

Research Article



In vitro Free Radical Scavenging and Antioxidant Effect of *Ocimum basilicum*

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ABSTRACT

Oxidative stress has been identified as the root cause of the development and progression of several diseases. Supplementation of exogenous antioxidants or boosting endogenous antioxidant defenses of the body is the most promising way of combating the detrimental effects of reactive oxygen species (ROS) induced oxidative damage. *Ocimum basilicum* L (*Lamiaceae*), a medicinally important plant commonly known as "Holy basil" found to contain many biological properties. The present study is aimed at evaluating the free radical scavenging potentials of *Ocimum basilicum* leaf extract. The antioxidant nature of the leaves extract was proved from Free radical scavenging was determined by using 1,1-diphenyl-2-picrylhydrazyl (DPPH), ABTS (2,2'-azinobis (3ethylbenzthiazoline-6-sulphonic acid), ferric reducing antioxidant power (FRAP), nitric oxide scavenging assay (NO), reducing power, hydroxy radical scavenging assay, superoxide radical scavenging (SOD), hydrogen peroxide radical assay, metal chelating activity as well as phosphomolybdenum assay. From the results obtained, *O.basilicum* leaves extract can be considered as a therapeutic agent for the treatment of free radical mediated diseases.

Keywords: *Ocimum basilicum*, Phytochemicals, Free radicals, Antioxidant property.

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INTRODUCTION

Oxidative stress has been identified as a major causative factor in the development and progression of several life threatening diseases, including diabetes mellitus, cancer, neurodegenerative and cardiovascular disease. During metabolic process and exposure to external environment, a large amount of free radicals is generated in the human body which pose a major influence on biological macromolecules such as proteins, fatty acids and nucleic acids causing oxidative damage on cells or tissues or even resulting in gene mutation. Free radicals at high concentration in human body cause oxidative stress, thus destroying the internal redox balance and causing a variety of chronic diseases, even premature senility. In addition, supplementation with exogenous antioxidants or boosting of endogenous antioxidant defenses of the body has been found to be a promising method of countering the undesirable effects of oxidative stress¹.

Natural antioxidants of plant origin, especially through foods are essential to eradicate a number of free radical mediated diseases². In the series of various medicinal plants, *Ocimum basilicum*. Plays a significant role for its folkloric use in the treatment of various ailments. *Ocimum*

basilicum L (*Lamiaceae*) also commonly known as "Holy basil" have possessed different biological effects. The oil of the plant contains eugenol, methyl eugenol, citral, and methyl chavicol³. *O. Basilicum* leaves are used as antispasmodic, carminative, digestive, stomachic, and tonic^{4,5}.

O. basilicum produces triterpenoids, polyphenols, steroids, and phenylpropanoids some of which, such as baseball, ocimol, basilimoside, rosmarinic acid, hydroxycinnamic acids, oleanolic acid, and betulinic acid which possess various biological properties^{6,7}. *Ocimum* sp., is shown to exert antibactericidal, antiinflammatory, antioxidative, antiulcer, antidiarrheal, chemopreventive and hypoglycemic properties⁸⁻¹³. The observed effects of the plants might be due to its antioxidant power which in turn is attributed to the presence of flavonoids and polyphenols¹⁴. In the absence of systematic studies in the literature, the present study was aimed to investigate the antioxidant properties of *Ocimum basilicum* leaf extract.

MATERIALS AND METHODS

Plant material- Identification and authentication

Matured *O. basilicum* leaves were selectively removed from the plant in and around areas of Pudussery, Palakkad, Kerala and identified by a plant taxonomist. BSI/SRC/5/23/2022/Tech/630.

Preparation of *O.basilicum* leaves extract

Delipidation and extraction

O. basilicum leaves were washed, dried in a hot air oven at 40°C and subsequently ground in to powder in an electrical grinder. Delipidation was performed with methanol, ethyl



acetate for overnight. Soxhlation was performed with 95% methanol, ethyl acetate evaporated in a rotary evaporator at 40-50°C under reduced pressure. The yield of the leaves extract was around 13.5 % of dry weight.

Free Radical Scavenging Assays

The *in vitro* anti radical scavenging potential of *O.basilicum* leaves extract (100-500µg/ml) was determined using DPPH¹⁵, ABTS¹⁶, FRAP¹⁷, Nitric oxide¹⁸, Reducing power¹⁹, hydroxy radical²⁰ superoxide scavenging²¹, hydrogen

peroxide²², metal chelating activity as well as phosphomolybdenum assay^{23,24}.

Statistical analysis

All the assays were carried out in triplicate. Experimental results are expressed as mean ± standard deviation. The results were analyzed using one-way analysis of variance and the group means were compared using Duncan's multiple range tests using SPSS version 16.

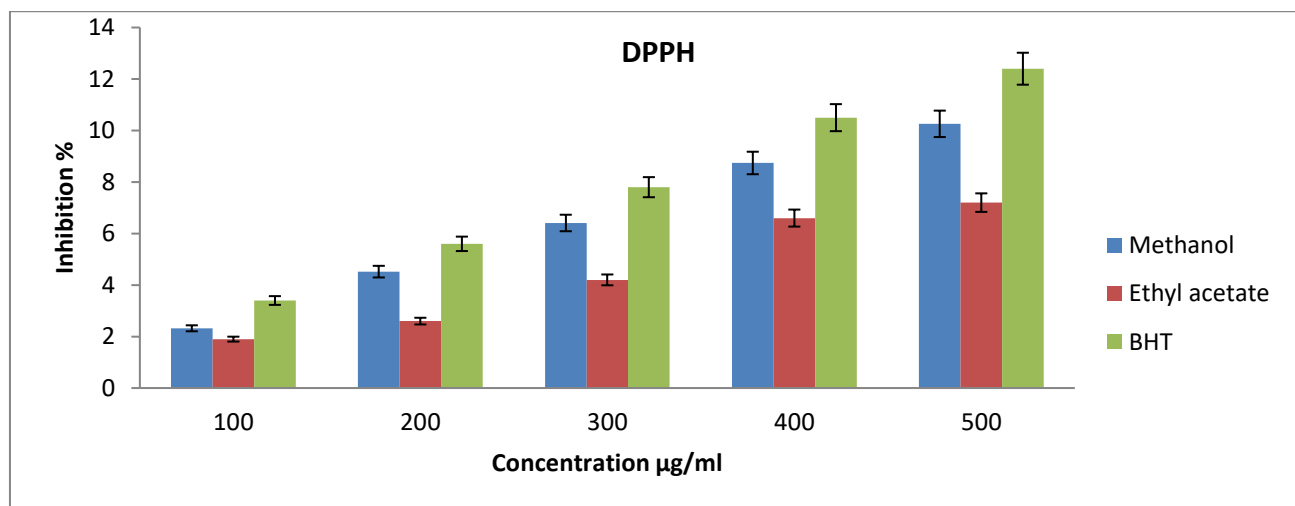


Figure 1: DPPH effect of Methanol and ethyl acetate leaves extract of *Ocimum basilicum*

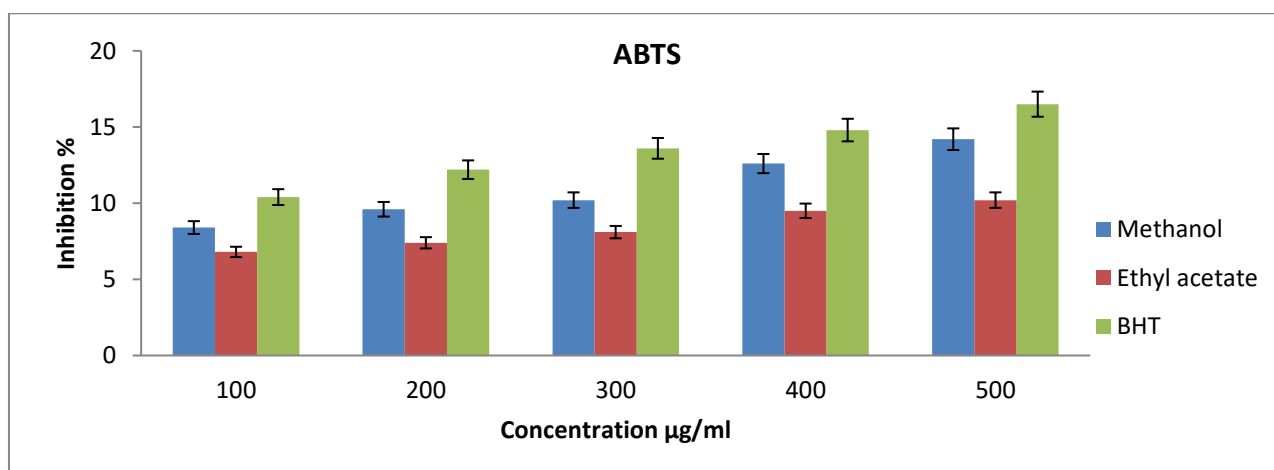


Figure 2: ABTS effect of Methanol and ethyl acetate leaves extract of *Ocimum basilicum*

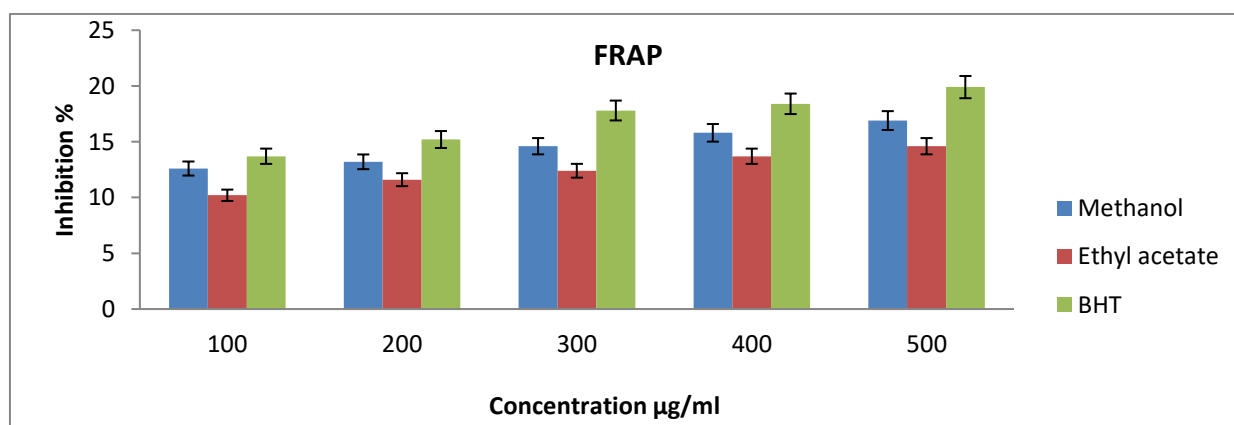


Figure 3: FRAP effect of Methanol and ethyl acetate leaves extract of *Ocimum basilicum*

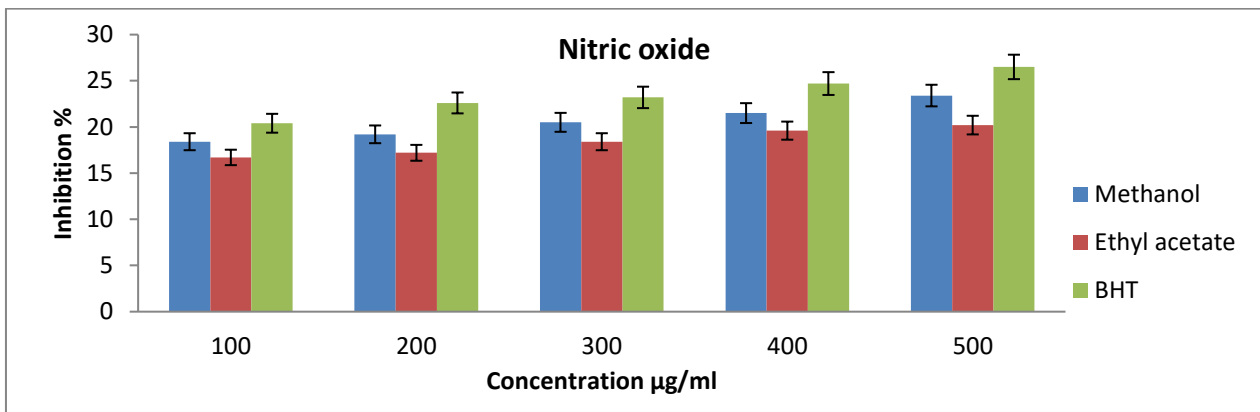


Figure 4: Nitric oxide effect of Methanol and ethyl acetate leaves extract of *Ocimum basilicum*

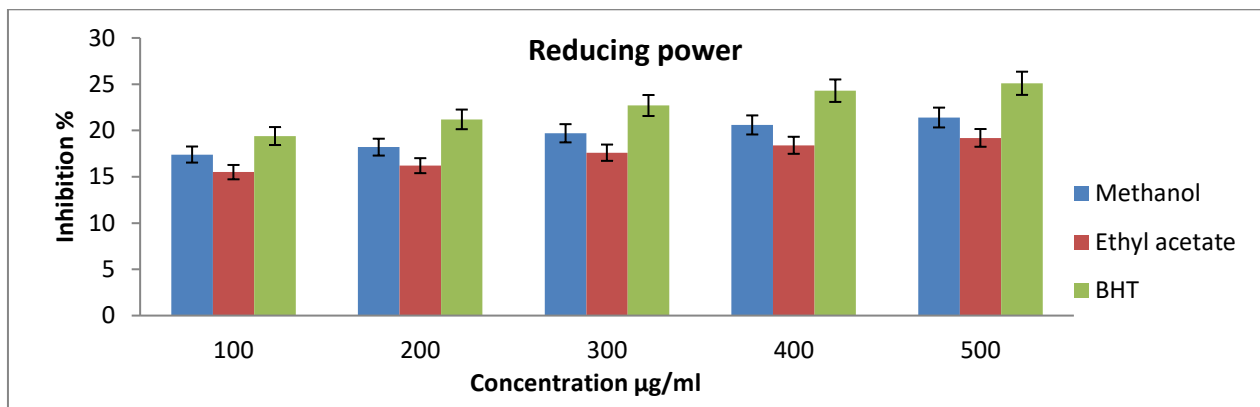


Figure 5: Reducing power effect of Methanol and ethyl acetate leaves extract of *Ocimum basilicum*

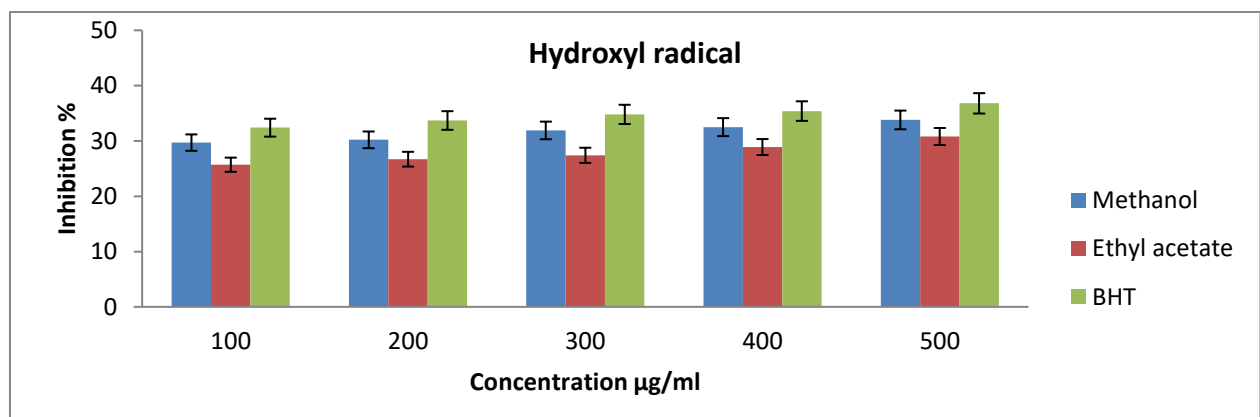


Figure 6: Hydroxyl radical effect of Methanol and ethyl acetate leaves extract of *Ocimum basilicum*

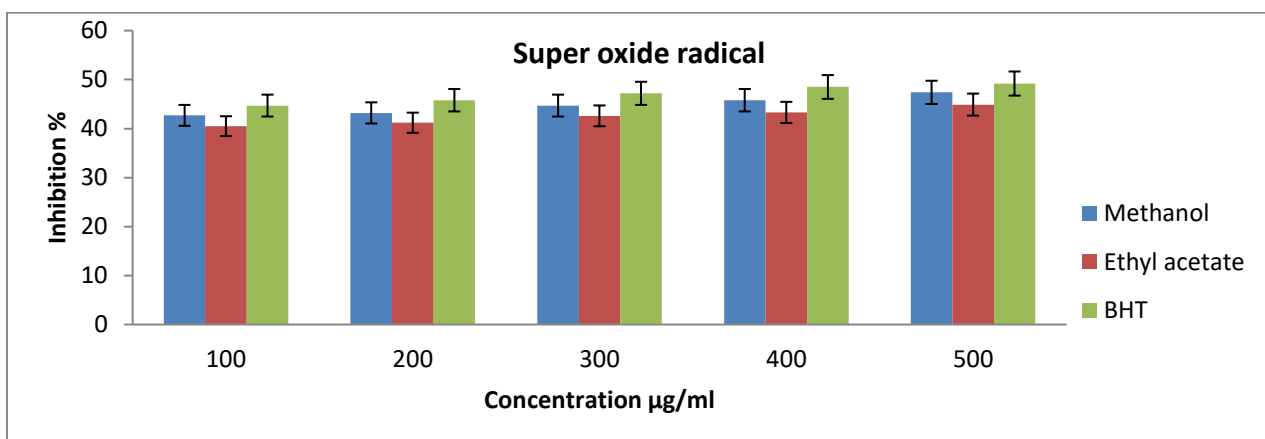


Figure 7: Superoxide radical effect of Methanol and ethyl acetate leaves extract of *Ocimum basilicum*

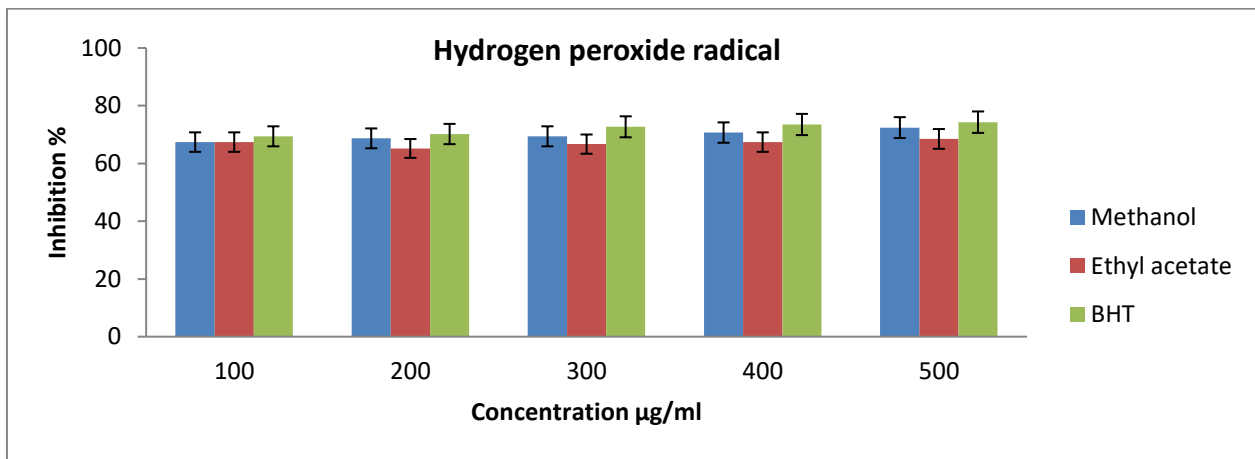


Figure 8: Hydrogen peroxide effect of Methanol and ethyl acetate leaves extract of *Ocimum basilicum*

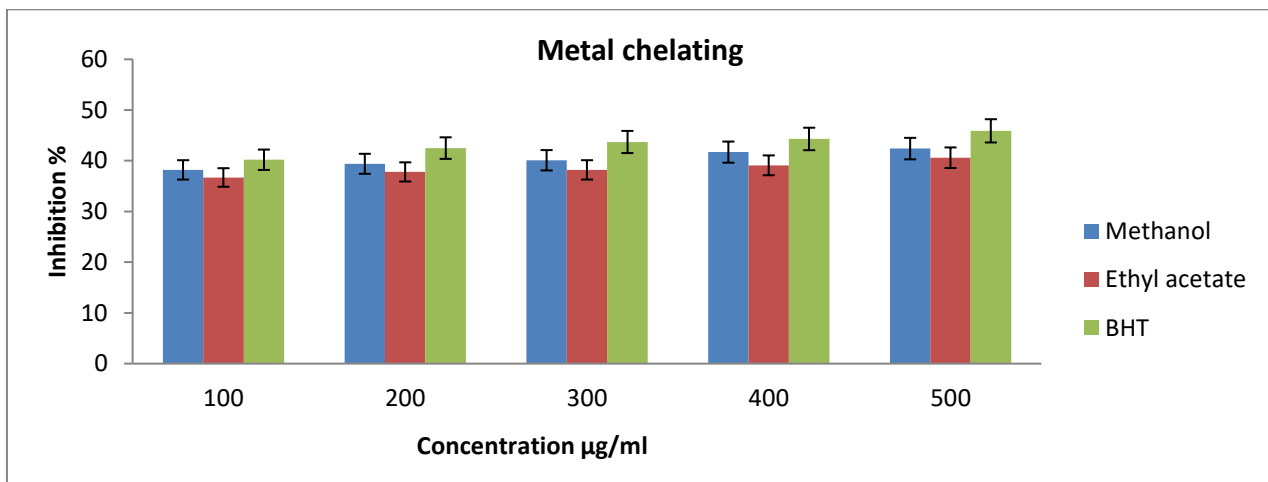


Figure 9: Metal chelating effect of Methanol and ethyl acetate leaves extract of *Ocimum basilicum*

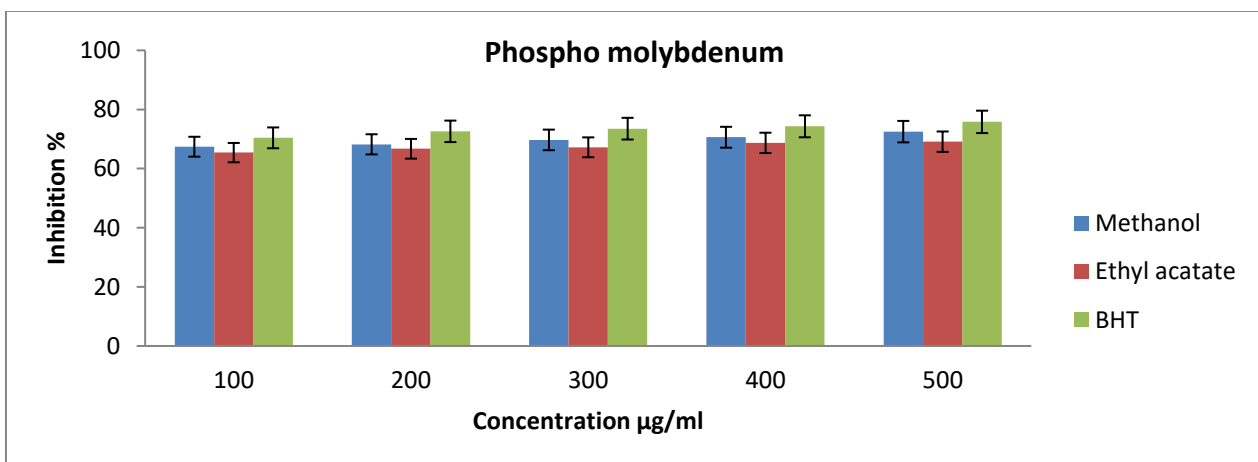


Figure 10: Phospho molybdenum effect of Methanol and ethyl acetate leaves extract of *Ocimum basilicum*

RESULTS

Figure 1 and 2 shows the effect of *O. basilicum* leaves extract methanol, ethyl acetate on the DPPH and ABTS radicals present in the reaction mixtures. The extract at a concentration of 100 -500µg/ml, significantly scavenged of DPPH radicals with an IC₅₀ value of 10.26, 7.2µg/ml and ABTS radicals having IC₅₀ values of 14.2, 10.2µg/ml. BHT values 12.4, 16.5µg/ml.

Figure 3 and 4 shows the effect of the FRAP power of the *O. basilicum* increases extract methanol, ethyl acetate with the increasing concentration was 16.9, 14.6µg/ml and the IC₅₀ value of BHT was 19.9µg/ml. The scavenging of nitric oxide by *O. basilicum* was increased concentration of 23.4, 20.2 µg/ml of *O. basilicum* 50% of nitric oxide generated by incubation was scavenged. The IC₅₀ value of BHT was 26.5µg/ml.

Figure 5 shows the effect the reducing power *O.basilicum* methanol, ethyl acetate was increased in quantity of sample. The *O.basilicum* could reduce the most Fe^{3+} ions, which had a lesser reductive activity than the standard of BHT. The IC_{50} value of *O.basilicum* and BHT was 21.4, 19.2 μ g/ml and 25.1 μ g/ml respectively.

The results for hydroxyl scavenging assay are shown in Fig.6. The concentrations for inhibition were found to be 33.8, 30.8 and 36.8 μ g/ml for the *O.basilicum* and BHT, respectively.

Figure 7 and 8 shows the effect of the superoxide scavenging activity of *O.basilicum* leaves methanol, ethyl acetate extract showed superoxide scavenging activity (IC_{50} = 40.4,44.9 μ g/ml), *O.basilicum* showed concentration dependent activity and the H_2O_2 scavenging effect at a concentration was 72.42, 68.5 μ g/ml. This was comparable to the scavenging effect on the concentration of BHT was 74.3, 49.2 μ g/ml.

Figure 9 and 10 shows the effect of the metal chelating activity and phosphomolybdenum reduction of *O.basilicum* methanol, ethyl acetate to the quantity of the sample. The IC_{50} value of *O.basilicum* was 42.4,40.6 μ g/ml and 45.9 μ g/ml as a standard BHT significantly the phosphomolybdenum 72.5 and 69.1 μ g/ml and 75.8 μ g/ml for the *O.basilicum*, BHT.

DISCUSSION

Phytochemicals, are produced in plants to protect themselves from the environmental stress and infections. Phytochemicals play a preventive role in the treatment of diabetes and cancer²⁵⁻²⁷. Primary metabolites produced in plants are maintained plant cells, while secondary metabolites are responsible for normal growth, development and defense of plants²⁸. These compounds are mostly nitrogen-containing alkaloids or nitrogen-deficient terpenoids and phenolics²⁹. Flavonoids and phenolic acids are biosynthetically derived from the acetate and shikimate pathways (from phenylalanine or tyrosine)^{30,31}. It has been reported that *Ocimum basilicum* L. Contains various compounds such as the flavonoid, alkaloid, phenol and essential oil contains flavonoid compounds with the greatest potential as an antioxidant³².

The antioxidant property of plant confers their free radical scavenging potential their bioactive components and to understand the mechanism of action of their phytoconstituents³³. In the present study, *O. basilicum* leaves scavenge DPPH and ABTS radicals in a concentration dependent manner. The amount of DPPH which is reduced may be estimated by observing a decrease in absorbance at 517 nm. ABTS assay involves reduction of the color intensity of ethanolic solution containing pre-formed radical monocation of ABTS which is generated by oxidation of ABTS with potassium persulfate due to the radical scavenging activity of anti-oxidants present in the plants³⁴. The change in intensity of the color is directly proportional to the antioxidant efficiency of the compound in plant extract. *O. basilicum* leaves extract at a concentration of

1000 μ g/ml, the extract significantly scavenged of DPPH radicals (IC_{50} =10.26,7.2 μ g/ml) and ABTS radicals (IC_{50} =14.2,10.2 μ g/ml)

Antioxidants can be explained as reductants, and inactivators of oxidants³⁵. Some previous studies have also reported that the reducing power may serve as a significant indicator of potential antioxidant activity.. In this study, we used a FRAP assay because it is quick and simple to perform, and the reaction is reproducible and linearly related to the molar concentration of the antioxidant and FRAP assay was used by several authors for the assessment of antioxidant activity of various food product samples^{36, 37}. The reducing power of the *Ocimum basilicum* increases with the increasing concentration was 16.9,14.6 μ g /ml.

Nitric oxide is a free radical produced in mammalian cells, involved in the regulation of various physiological process including neurotransmission, vascular homeostasis, antimicrobial and antitumor activities. However, excess production of NO is associated with several diseases³⁸. *Ocimum basilicum* inhibited nitrite formation in a concentration dependent manner (100-500 μ g/ml). This may be due to the presence of antioxidant principles in the *Ocimum basilicum* which complete with oxygen to react with nitric oxide. The scavenging of nitric oxide 23.4, 20.2 μ g/ml of *Ocimum basilicum* of nitric oxide generated by incubation was scavenged.

The reducing power of the *Ocimum basilicum* was evaluated by the transformation of Fe^{3+} to Fe^{2+} through electron transfer ability, which serves as a significant indicator of its antioxidant activity. Reductions are also reported to react with certain precursors of peroxide, thus preventing peroxide formation³⁹. The presence of antioxidant substances in the compound samples causes the reduction of the Fe^{3+} ferric cyanide complex to the ferrous form. Therefore, Fe^{2+} can be monitored by measuring the formation of Perl's Prussian blue at 700 nm. The IC_{50} value of *Ocimum basilicum* was 21.4,19.2 μ g/ml respectively.

Hydroxyl radical scavenging capacity of *Ocimum basilicum* is directly related to its antioxidant activity⁴⁰. This method involves in vitro generation of hydroxyl radicals using Fe^{3+} /ascorbate/EDTA/ H_2O_2 system using Fenton reaction. The concentrations for inhibition were found to be 33.8, 30.8 μ g/ml for the *Ocimum basilicum* respectively.

Superoxide radicals generated *in vitro* by the system was determined by NBT photo reduction method. The decrease of absorbance at 560 nm with the plant extract indicates the consumption of superoxide anion in the reaction mixture⁴¹. *O. basilicum* leaves extract exhibited a maximum of superoxide scavenging activity (IC_{50} = 47.4, 44.9 μ g/ml).

Hydrogen peroxide is a weak oxidizing agent that inhibits the oxidation of essential thiol (-SH) groups directed by few enzymes. It can probably react with Fe^{2+} and possible Cu^{2+} ions to form hydroxyl radicals⁴². From the results, *Ocimum basilicum* showed concentration dependent activity and the



H₂O₂ scavenging effect at a concentration was 72.4, 68.5µg/ml.

Iron is an essential mineral for normal physiology, but an excess of it may result in cellular injury^{43,44}. The chelating ability of ferrous ions by the *Ocimum basilicum* was estimated by the method: Ferrozine can quantitatively form complexes with Fe²⁺. In the presence of chelating agents, the complex formation is disrupted with the result that the red color of the complex is decreased. The metal chelating activity of *Ocimum basilicum* is present 42.4µg/ml and 40.6µg/ml.

The phosphomolybdenum method is based on the reduction of Mo₆ (VI) to Mo₅ (V) by the antioxidant compounds and the formation of green phosphate/Mo₅ (V) complex with the maximal absorption at 695 nm⁴⁵. The IC₅₀ value of *Ocimum basilicum* was 72.5µg/ml and 69.1µg/ml.

CONCLUSIONS

O.basilicum leaves is rich in biologically active ingredients and minerals of known pharmacological actions. Free radical potential of the *O.basilicum* leaves extract is evident from in vitro antioxidant assays. The antioxidant property might be due to the presence of flavonoids and phenols present in the leaves. Thus, *Ocimum basilicum* leaves can be used for the treatment of free radical mediated diseases such as diabetes mellitus and cancer.

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REFERENCES

- Kasote DM, et al. Significance of antioxidant potential of plants and its relevance to therapeutic applications, *Int J. Biol. Sci*, 2015; 11(8): 982-991.
- Kasote DM, Hegde MV, Katyare SS. Mitochondrial dysfunction in psychiatric and neurological diseases: cause(s), consequence(s), and implications of antioxidant therapy, *Biofactors*, 2013; 39(4): 392- 406.
- Bunrathep S, Palanuvej C, Ruangrunsi N. Chemical compositions and antioxidative activities of essential oils from four *ocimum* species endemic to Thailand, *J. Health. Res*, 2007; 21: 201–203.
- Rahimi R, Shams-Ardekani MR, Abdollahi M. A review of the efficacy of traditional Iranian medicine for inflammatory bowel disease, *World. J. Gastroenterol*, 2010; 16(36): 4504-14.
- Velázquez C, Calzada F, Torres J, González F, Ceballos G. Antisecretory activity of plants used to treat gastrointestinal disorders in Mexico, *J. Ethnopharmacol*, 2006; 103(1): 66-70.
- Siddiqui BS, Aslam H, Ali ST, Begum S, Khattoon N. Two new triterpenoids and a steroidal glycoside from the aerial parts of *Ocimum basilicum*. *Chem. Pharm. Bull (Tokyo)*, 2007; 55(4): 516-9.
- Strazzer P, Guzzo F, Levi M. Correlated accumulation of anthocyanins and rosmarinic acid in mechanically stressed red cell suspensions of basil (*Ocimum basilicum*), *J. Plant. Physiol*, 2011; 168(3): 288-93.
- Chattopadhyay RR. A comparative evaluation of some blood sugar lowering agents of plant origin, *Journal of Ethnopharmacology*, 1999; 67: 367–372.
- Klem MA, Nair MG, Sraassburg GM, and Dewitt DL. Antioxidant and cyclooxygenase inhibitory phenolic compounds from *Ocimum sanctum Linn*, *Phytomedicine*, 2000; 7: 7–13.
- Koga T, Hirota N, and Takumi K. Bactericidal activities of essential oils of Basil and Sage against a range of bacteria and the effect of these essential oils on *Vibrio parahaemolyticus*, *Microbiological Research*, 1999; 154: 267–273.
- Maity TK, Mandal SC, Saha BP, and Pal M, Effect of *Ocimum sanctum* roots extract on swimming performance in mice, *Phytotherapy Research*, 2000; 14: 120–121.
- Offiah VN and Chikwendu UA. Antidiarrheal effects of *Ocimum gratissimum* leaf extract in experimental animals, *Journal of Ethnopharmacology*, 1999; 68: 327–330.
- Uma Devi P, Ganasoundri A, Vindra B, and Srinivasan KK. Radiation protection by the *Ocimum* flavonoids orientin and vicerin: mechanism of action. *Radiation Research*, 2000; 154: 455–460.
- Kaushik G, Satya S, Khandwal RK, and Naik SN. Commonly consumed Indian food materials in the management of diabetes mellitus, *Diabetes and Metabolic Syndrome: Clinical Research and Reviews*, 2008; 5: 31–37.
- Brand-Williams W, Cuvelier ME, Berset C. Use of a free-radical method to evaluate antioxidant activity, *Food. Sci. Technol-Leb*, 1995; 28: 25–30.
- Re R, Pellegrini N, Proteggente A, Pannala A, Yang M, Rice-Evans C. Antioxidant activity applying an improved ABTS radical cation decolorization assay, *Free Radic. Biol. Med*, 1999; 26 (9-10): 1231- 1237.
- Benzie IF1, Strain JJ. The ferric reducing ability of plasma (FRAP) as a measure of "antioxidant power": the FRAP assay, *Anal. Biochem*, 1996; 239(1): 70-6.
- Marcocci L, Maguire JJ, Droylefaix MT, Packer L. The nitric oxide-scavenging properties of Ginkgo biloba extract EGb, *Biochem. Biophys. Res. Commun*, 1999; 201: 748-755.
- Oyaizu M. Studies on product of browning reaction prepared from glucose amine Japanese, *Journal of Nutrition*, 1986; 44: p. 307.
- Gülçin I. Antioxidant activity of caffeic acid (3,4-dihydroxycinnamic acid), *Toxicology*, 2006; 217: p. 213
- Fontana M, Mosca L, Rosei MA. Interaction of enkephalines with oxyradicals, *Biochemical Pharmacology*, 2001; 61: 1253-1257.
- Ruch RJ, Cheng SJ, Klaunig JE. Prevention of cytotoxicity and inhibition of intracellular communication by antioxidant catechins isolated from Chinese green tea, *Carcinogenesis*, 1089; 10 : p. 1003.
- Baskaran K, Nirmaladevi N, Ilangovan M. Antioxidant properties *cladophora socialis* green algae of seaweeds collected from rameswaram in india, *International journal of current pharmaceutical research*, 2022; 14: 6.



24. Baskaran K, Pugalendi KV, Ramalingam S. Free radical scavenging activity of ethyl acetate extract of *Cardiospermum halicacabum* by *in vitro* assays, *Am. J. Biol. Pharm. Res*, 2014; 2: 300-8.
25. Malireddy S, Kotha SR, Secor JD, Gurney TO, Abbott JL, Maulik G, Maddipati KR, Parinandi NL. Phytochemical antioxidants modulate mammalian cellular epigenome: implications in health and disease, *Antioxidant Redox Signal*, 2012; 17(2): 327-339.
26. Doughari JH, Dr.Venketeshwer Rao (Ed.). *Phytochemicals: Extraction Methods, Basic Structures and Mode of action as Potential Chemotherapeutic Agents*, *Phytochemicals - A Global Perspective of their Role in Nutrition and Health*, 2012; 978-953.
27. Mamta S, Jyoti S, Rajeev N, Dharmendra S, Abhishek G. Phytochemistry of medicinal plants, *Journal of Pharmacognosy and Phytochemistry*, 2013; 1: 2278- 4136.
28. Kliebenstein DJ, Osbourn A. Making new molecules - evolution of pathways for novel metabolites in plants, *Curr. Opin. Plant. Biol*, 2012; 15: 415–23.
29. Patra B, Schluttenhofer C, Wu Y, Pattanaik S, Yuan L. Transcriptional regulation of secondary metabolite biosynthesis in plants, *Biochim. Biophys. Acta*, 2013; 1829: 1236–47.
30. Havsteen BH. The biochemistry and medical significance of the flavonoids, *Pharmacol. Ther*, 2002; 96: 67-202.
31. Dewick PM. *The Shikimate Pathway: Aromatic Amino Acids and Phenylpropanoids*, in *Medicinal Natural Products: A Biosynthetic Approach*, 3rd Edition. Chichester, UK: John Wiley & Sons, Ltd; 2009.
32. Shuaib OR, Adeniran OI, Musah M, Yerima H, Sani H, Amusat K. Comparative Nutritional and Anti-Nutritional Analysis of *Ocimum grattissimum* and *Ocimum basilicum*, *Academia Arena*, 2015; 7(7): 77-81.
33. Antolovich M. et al. Methods for testing antioxidant activity, *Analyst*, 2002; 127:183-198.
34. Re R. et al. Antioxidant activity applying an improved ABTS radical cation decolorization assay, *Free Radic. Biol. Med*, 1999; 26(9-10): 1231-1237.
35. Siddhuraju P, Becker K, The antioxidant and free radical scavenging activities of processed cowpea (*Vigna unguiculata*. L.) Walp.) seed extracts, *Food Chemistry*, 2007;101:10–19.
36. Halvorsen BL, Carlsen MH, Phillips KM, Bohn K, Holte K, Jacobs Jr, Blomhoff R, Content of redox-active compounds (ie, antioxidants) in foods consumed in the United States, *American Journal of Clinical Nutrition*, 2006; 84: 95–135.
37. Pellegrini N, Serafini M, Colombi B, Rio DD, Salvatore S, Bianchi M, Brighenti F, Total antioxidant capacity of plant foods, beverages and oils consumed in Italy assessed by three different *in vitro* assays, *Journal of Nutrition*, 2003; 133:2812–2819.
38. Hepsibha B.T, Sathiya S, Babu C.S, Premalakshmi V, Sekar T, *In vitro* studies of antioxidant and free radical scavenging activities of *Azima tetraacantha*. Lam leaf extract, *Indian Journal of Science Technology*, 2010; 3:571-577.
39. Rajkapoor B, Burkan Z.E, Senthilkumar R, Oxidants and human diseases: role of antioxidant medicinal plants-a review, *Pharmacologyonline*, 2010; 1: 1117-1131.
40. Cross AR, Jones OTG. Enzymatic mechanism of superoxide, *Biochimica. Biophysica. Acta*, 1991;38:281-285.
41. Gulcin, I., et al. Anti-oxidant and analgesic activities of turpentine of *Pinus nigra* Arn. subsp. pallsiana (Lamb) Holmboe, *Journal of Ethanopharmacology*, 1991; 86: 51-58.
42. Battu GR, Ethadi SR, Veda P.G, Evaluation of antioxidant and anti-inflammatory activity of *Euphorbia heyneana Spreng*, *Asian Pacific Journal of Tropical Biomedicine*, 2002;S191-S194.
43. Miller N.J, Rice-Evans C, Factors influencing the antioxidant activity determined by the ABTS⁺ radical cation assay, *Free Radical Research*, 1997;26(3):195-199.
44. Hippeli S and Elstner E.F, Transition metal ion catalyzed oxygen activation during pathogenic processes, *FEBS Letters*, 1999; 443: 1-7.
45. Dinis T.C.P, Madeira V.M.C and Almeida L.M, Action of phenolic derivatives (acetaminophen, salicylate, and 5-aminosalicylate) as inhibitors of membrane lipid peroxidation and as peroxyl radical scavengers, *Archives of Biochemistry and Biophysics*, 1994; 31: 161–169.

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