350 and 175 mg/ml to carry out the anticoagulant potentialities. The petroleum ether did not exhibit any significant rise in the mean clotting time. However, the water fraction at 350 mg/ml significantly increased coagulation time by 4.98 minutes compared to the control (0.9% saline) clotting time of 3.36 minutes which indicates that the water fraction has lower anticoagulant activity (Table 4).

Table 2: Anti-hyperglycemic effect of fractions of *J. aurea* extract

Group	Fasting state	30 min	90 min	150 min
Control	5.93±.29	10.33±.17	9.13±.17	7.85±.13
Standard	5.70±.18	3.38±.17***	2.43±.17***	3.08±.18***
Petroleum ether fraction (250 mg/kg)	6.30±.37	10.63±.28	9.03±.67**	8.55±.30*
Petroleum ether fraction (500 mg/kg)	5.58±.25	10.23±.20	9.33±.58	7.60±.43
Water fraction (250 mg/kg)	5.57±.19	9.00±.18*	8.28±.26	7.10±.30
Water fraction (500 mg/kg)	5.63±.15	8.70±.40**	7.30±.51***	6.28±.21**

Values are expressed as mean ± standard error of the mean (n = 3); * indicates P<0.05, ** indicates P<0.01, and *** indicates P<0.001 when compared with control.

Table 3: Anti-inflammatory effect of fractions of *J. aurea* extract

Test group	Dose	% Inhibition of inflammation
Negative control (normal water)	10 ml/kg	0
Positive Control (ibuprofen)	100 mg/kg	70±0.029***
Water fraction	250 mg/kg	20±0.041**
	500 mg/kg	31.67±0.063**
Petroleum ether fraction	250 mg/kg	13.33±0.041*
	500 mg/kg	18±0.075*

Values are expressed as mean ± standard error of the mean (n = 3); * indicates P<0.05, ** indicates P<0.01, and *** indicates P<0.001 when compared with control.

Table 4: Anticoagulant effect of fractions of *J. aurea* extract

Groups	Concentration of sample	Average time of coagulation (min)
Control	0.9% saline	3.36±0.09***
Warfarin	5 mg/ml	62.86±2.87***
Petroleum ether fraction	175 mg/ml	3.11±0.02
	350 mg/ml	3.44±0.03
Water fraction	175 mg/ml	3.74±0.34
	350 mg/ml	4.98±0.22***

Values are expressed as mean ± standard error of the mean (n = 3); * indicates P<0.05, ** indicates P<0.01, and *** indicates P<0.001 when compared with control.



DISCUSSION

To determine the toxicity and establish a safe dose of different fractions of crude plant extract, toxicity studies are carried out in various test animals. Since *J. aurea* has not been previously studied in mice model, the acute toxicity study was carried out following the Organisation for Economic Co-operation and Development (OECD) recommendations to evaluate its toxicity and identify the safe dose that might be used in further studies^{18, 19}. In this study, the water and petroleum ether fractions appear to be non-toxic up to 2000 mg/kg.

A very little number of compounds that resemble antidiabetic drugs present in *J. aurea* may lower the blood glucose level in mice. The effect of the crude fractions was recorded at 30, 90, and 150 minutes after glucose administration. The observed reduction in blood glucose after administration of the crude fractions could be due to different mechanisms. As seen with *Mangifera indica* L., compound (s) found within the leaf may decrease glucose absorption in the gut²⁰. Alternately, any bio-active natural product (s) in the crude fractions may lower blood glucose by enhancing the pancreatic insulin release or enhancing glucose uptake, as has been shown in experiments using the extract from *Artemisia* and *Ageratum conyzoides* L.^{21, 22}. Another probable mechanism can be the rise of peripheral glucose consumption induced by the extract, as has been exhibited by the extract of *Sapindus trifoliatus* L.²³. There have been reports of antidiabetic action of herbal extracts containing flavonoids and tannins²⁴. Based on this fact, it may be postulated that the flavonoids or tannins present in this plant may be the cause of the observed decrease in blood sugar levels.

Inflammation is a complicated series of reparative and protective reactions to tissue injury whatever the cause - infection, auto-immune response, or mechanical injury²⁵. Histopathologically, extreme vasodilation, edematous skin alterations, and infusion of inflammatory cells are recognized as indications of acute inflammation after topical xylene application. Xylene-induced ear edema model is partly related to substance P, an undecapeptide that is widely distributed in the central and peripheral nervous systems and serves as a neurotransmitter or a neuromodulator in a range of physiological conditions²⁶. When substance P is released from sensory neurons, it causes plasma extravasations and vasodilatation, which suggests that it plays a part in the neurogenic inflammation that results in the swelling of the ear in mice. By converting arachidonic acid into prostaglandins, the Cyclooxygenase (COX) enzyme is recognized to have a significant role in the development of the later stage of inflammation²⁷. Many non-steroidal anti-inflammatory drugs (NSAIDs), including ibuprofen, are thought to have this enzyme as a target and can reduce ear edema at a later stage after xylene treatment. Because the release of free radicals is known to result in tissue damage during the inflammatory process, free radical scavenging agents also play a part in inflammation²⁸. It is believed that flavonoids and phenolics

work by inhibiting the production or activation of free radicals²⁹. In our study, results exhibited that both fractions of *J. aurea* significantly subdued ear edema compared to the control group.

The hemostatic processes are designed to stop bleeding at the site of damage by creating a hemostatic plug, which is eventually eliminated once healing is complete³⁰. Normal physiology maintains a careful balance between these systems, and bleeding or thrombosis result when one mechanism is inadequate or overactive³¹. Platelets, blood vessels, coagulation factors, plasma inhibitors, and the fibrinolytic system are only a few of the components that maintain physiology³². Blood clotting, which is the main cause of heart attacks and strokes, is inhibited by anticoagulant medications³³. When there is a high risk of blood clots, anticoagulant medications may be taken³⁴. Since anticoagulants are prescribed for cardiac issues, hence, instead of depending on blood thinners, physicians can move to herbal remedies. According to certain reports, antioxidants can prevent oxidative stress, hepatocellular damage, and problems associated with blood coagulation and hematology²⁷. As *J. aurea* is abundant with antioxidants, which could be the contributors to its anticoagulant action.

CONCLUSION

It can be concluded that both nonpolar and polar parts of *Justicia aurea* extract possess variable biological potentialities. However, the water fraction containing polar components showed better antihyperglycemic, anti-inflammatory, and anticoagulant effects than the petroleum ether fraction containing nonpolar components. Further studies are required to isolate and identify the secondary metabolite(s) liable for these biological activities.

Acknowledgements: The authors are grateful to Pharmacy Discipline, Khulna University for providing the laboratory and chemical facilities required to conduct this research.

REFERENCES

- Xu J-B, Li Y-Z, Huang S, Chen L, Luo Y-Y, Gao F, et al. Diterpenoid alkaloids from the whole herb of *Delphinium grandiflorum* L. *Phytochemistry*. 2021;190:112866.
- Ramproshad S, Afroz T, Mondal B, Khan R, Ahmed S. Screening of phytochemical and pharmacological activities of leaves of medicinal plant *Plumeria rubra*. *International journal of research in pharmacy and chemistry*. 2012;2(4):1001-7.
- Ortiz-Andrade R, Cabañas-Wuan A, Arana-Argáez VE, Alonso-Castro AJ, Zapata-Bustos R, Salazar-Olivo LA, et al. Antidiabetic effects of *Justicia spicigera* schltld (acanthaceae). *Journal of ethnopharmacology*. 2012;143(2):455-62.
- Corrêa GM, Alcântara AFdC. Chemical constituents and biological activities of species of *Justicia*: a review. *Revista Brasileira de farmacognosia*. 2012;22:220-38.
- Anyasor GN, Okanlawon AA, Ogunbiyi B. Evaluation of anti-inflammatory activity of *Justicia secunda* Vahl leaf extract using in vitro and in vivo inflammation models. *Clinical Phytoscience*. 2019;5(1):1-13.



6. Sarker SD, Nahar L. An introduction to natural products isolation. Natural products isolation. 2012:1-25.
7. Evans W. Trease and Evans Pharmacognosy. 15th edition. Edinburgh, Saunders. 2002;249.
8. Kundu P, Debnath SL, Sadhu SK. Exploration of Pharmacological and Toxicological Properties of Aerial Parts of *Blumea lacera*, a Common Weed in Bangladesh. Clinical Complementary Medicine and Pharmacology. 2022;2(3):100038.
9. Sultana MS, Golder M, Biswas B, Karmakar UK, Bokshi B, Alam MJ, et al. Antioxidative and Antidiabetic Potentials of the Pneumatophores of *Heritiera fomes* Buch. Ham. Dhaka University Journal of Pharmaceutical Sciences. 2022:283-91.
10. Devnath HS, Ahmed MI, Medha MM, Islam MN, Biswas RP, Islam MA, et al. HPLC Analysis and Antimicrobial, Antidiarrheal and Antihyperglycemic Properties of *Eurya acuminata* along with in silico Profiles. Phytomedicine Plus. 2022;2(3):100291.
11. UA O OD, Udokang N, Udobang J, Ekpenyong C. Oral administration of aqueous leaf extract of *Ocimum gratissimum* ameliorates polyphagia, polydipsia and weight loss in streptozotocin-induced diabetic rats. American Journal of Medicine and Medical Sciences. 2012;2(3):45-9.
12. Medha MM, Devnath HS, Biswas B, Bokshi B, Sadhu SK. In silico profiling of analgesic and antihyperglycemic effects of ethanolic leaves extract of *Amischotholype mollissima*: evidence from in vivo studies. Saudi Journal of Biological Sciences. 2022:103312.
13. Andrikopoulos S, Blair AR, Deluca N, Fam BC, Proietto J. Evaluating the glucose tolerance test in mice. American Journal of Physiology-Endocrinology and Metabolism. 2008;295(6):E1323-E32.
14. Jahan T, Kundu P, Sultana T, Saha L, Chakraborty A, Islam MA, et al. Phytochemical investigation and assessment of pharmacological properties of leaves of *Duabanga grandiflora*. J Med Plants. 2021;9:25-32.
15. Prasad S, Kumar S, Patel K, Dumater C, Vajpeyee S, Bhavsar V. To investigate the action of ginger-juice *Zingiber officinale* roscoe (Zingiberaceae) on blood coagulation process. International Journal of Pharma Sciences and Research. 2012;3(7):407-15.
16. Dey P, Bhakta T. Evaluation of in vitro anticoagulant activity of *Molineria recurvata* leaf extract. J Nat Prod Plant Resour. 2012;2(6):685-8.
17. Shaikh JR, Patil M. Qualitative tests for preliminary phytochemical screening: An overview. International Journal of Chemical Studies. 2020;8(2):603-8.
18. Kifayatullah M, Mustafa MS, Sengupta P, Sarker MMR, Das A, Das SK. Evaluation of the acute and sub-acute toxicity of the ethanolic extract of *Pericampylus glaucus* (Lam.) Merr. in BALB/c mice. Journal of Acute Disease. 2015;4(4):309-15.
19. Saganuwan S. Toxicity studies of drugs and chemicals in animals: an overview. Bulgarian Journal of Veterinary Medicine. 2017;20(4):22-29.
20. Bhowmik A, Khan LA, Akhter M, Rokeya B. Studies on the antidiabetic effects of *Mangifera indica* stem-barks and leaves on nondiabetic, type 1 and 2 diabetic model rats. ||| Bangladesh Journal of Pharmacology. 2009;4(2):110-4.
21. Farjou I, Avadai T, Karim K. Hyperglycemia-induced alterations in brain GABA and seizure threshold. IRCS medical science-biochemistry. 1985;13(7):645-6.
22. Nyunaï N, Njikam N, Abdenneb E, Mbafor J, Lamnaouer D. Hypoglycaemic and antihyperglycaemic activity of *Ageratum conyzoides* L. in rats. African Journal of Traditional, Complementary and Alternative Medicines. 2009;6(2):44-49.
23. Sahoo P, Padhy K, Pradhan D, Tripathy G, Bhoi R, Pattanayak S, et al. Antidiabetic and antioxidant activity of ethanolic extract of *Sapindus trifoliatus* Linn. International Journal of Pharma and Bio Sciences. 2010;1(2):31-37.
24. Suba V, Murugesan T, Arunachalam G, Mandal S, Saha B. Anti-diabetic potential of *Barleria lupulina* extract in rats. Phytomedicine. 2004;11(2-3):202-5.
25. Chapman CR, Tuckett RP, Song CW. Pain and stress in a systems perspective: reciprocal neural, endocrine, and immune interactions. The Journal of Pain. 2008;9(2):122-45.
26. Agbaje E, Fageyinbo M. Evaluating Anti-Inflammatory activity of aqueous root extract of *Strophanthus hispidus* DC.(Apocynaceae). International Journal of Applied Research in Natural Products. 2012;4(4):7-14.
27. Inoue H, Nagata N, Koshihara Y. Profile of capsaicin-induced mouse ear oedema as neurogenic inflammatory model: comparison with arachidonic acid-induced ear oedema. British journal of pharmacology. 1993;110(4):1614-20.
28. Kehrer JP, Klotz L-O. Free radicals and related reactive species as mediators of tissue injury and disease: implications for health. Critical reviews in toxicology. 2015;45(9):765-98.
29. Galati G, O'brien PJ. Potential toxicity of flavonoids and other dietary phenolics: significance for their chemopreventive and anticancer properties. Free radical biology and medicine. 2004;37(3):287-303.
30. Pourshahrestani S, Zeimaran E, Kadri NA, Mutlu N, Boccaccini AR. Polymeric hydrogel systems as emerging biomaterial platforms to enable hemostasis and wound healing. Advanced Healthcare Materials. 2020;9(20):2000905.
31. Ismail H, Mirza B. Evaluation of analgesic, anti-inflammatory, anti-depressant and anti-coagulant properties of *Lactuca sativa* (CV. Grand Rapids) plant tissues and cell suspension in rats. BMC complementary and alternative medicine. 2015;15(1):1-7.
32. Chan AK, Paredes N. The coagulation system in humans. Haemostasis: Springer; 2013. p. 3-12.
33. Mine Y, Wong AHK, Jiang B. Fibrinolytic enzymes in Asian traditional fermented foods. Food Research International. 2005;38(3):243-50.
34. Dahlbäck B. Blood coagulation and its regulation by anticoagulant pathways: genetic pathogenesis of bleeding and thrombotic diseases. Journal of internal medicine. 2005;257(3):209-23.

Source of Support: The author(s) received no financial support for the research, authorship, and/or publication of this article.

Conflict of Interest: The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

For any question relates to this article, please reach us at: globalresearchonline@rediffmail.com
New manuscripts for publication can be submitted at: submit@globalresearchonline.net and submit_ijpsrr@rediffmail.com

