



Development and Characterization of Film Forming Spray of 5% Minoxidil for Baldness Due to Androgenetic Alopecia

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ABSTRACT

This study focuses on the treatment of Androgenetic Alopecia (AGA), a common type of hair loss that affects both men and women. The research problem addressed in this study is the need for effective and safe treatment options for AGA. The primary aim is to evaluate the safety and efficacy of minoxidil, a commonly used medication for AGA, and to explore novel topical delivery systems to enhance treatment outcomes and reduce side effects. The methodology employed in this study involves a comprehensive review of existing literature on AGA treatment, with a focus on minoxidil. Various studies and clinical trials exploring the safety and efficacy of minoxidil in the treatment of AGA are analyzed. Additionally, novel topical delivery systems for minoxidil, such as film-forming sprays, are discussed as potential solutions to enhance treatment outcomes. The findings of the study suggest that minoxidil is effective in promoting hair growth in patients with AGA, with variable levels of success reported in different studies. However, the use of traditional minoxidil formulations containing high ethyl alcohol and propylene glycol may lead to side effects such as dry scalp, irritation, and allergic reactions. Novel delivery systems like film-forming sprays offer a promising solution to reduce side effects and improve treatment efficacy. The implications of this study highlight the importance of exploring innovative delivery systems for existing medications to enhance their therapeutic effects and minimize adverse reactions. By developing novel formulations like film-forming sprays, healthcare providers can offer a more effective and patient-friendly treatment option for individuals with AGA.

Keywords: Androgenic alopecia, minoxidil, hair loss, treatment, topical delivery systems, film-forming sprays.

INTRODUCTION

Hair follicles control hair growth by using immunological cells, neuroproteins, and hormones. The intricate interplay among hair follicles found in various body parts results in the production of distinct hair types. The phases of hair growth are Anagen, which is the active growth stage; Catagen, which is the resting stage; Telogen, which is the hair follicle's regression phase; Exogen, which is the active phase of hair erosion; and Kenogen, which is the interval between the time when a hair follicle is empty and when new hair grows. The follicle has a bulbous bottom during the growing phase, with the dermal papilla located in the center. The papilla receives nourishment and oxygen from tiny blood vessels, which removes waste products and Hormones have a great effect on the papilla. This is the area where body's hormones and chemicals, either secreted or injected, affect the growth of hair, causing it to grow at a different rate or not at all. An increase in the amount of hair that falls out every day (effluvium) or noticeable hairlessness (alopecia) can both be referred to as "hair loss" in complaints. A normal day can see up to 100 hairs fall out. Hair loss is caused by several factors. It can be brought on by genetics, food, endocrine disorders, systemic diseases, drug use, and abnormalities in the hair shaft. The clinical examination should start with a thorough examination of the entire skin as well as the distribution of body and scalp hair. Particularly for androgenetic alopecia (AGA) and alopecia areata, grading systems have been developed. The Hamilton-Norwood scale is the most

widely used classification for male pattern baldness. The most well-known 3-point Ludwig scale for female pattern AGA is used with females. The Savin scale has eight differentiation classes and the Gan-Sinclair scale has five; both are more accurate at defining specifications and are now widely used. The scientific term for the genetic predisposition for pattern baldness or pattern hair loss in both men and women is androgenetic alopecia. Alopecia areata is an immune system disorder that results in the loss of hair growth from hair follicles. Hair loss that appears suddenly in small patches on the head is a typical sign. Alopecia universalis, which causes all body hair to disappear, and alopecia totalis, which results in the loss of all hair on the head, are advanced forms of the disorder. Androgenetic alopecia (AGA), a common type of alopecia, is a multifactorial, age-related chronic illness that has a major negative impact on the psychological well-being and quality of life of its patients. By the age of 70, it is estimated that between 80% and 50% of Caucasian men and women will have some form of AGA. The Food and Drug Administration (FDA) has approved finasteride, an oral medication, and minoxidil (2% and 5%) as topical formulations for the treatment of men with AGA. It is still unknown how precisely pilosebaceous transport mechanisms work.¹⁻¹³

Minoxidil is an effective oral medication for severe hypertension. As a hypertension treatment, it was first made available in the early 1970s. One common side-effect of minoxidil was hypertrichosis, which included hair growth in male baldness. When taken systemically for



hypertension, minoxidil functions as a vasodilator; however, when it comes to hair loss, its mode of action is through direct stimulation of the follicular hair matrix cell or dermal papillae. Although the exact mechanism by which minoxidil stimulates hair growth is still unknown, Hair follicle miniaturization is reversed by minoxidil's angiogenic effects, which also seem to lengthen the anagen phase. By doing this, the formation of hair is not inhibited by epidermal growth factor. Hair growth is thereby encouraged. The creation of novel topical delivery systems for minoxidil is necessary to reduce side effects and increase therapeutic efficacy. Minor side effects such as allergic contact dermatitis and scalp pruritus have been associated with topical minoxidil. Several earlier studies that looked into the safety and effectiveness of 5% minoxidil in the treatment of AGA discovered that 54-62% of patients had variable levels of hair growth.¹⁴⁻¹⁹

Pharmacokinetics of Minoxidil:
<https://www.ncbi.nlm.nih.gov/books/NBK482378/>

Absorption: The skin absorbs only 65% of topical minoxidil,

Distribution: There is no binding affinity of minoxidil to plasma proteins. With a distribution volume ranging from 2.8 to 3.3 L/kg, this medication has a wide distribution.

Metabolism: Topical minoxidil is converted to minoxidil sulfate in hair follicles through the action of the sulfotransferase enzyme, **Elimination:** Although minoxidil has a 3 to 4-hour elimination half-life, the drug's hypotensive effects can last up to 72 hours. Notably, the kidneys are primarily responsible for excreting minoxidil and its metabolites.

Structure:²¹

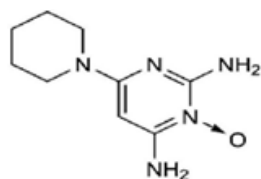


Figure 1: Minoxidil chemical structure

Advantages:²² Treatment of androgenetic alopecia, treatment of female pattern hair loss, alopecia areata, beard enhancement, eyebrow enhancement, chemotherapy-induced alopecia, doesn't induce hypotension, frontal fibrosing alopecia, etc.

Disadvantages:¹⁴ hair loss (telogen effluvium), scalp pruritus, scaling, irritant, allergic contact dermatitis, itching, erythema, etc.

FILM-FORMING SPRAY [FFS]

Numerous advancements have been made in recent decades to produce effective and efficient spray preparations. In general, FFS is made up of polymers, enhancers, and active ingredients dissolved in organic solvents. In contrast to other common topical preparations, a thin, non-sticky film can form that can prolong the drug's contact time and permeability, resulting in continuous drug release. It can also prevent

crystallization, increasing the amount of drug available to provide therapeutic effects.³⁹ A transdermal spray that forms a film would function more effectively, show more efficacy, cause fewer side effects, require fewer doses, and have a longer retention period. Through the formation of a uniform thin layer on the skin, the developed formulation will facilitate the rapid evaporation of solvents, resulting in a cooling effect and lowering the tendency to rub off. Because the formulation will stick to the skin longer and penetrate the skin more quickly, it will take effect quickly.⁴⁰ A non-solid dosage form that forms a film in situ or after being implemented to the pores and skin or another frame surface. These structures incorporate the drug and movie-forming excipients in a solvent that evaporates whilst coming in touch with the skin, leaving a movie of excipients and the drug behind the produced film can be a solid polymeric material that works as a matrix for drug release to the skin over time or a residual liquid film that is quickly absorbed in the stratum corneum.⁴¹ The film-forming transdermal spray keeps the same level of medication effectiveness but requires a much lower dosage, and it even eliminates more side effects.²² The sprayability of FFS is greatly influenced by the kind of nozzle, the size of the aperture, the pressure of the spray, and the type of liquid. Studying the viscoelastic, in situ gel, pH, and heat-sensitive characteristics of FFS is crucial to figuring out what factors to take into account when choosing polymers, solvents, and other excipients.³⁹

FILM-FORMING SPRAY OF 5% MINOXIDIL

A derivative of piperidine and pyrimidine, minoxidil has the following chemical structure: 2,6-diamino-4-piperidinopyrimidine-1-oxide (C₉H₁₅N₅O). The metabolite (minoxidil sulfate) is the primary factor contributing to minoxidil's beneficial effects on hair growth. While, Sulfotransferase, an enzyme found in hair follicles and with varying levels of production across individuals, is responsible for this conversion of minoxidil to minoxidil sulfate.²² Moreover, minoxidil is a 5-alpha reductase (5AR) inhibitor used in the pharmacologic therapy of AGA to reduce dihydrotestosterone (DHT) and stimulate hair follicles¹⁰. However, Hair thinning in an "M" shape characterizes androgenetic alopecia (AGA), also known as male-pattern baldness; the back and sides of the head are spared while the temples and crown of the head experience hair loss. Most people's distribution of androgen-sensitive follicles is reflected in this pattern. Androgens cause follicular miniaturization and shorten the anagen phase beginning at puberty, which results in the formation of vellus-like hair and progressive hair thinning.⁷ The 2% minoxidil solution was first launched in the market in 1986, followed by the 5% solution in 1993. Additionally, MDX exhibits side effects like redness, swelling, and itching, which makes treatment more difficult because a twice-daily application is advised for the recommended long-term course of care.

Furthermore, most commercially available (CA) formulations of MXD dissolve in propylene glycol (PG)

and/or ethanol due to the drug's poor water solubility; one such formulation is marketed using a mixture solvent of PG, water, and ethanol (20/30/50, v/v/v).³⁰ To address minoxidil solution side effects a new form of drug delivery system is developed called film-forming spray. FFS is made up of polymers, enhancers, and active ingredients dissolved in organic solvents. In contrast to other common topical preparations, a thin, non-sticky film can form that can prolong the drug's contact time and permeability, resulting in continuous drug release. It can also prevent crystallization, increasing the amount of drug available to provide therapeutic effects.³⁹ A transdermal spray that forms a film would function more effectively, show more efficacy, cause fewer side effects, require fewer doses, and have a longer retention period. Through the formation of a uniform thin layer on the skin, the developed formulation will facilitate the rapid evaporation of solvents, resulting in a cooling effect and lowering the tendency to rub off. Because the formulation will stick to the skin longer and penetrate the skin more quickly, it will take effect quickly.⁴⁰ Depending on the volume of solution sprayed, drug dosages in film-forming sprays can also be adjusted to regulate either local or systemic effects. Prescriptions are dispersed efficiently and uniformly thanks to an FFS. Additionally, ease of use can increase patient compliance. Removing the thin layer with water is an easy task. This thin, non-sticky film improves patient comfort during activities in contrast to patches, ointments, gels, and other moisture products that dry out harshly and become sticky when applied. In addition, the thin film facilitates wound/skin penetration, preserving equilibrium. Insufficient wound/skin humidity may cause irritation or infection, just like when patch preparations are used. Droplets are created using the film-forming solution. The film-forming fluid is sprayed with the usage of any sort of sprayer. Even though they are all intended for medical use, each sprayer has unique features and purposes. The different types of sprayers that could be used as drug delivery systems in film-forming systems.^[42] **BENEFITS:** Low Incidence of Irritation, Sterility of the dosage, excellent coverage of the skin, Adjustable Dosage, Increase the contact time, Increase the Permeability of the drug, Continuous drug release, Prevent crystallization, No Irritation, No Contamination Due to Repeat Exposure of Hands.

Application: Treatment of androgenetic alopecia, Treatment of Alopecia areata, Treatment of Chemotherapy-induced alopecia, Treatment of Hair transplant, Treatment of Scarring alopecia, Treatment of monilethri, Treatment of Hereditary alopecia or hypotrichosis.

MATERIALS AND METHODS

Formulation of Film- Forming Spray⁸¹

Method used for hydrophilic drug:

Take solvent in a beaker. Dissolve film forming agents completely (polymer) and then add the drug. Lastly add

Plasticizer and stabilizer of different grades according to formulation and dissolve completely. Transfer this FFS into a Spray bottle (container). Mixing time can vary according to the polymer and solvent used for film formations

Method Used for Hydrophobic Drugs:⁴⁰

First, 0.5% w/w plasticizer (PEG-400) and film formers [EC: Eudragit(:1:2 ratio)] were dissolved in the eutectic mixture, which contained equal amounts of menthol and camphor. A vehicle blend comprising 80 parts alcohol and 20 parts acetone was used to separately dissolve 0.5% w/w drugs. The mixture was then sonicated for 20 minutes in an ultrasonic cleaner. The final mixture was put into an assembly of containers. Mixing time can vary according to the polymer and solvent used for film formations.

Preparation of Formulation

first, Mix Ethanol and Acetone in a beaker (8:2), then, Dissolve Film-forming agents (HPMC E15) completely, and then add the drug, and mix it on a magnetic stirrer till it becomes homogeneous for 20 min. Lastly, add PEG 400 and glycerin to the formulation and stir it again for 15 minutes, then Transfer this FFS (film-forming solution) into a Spray bottle (container).



Figure 1: Film-forming formulation of different concentrations of ingredients



Figure 2: Film of minoxidil 5% film-forming spray

Table 1: Formulation composition

Ingredients	F1	F2	F3
Minoxidil	5	5	5
HPMC E15	1.2	1.8	2.4
PEG 400	0.25	0.25	0.25
Glycerin	0.5	0.5	0.5
Ethanol	60	60	60

EVALUATION PARAMETERS

pH: A digital pH meter that has been calibrated is used to measure the pH of the optimized formulation. The film-forming solution being tested is dipped into the pH meter's rod, and the meter's reading is recorded.⁴⁰

Viscosity: Using a Brookfield viscometer at room temperature, the viscosity of the prepared spray formulations was determined.⁴⁰ The film-forming solution's viscosity is a crucial parameter, particularly in MDS, as it impacts its spray ability. The spray's coverage area may decrease as the film-forming solution's concentration rises.³⁹

Film Formation: A Petri dish was used to administer film-forming systems. The film is labeled as either transparent or opaque, sticky or dry, and detachable or non-detachable based on its visual properties.⁸²

Drying Duration: To measure the drying time, the formulation is applied to the inner forearm sides of a volunteer or on a petri dish. After a set period, a glass slide is gently placed on the film. When there are no discernible liquid remnants on the glass slide following removal, the film is said to be dry. A minimum drying time should be a part of an efficient FFS to avoid making the patient wait a long time.⁸²

Water Washability: The dried film is used to evaluate the film's wetting ease. After being cleaned with water, the film is rated on an ordinal scale: easily, partially, and poorly cleaned.

Spray Angle⁶⁹: The pigmentation technique of a spray-on slip of paper was used to calculate the spray angle. The nozzle was positioned 15 cm away from the white paper, and the sprays were activated horizontally. To help with visual representation, 10 mg of Sudan red was dissolved in the formulation. Using the formula below, the spray angle was determined. $\text{spray angle } (\theta) = \tan^{-1}(l/r)$, Where, r = circle's mean radius, l = distance between the nozzle and sheet.

Volume Actuated Upon Each Spray⁶⁹: After each actuation, the weight difference was recorded and the volume delivered was estimated using the formula, $WT = WO - AL/Dn$, Where, AL = amount of solution delivered with each actuation, WT = formulation's load following an actuation, WO = formulation's initial weight before an actuation, Dn = density.

Drug Content⁶⁹: A ml of the prepared formulation was immersed for four hours at 100 rpm in phosphate buffer with a pH of 7.4 to assess the drug content. After filtering, the samples were suitably diluted. The mixture obtained was subjected to additional sonication, and spectrophotometric absorbance measurements were made at λ_{max} .

Stickiness: Using cotton wool, gently press the dried film. The amount of cotton wool fibers adhered to the film serves as a proxy for the viscosity of the material. If the attached fiber is thick, the film adhesiveness is deemed high; if the attached fiber is thin, it is deemed medium; and if there is little to no attached fiber, it is deemed low.⁸³

Flexibility: If the film is flexible and free of skin fixation disorders or cracks, it will be deemed flexible.⁶⁶

In-Vitro Drug Permeation Studies: The Franz diffusion cell's onion membrane was used for the *in-vitro* drug permeation process. The shape and size of the membrane used in the permeation studies were cut to match the dimensions of the diffusion cell. 7.4 pH phosphate buffer was added to the diffusion cell's receptor compartment. The donor and receptor compartments were separated by a 2.4 cm- diameter onion membrane. The film-forming solution was taken and added to the receptor compartment in an amount of 1 mL, which is equivalent to 5 mg of the drug. Subsequently, the stirrer was adjusted to 100 revolutions per minute. At precise intervals, 1 mL of sample were taken out of the receptor chamber. A U.V. spectrophotometer was used to measure the absorbance of these samples, which were prepared up to 10 mL in a volumetric flask and recorded at λ_{max} .⁴¹

RESULTS AND DISCUSSION

Viscosity, pH, Spray Angle, Volume Actuated Upon Each Spray, Evaporation Time, And Drug Content

The pH of the solutions was found to be between 5.5 and 6.5, which is within the normal range for the pH of the skin and scalp and is therefore it not irritating to the scalp. All formulations from F1 to F6, however, have viscosities between 26 and 48 cps. Additionally, the film-forming spray's uniform delivery on the scalp's surface is demonstrated by its spray angle, which lies between 36 and 40 degrees. The volume activated during each spray was within a narrow range and changed slightly in response to changes in the polymer's concentration. The concentration of polymers added to the formulation directly affects the drying time or timing of the film's formation; the higher the concentration, the longer the film takes to form while changing. Nevertheless, the drug concentration of the formulation was observed in spectrophotometry at λ_{max} of 279 nm. Drug content was also not affected significantly by changes in the concentration of polymers in the formulation.

Table 2: Result of Viscosity, pH, spray angle, volume actuated upon each spray, evaporation time, and drug content

Formulation number	pH	Viscosity cps	Spray angle	Volume actuated upon each spray (ml)	Evaporation time (min)	Drug content
F1	5.7	26.86	36.8±0.2	0.2436	5.5	98
F2	6.2	36.92	37.95±0.1	0.3665	6	97
F3	5.9	41.68	39.69±0.05	0.508	7.5	95

Film Washability, Stickiness, and Flexibility: The washability and stickiness of films were observed which is followed by rupturing and flexibility tests of film shown in table.

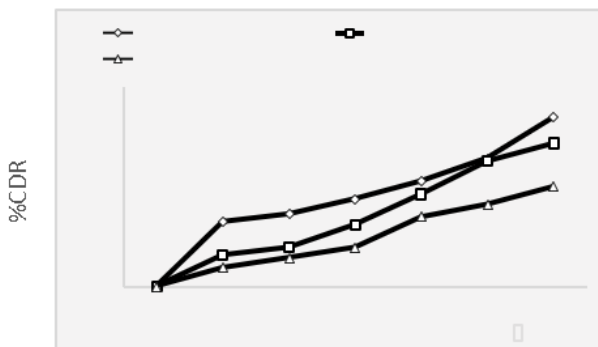
Table 3: Result of Film washability, stickiness, and flexibility.

Formulation number	Flexibility	Stickiness	Washability
F1	++	+++	+++
F2	++	+++	+++
F3	+	++	+++

Good (+++), moderate (++), poor(+).

In vitro permeation study of 5% minoxidil film-forming spray using a thin layer of onion.

Using an onion-thin layer, an in vitro drug permeation experiment was conducted for each formulation, in which F1 exhibited the highest drug content and cumulative drug release percentage. The percentage of cumulative permeation was then plotted against time. The above % cdr shows that the kinetic release of drug shows zero order kinetics.

**Figure 3:** Cumulative drug release of formulation F1, F2, F3

CONCLUSION

In this work, an attempt is made to create and describe a minoxidil film-forming spray by employing polymer at varying concentrations to create a consistent film after the solution is sprayed. The prepared formulation was examined for several evaluation factors, such as pH, drying time, and spray angle. Based on the above results, it was determined that formulation F2, which contains 1.8% hydroxy properly methyl cellulose E15, has good physical properties, good drug concentration, and cumulative drug release. Considering the outcomes of evaluation studies, it can be concluded that film-forming spray formulation can be a good alternative approach for treating patchy hair loss or baldness in men who have androgenetic alopecia.

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REFERENCES

- Salimi A and Soleymani S.M, "Transfollicular Drug Delivery Systems", *Jundishapur J Nat Pharm Prod* e82403, 2018. <https://doi.org/10.5812/ijpp.82403>.
- Srivastava A.K, Srivastava S.C, Srivastava N, "HAIR DISORDERS, TREATMENT AND CARE: AN OVERVIEW", *Journal of Recent Advances in Applied Sciences*, 2015;29(1):1-16.
- Wolff H, Tobias W. Fischer, Peytavi U.B, "The Diagnosis and Treatment of Hair and Scalp Diseases", *Dtsch Arztebl Int* 2016;113: 377–86. DOI: 10.3238/arztebl.2016.0377
- Brenner F.M, M.D. and Wilma F. Bergfeld, M.D., "hair Disorders", 2015.
- Hillmann H and Peytavi U.B, "Diagnosis of Hair Disorders", *Semin Cutan Med Surg*, 2009;28:33-38, doi:10.1016/j.sder.2008.12.005
- Ross EK, Vincenzi C, Tosti A, "Videodermoscopy in the evaluation of hair and scalp disorders", *J Am Acad Dermatol* 2006;55:799–806.
- Shapiro J and Price V.H, "Hair regrowth. Therapeutic agent.", *Dermatol Clin*, 1998;16:341-56.
- Drake L.A, Dinehart S.M, Farmer E.R, Goltz R.W, Graham G.F, Hordinsky M.K, " Guidelines of care for androgenetic alopecia", *American Academy of Dermatology*, 1996;35(3 pt 1):465-9.
- Yuan A.R, Bian Q, Gao J.Q., " Current advances in stem cell-based therapies for hair regeneration", *Eur J Pharmacol*; 2020,881:173197–208 .
- Hamza A.M, Walid M.M, "The Effect of topical Minoxidil in Treatment of androgenetic Alopecia", *MJMR*, 2020;31(4):350-351.
- Yueting Gu, Qiong Bian, "Hair follicle-targeting drug delivery strategies for the management of hair follicle-associated disorders", *Asian Journal of Pharmaceutical Sciences*, 2022;17:333–352, <https://doi.org/10.1016/i.ajps.2022.04.003>
- Kajimoto K, Yamamoto M, Kogure K, Kanamura K, "Noninvasive and Persistent Transfollicular Drug Delivery System Using a Combination of Liposomes and Iontophoresis", *International Journal of Pharmaceutics*, 2010;403(1-2):57-65.

- <https://doi.org/10.1016/j.ijpharm.2010.10.021>
13. Verma A, Jain A, Jain S.K, Hurkat P, "Transfollicular drug delivery: current perspectives", *Research and Reports in Transdermal Drug Delivery*, 2022;1-17. <https://doi.org/10.2147/RRTD.S75809>
 14. Rossi A, Cantisani C, Melis L, Lorio A, "Minoxidil Use in Dermatology, Side Effects and Recent Patents", *Recent Pat Inflamm Allergy Drug Discov.* 2012;6(2):130-6. <https://doi.org/10.2174/187221312800166859>
 15. Walsh D.S, Dunn C.L, James W.D, "Improvement in androgenetic alopecia (stage V) using topical minoxidil in a retinoid vehicle and oral finasteride", *Arch Dermatol*, 1995;131:1373-5.
 16. Shumi F.R, Sarker Mahbub Ahmed Shamim, "Adverse Effects Encountered during the Therapy of Topical Minoxidil combined with Systemic Finasteride and Topical Minoxidil Alone in Male Androgenetic Alopecia", *Annals of International Medical and Dental Research*, 2021;7(3):18-24.
 17. Uprit S, Sahu R.K, Roy A, "Preparation and characterization of minoxidil loaded nanostructured lipid carrier gel for effective treatment of alopecia", *Saudi Pharmaceutical Journal*, 2012;21:379-385. <http://dx.doi.org/10.1016/j.sjps.2012.11.005>
 18. Robert L. Rietschel, M.D, "Safety and efficacy of topical minoxidil in the management of androgenetic alopecia", *Journal of the American Academy of Dermatology* 1987;16(2):50-56.
 19. Sattur S, Talathi A, Sheety G, Arsiwala S, Dhoot D, "Comparative Clinical Study Evaluating the Efficacy and Safety of Topical 5% Cytosomal Minoxidil and Topical 5% Alcohol- Based Minoxidil Solutions for the Treatment of Androgenetic Alopecia in Indian Men", *Cureus* 2023;15(10):e46568. <http://dx.doi.org/10.7759/cureus.46568>
 20. Ervin Novakm, D. Thomasj.FRANZ, "Topically Applied Minoxidil in Baldness", *International journal of dermatology*, 1985;24:44-52.
 21. Parhi R, Terapalli B.R, Teja B.B, "Formulation and *In Vitro* Evaluation of Minoxidil Topical Gel", *Turk J Pharm Sci*, 2014;11(2):153-162.
 22. Suchonwanit P, Thammarucha S, Leerunyakul K, "Minoxidil and its use in hair disorders: a review", *Drug Design, Development and Therapy*: 2019;13:2777–2786, <http://doi.org/10.2147/DDDT.S214907>
 23. Purnak T, Senel E, Sahin C, "LIQUID FORMULATION OF MINOXIDIL VERSUS ITS FOAM FORMULATION", *Indian J Dermatol* 2011;56(4):462-8. http://dx.doi.org/10.4103/0019_5154.84714
 24. Clarence T. Ueda, "Topical and Transdermal Drug Products", *The United States Pharmacopeial Convention, Inc.* 2009.
 25. Barbareschi M, Vescovi V, Starace M, Milani M, "Propylene glycol free 5% minoxidil lotion formulation: cosmetic acceptability, local tolerability, clinical efficacy and *in vitro* skin absorption evaluations", *Giornale Italiano di Dermatologia e Venereologia* 2020;3(2):20-25. <http://dx.doi.org/10.23736/S0392-0488.20.06554-2>
 26. Kumar P, Singh S.K, Jindal D.K, Handa V, "Formulation and Evaluation of Minoxidil Gel Using Acrylamide/Sodium Acryloyldimethyl taurate copolymer for Alopecia areata", *International Journal of Pharmaceutical Sciences and Drug Research*; 2018;10(1):01-06, <http://dx.doi.org/10.25004/IJPSDR.2018.100101>
 27. Hasanzadeh H, Halavati N, Saberi M, "Efficacy and safety of 5% minoxidil topical foam in male pattern hair loss treatment and patient satisfaction", *Acta Dermatovenerol APA*; 2016;25:41-44, <http://dx.doi.org/10.15570/actaapa.2016.12>
 28. Cheng Zhou, Weixin Fan, Yang Q, Chen A, "Comparison of a New 5% Minoxidil Foam and Rogaine in the Treatment of Androgenetic Alopecia in Chinese Men: A Randomized, Double-Blind, Phase III, Equivalence Trial", *Hindawi Dermatologic therapy*, 2023. <https://doi.org/10.1155/2023/4582911>
 29. Santos A.C, Pereira I, Pita I, Veiga F, Guerra C, Costa D, "Topical Minoxidil-Loaded Nanotechnology Strategies for Alopecia", *Cosmetics*, 2020;7:21-7; <https://doi.org/10.3390/cosmetics7020021>
 30. Nagai N, Yoshie Iwai, "Drug Delivery System Based On Minoxidil Nanoparticles Promotes Hair Growth In C57BL/6 Mice", *International Journal of Nanomedicine*: 2019;14:7921–7931, <http://doi.org/10.2147/IJN.S225496>
 31. Padois K, Pirot F, Falson F, Bertholle V, "Solid lipid nanoparticles suspension versus commercial solutions for dermal delivery of minoxidil", *Int J Pharm*; 2011;416:300–304. <https://doi.org/10.1016/j.ijpharm.2011.06.014>
 32. Jung S, Otberg N, Thiede G, Richter H, Sterry W, Panzner S, "Innovative liposomes as a transfollicular drug delivery system: penetration into porcine hair follicles", *J Invest Dermatol*; 2006;126(8):1728–32. <https://doi.org/10.1038/sj.jid.5700323>.
 33. Sascha Jung, Otberg N, Thiede G, Panzner S, "Innovative Liposomes as a Transfollicular Drug Delivery System: Penetration into Porcine Hair Follicles", *Journal of Investigative Dermatology*; 2006;126:1728–1732. <https://doi.org/10.1038/sj.jid.5700323>
 34. Jain B, Singh B, Katara O.P, Vyas S.P, "Development and characterization of minoxidil- loaded liposomal system for delivery to pilosebaceous units", *Journal of Liposome Research*, 2010;20(2):105–114, <https://doi.org/10.3109/08982100903161449>
 35. Usmania, Bilandi A, Kataria M.K, "Formulation and evaluation of minoxidil emulgel for *Androgenic alopecia*", *Indo American Journal of Pharmaceutical Sciences*; 2016;3(12):2349-7750. <http://doi.org/10.5281/zenodo.258181>
 36. Donthi M.R, Singhvi G, Saha R.N, Munnangi S.R, "Nanoemulgel: A Novel Nano Carrier as a Tool for Topical Drug Delivery", *Pharmaceutics*, 2023;15:164-9. <https://doi.org/10.3390/pharmaceutics15010164>
 37. Eman Abd, Grice J.E, Roberts M.S, Heather A.E Benson, "Minoxidil Skin Delivery from Nanoemulsion Formulations Containing Eucalyptol or Oleic Acid: Enhanced Diffusivity and Follicular Targeting", *Pharmaceutics*, 2018;10:19-24; <https://doi.org/10.3390/pharmaceutics10010019>
 38. Umar A.K, Butarbutar M, Wathoni N, Sriwidodo S, "Film-Forming Sprays for Topical Drug Delivery", *Drug Design, Development and Therapy*: 2020;14:2909–2925. <http://doi.org/10.2147/DDDT.S256666>
 39. Desai M, Godbole A.M, "Design, Development, and Characterization of Film Forming Spray as Novel Antifungal Topical Formulation for Superficial Fungal Infections", *Indian Journal of Pharmaceutical Education and Research*, 2022;56(4):60-66. <https://doi.org/10.5530/ijper.56.4s.211>
 40. Sai Soujith N.B, Jawahar N, "Terbinafine HCl Film-Forming Spray for the Treatment of Topical Fungal Infections", *Ind. J. Pharm. Edu. Res*; 2023,57(1s):s85-s97, <https://doi.org/10.5530/ijper.57.1s.10>
 41. Tilekar S.K, Sayyad S.F, Phopse B.K, "A COMPREHENSIVE REVIEW ON FILM FORMING TOPICAL SPRAY DRUG DELIVERY," *World Journal of Pharmacy and Pharmaceutical Sciences*, 2022;11(10):496-02. <https://doi.org/10.20959/wjpps202210-23311>
 42. Haghghat A.N, Kosar. M, Rastegari A, Montazeri H, Faghihi H, Aghsami M, "Chitosan and its amphiphilic derivative nanoparticles loaded with Minoxidil for induction of hair growth: *In vitro* and *in vivo* evaluation", *International Journal of Biological Macromolecules*, 2024;259(1):12912. <https://doi.org/10.1016/j.ijbiomac.2023.129122>.
 43. Jiayu Mi, Zheng K, Jiang L, Sun Z, Pang, "Minoxidil/salicylic acid hydrogel formulated for alopecia treatment: Supramolecular interactions modulate physicochemical properties and biological", *Journal of Molecular Structure*, 2024;1306:137847, <https://doi.org/10.1016/j.molstruc.2024.137847>
 44. Lismare da Silva Prado, Grivicich I, Silva J.D, "Toxicological assessment of minoxidil: A drug with therapeutic potential besides alopecia", *Food and Chemical Toxicology*, 2023;182:114211 <https://doi.org/10.1016/j.fct.2023.114211>
 45. Francielle de Fátima Viana Santana, Lozi A.A, Silva J.D, Goncalves R.V,



- “Comparative effects of finasteride and minoxidil on the male reproductive organs: A systematic review of *in vitro* and *in vivo* evidence”, *Toxicology and Applied Pharmacology*, 2023;478:11671 <https://doi.org/10.1016/j.taap.2023.116710>
46. Arora G, Mishra A, Chandra N, Jain G.K, “In Vitro and In Vivo Scalp Retention and Penetration of 99mTc-Minoxidil Solution”, *Journal of Pharmaceutical Sciences*, 2023; 112(1):230-23. <https://doi.org/10.1016/j.xphs.2022.09.016>
 47. Makhlof A, Elnawawy T, “Hair regrowth boosting via minoxidil cubosomes: Formulation development, in vivo hair regrowth evaluation, histopathological examination and confocal laser microscopy imaging”, *International Journal of Pharmaceutics*, 2023;634:5122665. <https://doi.org/10.1016/j.ijpharm.2023.122665>
 48. Ying Sun, Zhang Y, Liu X, Kong X, Shen L, “The preparation of high minoxidil loaded transfersomes and its gel for effective topical treatment of alopecia”, *Journal of Drug Delivery Science and Technology*, 2023;84:104458. <https://doi.org/10.1016/j.jddst.2023.104458>
 49. Teerasong S, Praditweangkum W, Chompoosor A, “A new mechanism for resonance Rayleigh scattering detection of minoxidil based on catalytic oxidation of silver nanoparticles”, 2022;275(5):121147. <https://doi.org/10.1016/j.saa.2022.121147>
 50. Allam A.A, Fathalla D, Soliman G.M, Safwat M.A, “Transferosomes versus transthesomes for the dermal delivery for minoxidil: Preparation and *in vitro/ex vivo* appraisal,” *Journal of Drug Delivery Science and Technology*, 2022;76:103790. <https://doi.org/10.1016/j.jddst.2022.103790>
 51. Oliveira P.M, Silva T.A, Carvalho J.L, Gratieri T, “Nanostructured lipid carriers loaded with an association of minoxidil and latanoprost for targeted topical therapy of alopecia,” *European Journal of Pharmaceutics and Biopharmaceutics* volume, 2022;172:78-88. <https://doi.org/10.1016/j.ejpb.2022.02.003>
 52. Basuri P.P, kumar V, Nalini C.N, “Estimation of minoxidil in human plasma using UHPLC-MS/MS and its application in pharmacokinetic study”, *Journal of Chromatography B*, 2022;1192:123104. <https://doi.org/10.1016/j.ichromb.2022.123104>
 53. Seongryeong Han, Jang H.S, Shim J.H, Kim M, Lee Y “Development of minoxidil-loaded double emulsion PLGA nanoparticles for the treatment of hair loss”, *Journal of Industrial and Engineering Chemistry*, 2022;113(25):161-169. <https://doi.org/10.1016/j.jiec.2022.05.040>
 54. Kim M.J, Seong K.Y, Kim D.S, Jeong J.S “Minoxidil-loaded hyaluronic acid dissolving microneedles to alleviate hair loss in an alopecia animal model”, *Acta Biomaterialia*, 2022;143:189-202. <https://doi.org/10.1016/j.actbio.2022.02.011>
 55. Ray R, Sharma A, “Comparison of 5% minoxidil lotion monotherapy versus its combination with autologous platelet rich plasma in androgenetic alopecia in hundred males”, *Medical Journal Armed Forces India*, 2021;77(3):355-362. <https://doi.org/10.1016/j.mjafi.2020.11.010>
 56. Ngampanya A, Patel N.K, Sermsappasuk P, “Development and Qualification of a Physiologically Based Pharmacokinetic Model of Finasteride and Minoxidil Following Scalp Application”, *Journal of Pharmaceutical Sciences*, 2021;110(5):2301-231. <https://doi.org/10.1016/j.xphs.2021.02.016>
 57. Tara Barat MD, “Evaluation of the efficacy and safety of cow placenta extract lotion versus minoxidil 2% in the treatment of female pattern androgenetic alopecia”, *International Journal of Women's Dermatology*, 2020;6(4):318-321.
 58. Cardoso S.A, Thaís Nogueira Barradas, “Developing formulations for drug follicular targeting: Nanoemulsions loaded with minoxidil and clove oil”, *Journal of Drug Delivery Science and Technology*, 2020;59:101908. <https://doi.org/10.1016/j.jddst.2020.101908>
 59. Ramezani V, Honarvar M, Ranjbar A.M, Hashemi M, Karimollah A, “Formulation and optimization of transfersome containing minoxidil and caffeine”, *Journal of Drug Delivery Science and Technology*, 2018;44:129-13. <https://doi.org/10.1016/j.jddst.2017.12.003>
 60. Maitra M, Goyal A.K, Rath G, “A novel approach for follicular delivery of minoxidil for treatment of alopecia”, *Journal of Drug Delivery Science and Technology*, 2017;41:113-123. <https://doi.org/10.1016/j.jddst.2017.07.002>
 61. Maíra N. Pereira, Lima E.M, Gratieri T, Schulte H.L, “Solid effervescent formulations as new approach for topical minoxidil delivery”, *European Journal of Pharmaceutical Sciences*, 2017;96:411-419. <https://doi.org/10.1016/j.ejps.2016.10.016>
 62. Pawar N, Parmar A, Mishra D.N, Bahmani K, Malik R, Pawar R, Jalwal P, “Formulation, Optimization and Evaluation of Non-aerosol Topical Spray of Lidocaine for Pain Management”, *Int. J. Pharm. Investigation*; 2021;11(4):414-419. <https://doi.org/10.5530/ijpi.2021.4.74>
 63. Mohite P, “Film Forming Spray: A Comprehensive Review”, *International Journal of Innovative Science and Research Technology*, 2022;7(12):2456-2165.
 64. Ip40 Pawar N, Jalwal P, “Non-Pressurized Topical Spray Pharmaceutical- Methodology of Formulation Development and Quality Control Management”, *Int. J. Pharm. Investigation*; 2021;11(3):260-268, <https://doi.org/10.5530/ijpi.2021.3.46>
 65. Ip44 Elena O. Bakhrušina, “Spray Film-Forming systems as promising topical in situ Systems: A review”, *Saudi Pharmaceutical Journal*; 2023;31:154-169.
 66. Ip43 Rathi A, Pethe A, “COMPARATIVE EVALUATION OF FILM FORMING PROPERTIES OF SOME NATURAL POLYMERS”, *Int J Pharm*; 2014;4(1):347-356.
 67. Cesar M, Terence, Faldini, Braunstein S, Figueiredo L, Junqueira P, “Preparation and Characterization of a Polymeric Blend of PVP/PVAL for Use in Drug Delivery System”, *Journal of Biomedical Nanotechnology*, 2011;7:1-4, <https://doi.org/10.1166/jbn.2011.1303>
 68. Cesar M, Terence, Faldini, Braunstein S, Figueiredo L, Junqueira P, de Miranda, “Preparation and Characterization of a Polymeric Blend of PVP/PVAL for Use in Drug Delivery System”, *J. Biomed. Nanotechnol.* 2011;7(3):30-36. <https://doi.org/10.1166/jbn.2011.1303>
 69. Paola Franco and Iolanda De Marco, “The Use of Poly(N-vinyl pyrrolidone) in the Delivery of Drugs: A Review”, *Polymers* 2020;12:1114-21; <https://doi.org/10.3390/polym12051114>
 70. Horvat G, Zvab K, Knez Z and Novak Z, “Simple, One-Pot Method for Preparing Transparent Ethyl Cellulose Films with Good Mechanical Properties”, *Polymers* 2022;14:2399. <https://doi.org/10.3390/polym14122399>
 71. Archana D, Singh B.K, Dutta J, Dutta P.K, “Chitosan-PVP-nano silver oxide wound dressing: in vitro and in vivo evaluation”, *Int J BiolMacromol.* 2020;73:49-57.
 72. Zhuang C, Zhong Y, Zhao Y, “Effect of deacetylation degree on properties of Chitosan films using electrostatic spraying technique”, *Food Control.* 2019;97:231-8.
 73. Zhong Y, Zhuang C, GuW, Zhao Y. “Effect of molecular weight on the properties of chitosan films prepared using electrostatic spraying technique”. *CarbohydrPolym.* 2019;212:197–205.
 74. Sonje A, “Int.Res.J.Pharm,2013;4(5):55-62. <https://doi.org/10.7897/2230-8407.04515>
 75. Leichtnam M-L, Rolland H, Wüthrich P, Guy R.H, “Impact of anti-nucleants on transdermal delivery of testosterone from a spray”, *J Pharm Sci*, 2007;96(1):84–92. <https://doi.org/10.1002/jps.20670>
 76. Roy N, Agrawal M', “Review Article on Permeation Enhancers: A Major Break Through In Drug Delivery Technology”, *International Journal of Pharmaceutical Sciences and Research*, 2017;8(3):1001-1011, [http://dx.doi.org/10.13040/IJPSR.09758232.8\(3\).1001-11](http://dx.doi.org/10.13040/IJPSR.09758232.8(3).1001-11)



77. Shahadha R.W and Maraie N.K, "Mucoadhesive Film Forming Spray for Buccal Drug Delivery: A Review", *Al Mustansiriyah Journal of Pharmaceutical Sciences*, 2023;23(1):70-77.
78. Miroslava Spaglova, "Microemulsions as Solubilizers and Penetration Enhancers for Minoxidil Release from Gels", *Gels*, 2021;7:26-33. <https://doi.org/10.3390/gels7010026>
79. Larissa Carine Pünnel and Dominique Jasmin Lunter, "Film-Forming Systems for Dermal Drug Delivery", *Pharmaceutics*, 2021;13:932-8. <https://doi.org/10.3390/pharmaceutics13070932>
80. Parab A and Mehta D, "Design of Film Forming Spray solution for Antimicrobial Agents", *Eur. Chem. Bull*, 2023;12(07):3752 –3771.
81. Zhang, J. and Michniak-Kohn, B., "Investigation of microemulsion microstructures and their relationship to transdermal permeation of model drugs: ketoprofen, lidocaine, and caffeine", *International journal of pharmaceutics*, 2019;421(1):34-44.
82. Mhatre S, Kolhe N, "A REVIEW ON FILM-FORMING SPRAY SOLUTIONS", *IJCRT*, 2023;11(11):2320-2882.
83. Teixeira AZA, Saini G and Macgregor, "A. Factorial design used in optimization immediate release solid dosage ranitidine hydrochloric", *Estud Biol*. 2006;28(62):17-25. <https://doi.org/10.7213/rev.v28i62.22714.v28i62.22714>
83. Kathe K, Kathpalia H, "Film forming systems for topical and transdermal drug delivery", *Asian Journal of Pharmaceutical Sciences*, 2017;12:487-497.

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