



Sodium Selenite Pretreatment Ameliorates Aspects of the Nephropathy Induced by Mercuric Chloride in the Ratte *Albinos Wistar*

Youcef Necib^{a,*}, Ahlem Bahi^a, Sakina Zerizer^b, Cherif Abdennour^c, Mohamed Salah Boulakoud^c

^aDepartment of Biochemistry and biological cellular and molecular, Faculty of sciences, Mentouri university, BP 25000 Constantine, Algeria.

^bLaboratory of microbiological engineering and application, Faculty of sciences, Mentouri university, BP 25000 Constantine, Algeria.

^cAnimal Ecophysiology laboratory, Faculty of Sciences, Badji Mokhtar University, BP12 Sidi Amar, Annaba, Algeria.

*Corresponding author's E-mail: youcefnechib@yahoo.fr

Accepted on: 21-08-2013; Finalized on: 31-10-2013.

ABSTRACT

The study was designed to investigate the possible protective role of sodium selenite in mercuric chloride induced renal stress, by using biochemical approaches. Female rats *Albinos Wistar* were randomly divided into four groups. The first group was served as the control, the second group was given sodium selenite (1 mg/kg b.w), while the third group was given mercuric chloride (1 mg/kg), finally, the fourth group was given combined treatment of sodium selenite and mercuric chloride for 10 days. The effects of sodium selenite on mercuric chloride induced oxidative and renal stress were evaluated by serum creatinine, urea and uric acid, kidney tissue lipid peroxidation, GSH levels, GSH-Px and GST activities. Administration of mercuric chloride induced significant increase in serum: creatinine, urea and uric acid concentration showing renal stress. Mercuric chloride also induced oxidative stress, as indicated by decreased kidney tissue of GSH level, GSH-Px and GST activities along with increase the level of lipid peroxidation. Furthermore, treatment with mercuric chloride caused a marked elevation of kidney weight and decreased body weight. Sodium selenite treatment markedly reduced elevated serum: creatinine, urea and uric acid levels, and counteracted the deleterious effects of mercuric chloride on oxidative stress markers and attenuated histopathological changes caused by HgCl₂ in kidney. Our results indicate that sodium selenite could have a beneficial role against mercuric chloride induced nephrotoxicity and oxidative stress in rat.

Keywords: Mercury, sodium selenite, lipid peroxidation, reduced glutathione, antioxidant enzymes.

INTRODUCTION

Mercury is a well-known human and animal induces extensive kidney damage nephrotoxicant. acute oral or parenteral exposure induces extensive kidney damage¹. Studies *in vivo* and *in vitro* have demonstrated that mercury induced lipid peroxidation, suggesting the involvement of oxidative stress in its cytotoxicity. Lund et al. (1993)² reported that mercury enhances renal mitochondrial hydrogen peroxide formation *in vivo* and *in vitro*. However, causative correlation between mercury induced lipid peroxidation and cellular toxicity remains controversial. Some authors reported that lipid peroxidation plays a critical role in cell injury induced by mercury² in renal cells, whereas other investigators showed that lipid peroxidation is not directly responsible for mercury induced cell injury in hepatocytes and renal cells³. It is believed that antioxidants should be one of the important components of effective treatment for mercury poisoning. Indeed, HgCl₂ induced injury can be ameliorated by superoxide dismutase⁴ and non-enzymatic antioxidants like vitamin C, Vitamin E, cysteine and selenium⁶ have proven helpful to overcome oxidative damage.

The biological importance of selenium is at least 3-fold. First, it forms the prosthetic group of some critical selenocysteine containing enzymes, such as glutathione peroxidase, iodothyronine 5'-deiodinase, and thioredoxin reductase⁷. Second, sodium selenite is protective against

a number of toxicants. Third, selenium excessive intake cause toxic potential⁸. The purpose of this study was to evaluate the protective role of selenium on mercury chloride induced oxidative and renal stress in rats.

MATERIALS AND METHODS

All chemicals used in this work were purchased from sigma chemical company. Laboratory animals, *Albino Wistar* male rats, were brought from the Algiers Pasteur institute at the age of 8 weeks, with an average live weight of 200g. They were located in a room with an ambient temperature of 21±1°C and up to 12h of light daily. The rats were divided into four experimental groups; each consists of six rats. The first group was served as the control. The second group was given sodium selenite at a dose of 1 mg/kg body weight, while the third group (HgCl₂) was intraperitoneally given mercuric chloride at a dose of 1 mg/kg body weight. Finally, the fourth group was given combined treatment with sodium selenite and mercuric chloride. The treatment of all groups was lasted for 10 days.

Twenty four hours after the last administration, blood was collected by retro-orbital sinus puncture. After centrifugation at 3000 rpm for 10min, the serum was separated immediately and stored at -20°C until determination of urea, creatinine, and uric acid levels. Subsequently, rats were decapitated and kidneys were removed.



Tissue preparation

About 500mg of kidney was homogenized in 4ml of buffer solution of phosphate buffered saline (w/v: 500mg tissue with 4ml PBS, PH 7.4) homogenates were centrifuged at 10.000xg for 15min at 4°C. And the resultant supernatant was used for determination of: reduced glutathione (GSH) according to the method of Weeckbekeretory (1988)⁹, Thiobarbituric acid- reactive substance (TBARS) level by method of Buege and Aust (1978)¹⁰, and glutathione peroxidase (GSH-PX) and glutathione –S-transferase (GST) activities were measured by the method Flohe and Gunzler(1984)¹¹ and Habig et al (1974)¹² respectively. However, protein content was measured by the method of Bradford (1976)¹³.

Histopathological examination

Kidney from autopsied animals were excised out and fixed in formalin (10%). five micron thick section were prepared by using microtome and these section were stained with hematoxyline and eosin. For histological alterations these slides were observed under light microscope.

Determination of Biochemical parameters

Serum urea, creatinine and uric acid levels were determined using commercial kits (Spinreact).

Statistical analysis

The data were subjected to student *t* test for comparison between groups. The values are expressed as mean ±

SEM. Significance level was set at $P < 0.05$, $P < 0.01$, $P < 0.001$.

RESULTS

Effects of treatments on body, absolute and relative kidney weights

Table 1 shows the effect of mercuric chloride, sodium selenite and combined treatment with mercuric chloride and sodium selenite. The marked decreased in rats body weight was observed in mercuric chloride treated rats and mercuric chloride + sodium selenite group, but the result was not significant as compared to control. Along sodium selenite showed increased body weight but result was not significant. The kidneys of rats treated with mercuric chloride were enlarged. Mercuric chloride treated rats showed a highly significant increased kidney weight and relative kidney weight ($P \leq 0.001$) as compared to control. Combined treatment with sodium selenite showed significant increased relative kidney weight, while alone sodium selenite treatment had showed no significant effect.

Effects of treatment on serum biochemical parameters

A highly significant ($P \leq 0.001$) elevation in serum urea, creatinine and uric acid levels was observed in mercuric chloride intoxicated rats. Only sodium selenite treatment did not show any significant alteration. However, the combined treatment of sodium selenite with mercuric chloride show a highly significant decline in serum urea, creatinine and uric acid was noticed respect to mercuric chloride treated animals (table 2).

Table 1: Changes in body and absolute and relative kidney weights of control and rats treated with selenium (Se), mercuric chloride, and combined treatment of mercuric chloride with selenium after 10 days of treatment.

Parameters	Treatment groups			
	Control	Se	HgCl ₂	Se + HgCl ₂
Initial body weight (g)	226.66±26.56	222.5±12.56	221±35.36	224.66±25.5
Final body weight (g)	227±18.4	225.83±18.9	193.16±31.0	218.81±15.05
Absolute kidney weight (g)	1.04±0.24	1.11±0.53	2.23±0.82**	1.76±0.18**
Relative kidney weight (g/100g b.w)	0.45±0.008	0.49±0.006	1.15±0.02**	0.8±0.01***

Values are given as mean ± SEM for group of 6 animals each. * $P \leq 0.05$, compared to controls. ** $P \leq 0.01$, compared to controls. *** $P \leq 0.001$, compared to controls.

Table 2: Changes in biochemical parameters of control and rats treated with selenium (Se), mercuric chloride, and combined treatment of mercuric chloride with selenium after 10 days of treatment.

Parameters	Treatment groups			
	Control	Se	HgCl ₂	Se + HgCl ₂
Urea (g/l)	0.25±0.02	0.23±0.01	0.37±0.03	0.33±0.05**
Creatinine (mg/l)	2.89±0.71	2.92±0.55	3.94±0.98	3.17±0.54
Uric acid (mg/l)	15.55±4.50	16.55±2.43	24.81±6.87*	23.04±4.35**

Values are given as mean ± SEM for group of 6 animals each. * $P \leq 0.05$, compared to controls. ** $P \leq 0.01$, compared to controls. *** $P \leq 0.001$, compared to controls.

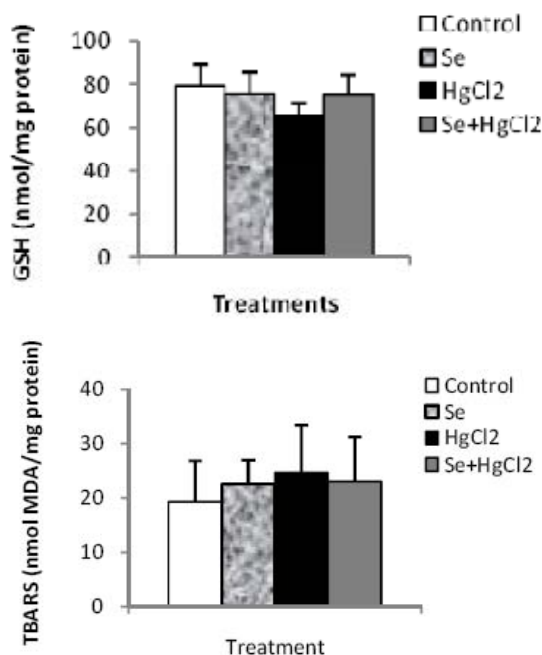


Figure 1: Reduced glutathione (nmol/ mg protein) and TBARS (nmol MDA/mg protein) levels in kidney of control and rats treated with selenium, mercuric chloride, and combined treatment of mercuric chloride with selenium after 10 days of treatment. Values are given as mean ± SEM for group of 6 animals each significant difference: * compared to controls (*P≤0.05; **P≤0.01; ***P≤0.001).

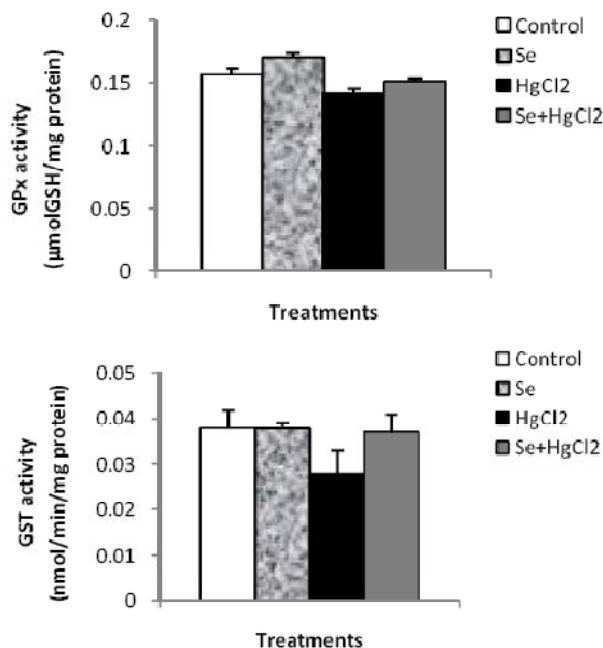


Figure 2: Enzyme activities of GPx (µmol GSH/ mg protein) and GST (nmol/min/mg protein) in kidney of control and rats treated with selenium, mercuric chloride, and combined treatment of mercuric chloride with selenium after 10 days of treatment. Values are given as mean ± SEM for group of 6 animals each significant difference: * compared to controls (*P≤0.05; **P≤0.01; ***P≤0.001).

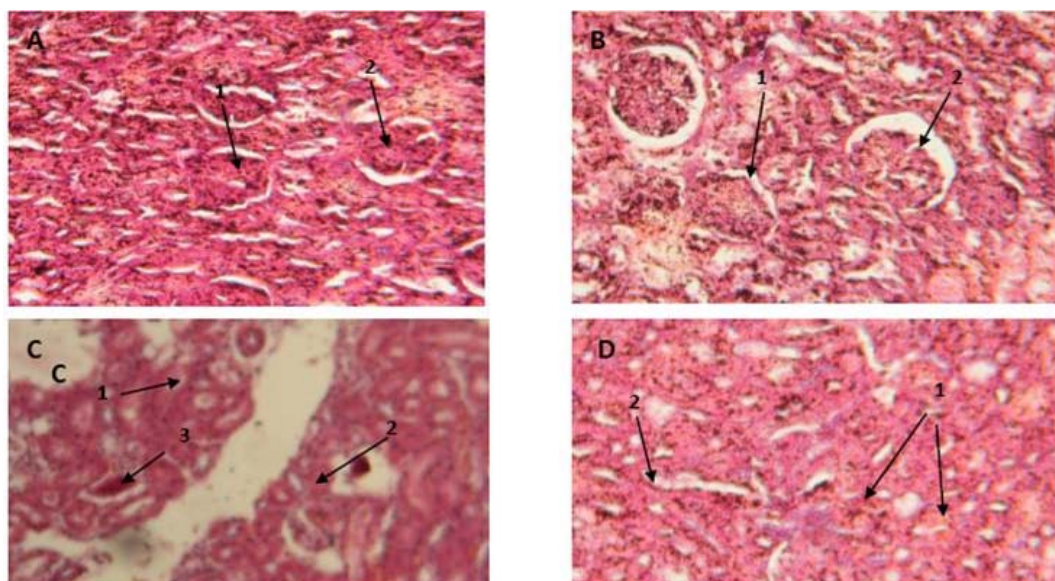


Figure 3: T.S. of kidney of Female rat treated with mercuric chloride (Hg) alone, and in combination with sodium selenite. (A) control (H&E100X): showing well develop glomerulus (1), with normal tubular cells; (B) sodium selenite alone treatment (H&E 100X): showing normal glomerulus (1), and normal tubular cells; (C) mercury treatment (H&E100X): showing degeneration of tubular cells (1), loss of nucleus (2), degeneration of glomerulus (3); (D) combined treatment of mercuric chloride with sodium selenite (H&E100X): showing normal glomerulus (1), normal tubular cells (2).

Effects of treatments on renal oxidative stress parameters

Mercuric chloride exposure a significant depleted in reduced glutathione level, GSH-Px and GST activities. And a significant increase in kidney lipid peroxidation level in mercury intoxicated rats was noticed. Sodium selenite

alone treatment did not show any significant decline. In combined treatment of mercuric chloride with sodium selenite highly significant increase in reduced glutathione level, GSH-Px and GST activities. And significant depletion in lipid peroxidation level was recorded with respect to mercury intoxicated rats (Fig.1 and 2).

Histological studies

The histological changes in Kidney are presented in Fig.3. Mercuric chloride induced various pathological alterations in kidney of rats. These alterations were characterized by renal tubular damage, indicating by tubular necrosis (Fig. 3C). In combination group were sodium selenite was administration with mercuric chloride showed reparative changes. Kidney showed prominent recovery in the form of normal renal tubular and very less tubular necrosis (Fig. 3D). kidney of the control group had a regular histological structure (Fig.3A). Furthermore, no histological alterations were observed in the kidney of sodium selenite treated group (Fig. 3B).

DISCUSSION

In the present study, oxidative stress induced by HgCl₂ was evidenced in kidney of rats by increase in lipid peroxidation level and the stimulation of GSH-Px, and GST activities. Accordingly, oxidative stress induced by HgCl₂ has been previously reported². As a consequence of lipid peroxidation biological membranes are affected causing cellular damage. Renal damage observed in rats exposed to HgCl₂ was also evidenced by increase in the plasmatic levels of urea, creatinine and uric acid, which are renal markers of damage. In the present study, serum urea, creatinine and uric acid levels were significantly increased after 10 days mercuric chloride (1mg/kg), showing insufficiency of renal function. Studies in animals have established that tubular injury plays a central role in the reduction of glomerular filtration rate in acute tubular necrosis. Two major tubular abnormalities could be involved in the decrease in glomerular function in mercuric chloride treated rats: obstruction and backleak of glomerular filtrate⁵. The alterations in glomerular function in mercuric chloride treated rats may also be secondary to ROS (reactive oxygen species), which induce mesangial cells contraction, altering the filtration surface area and modifying the ultrafiltration coefficient factors that decrease the glomerular filtration rate¹⁴. The activity of GSH-Px and GST that can clear to protect the cells from being injured represents the competence of clearing free radicals from the organism. MDA content manifests the level of lipid peroxidation, and then indirectly represents the level of damage of the cell of renal mitochondria. Evaluating from GSH, MDA levels and GSH-Px and GST catalase activities in kidney of rats. Hg alone significantly decreased GSH level, GSH-Px and GST activities and increased MDA content along with histological damage in kidney.

Co-administration of Hg and Se significantly increased GSH level, and activities of GSH-Px and GST, catalase and decreased MDA content. The effect of Hg and Se interaction depended on the molar ratio of these elements administrated to animals. The maximal effect of Se on Hg induced nephrotoxicity was observed when Se was given the same mol as Hg. Hg can give rise to free radicals that induce lipid, protein, and DNA oxidation. Hg has a great affinity for SH groups of proteins and enzymes

that are crucial in cell metabolism. Endogenous antioxidant enzymes such as GSH-Px and GST are involved in the protection against oxidative stress and lipid peroxidation in kidney¹⁵. Induction of these antioxidant enzymes indicates an adaptive onset of the redox defence system, whereas inhibition is thought to contribute to oxidative stress in mouse brain following mercury intoxication^{16,17}. Se can enhance antioxidant ability by enhancing activities of antioxidant enzymes and by increasing contents of the antioxidants. Xia et al. (2003)¹⁸ reported that Se is crucial in several enzymes with physiological antioxidant properties, including GSH-Px and thioredoxin. Besides, the ability of Se to reduce Hg toxicity has been extensively investigated. It has been demonstrated that HgCl₂ lower the activity of the selenoenzyme GSH-Px in the renal mitochondria after prolonged treatment. Their direct inhibitory action, presumably via covalent reaction with the selenol group of the selenocysteine residue¹⁹ is one mechanism whereby they impair the activity of GSH-Px and possible other selenoenzymes, following prolonged exposure. Selenite treatment prevented HgCl₂ induced decline in GSH-Px activity in the renal mitochondria of rats²⁰. The protective effect of Se against Hg induced nephrotoxicity may be related to the formation of a Se-Hg complex. This conclusion is based on previous studies demonstrating that pre-treatment with sodium selenite increased whole retention of Hg, conceivably due to the formation of inert Se-Hg complexes²¹ and the complexes reduced the availability of Hg²⁰. Yoneda and Suzuki (1997)²² also demonstrated that Se forms an equimolar complex with Hg in the plasma which subsequently binds to selenoprotein P. simultaneous administration equimolar doses of sodium selenite prevented not only methyl mercury induced increased of oxidized glutathione, inhibition of GSH-Px in kidney²³, but also histological and functional damage in kidney as well.²⁴ Although the exact mechanism of mercuric chloride induced nephrotoxicity is not well understood, several studies suggested the involvement of free radicals. Oxidative stress develops when the disturbances between reactive oxygen forms are produced in excess and the factors preventing their harmful effect occur. It has been shown in various studies that mercuric chloride administrations are associated with increased formation of free radicals, and with heavy oxidative stress. This will lead to oxidative damage cell components e.g proteins, lipids and nucleic acids²⁵. HgCl₂ inhibits activities of antioxidant enzymes (GSH-Px and GST) and there is depletion of cellular thiols²⁶ in rat kidney and testes suggesting that HgCl₂ toxicity results from generation of reactive oxygen species. Selenite metabolite are similar to thiols, and therefore compounds that react with thiols are expected to react also with selenols²⁷ may be the one mechanism to restore the activity of antioxidant. Mercuric chloride induced nephrotoxicity is associated with increased level of MDA. MDA and 4-HNE(4-hydroxy-2-nonenal) are the end products produced by the decomposition of W₃ and W₆ polyunsaturated fatty acids^{28,29} due to HgCl₂



administration, platinum sulphhydryl group complexes formed are taken up by renal cells and stabilized by intracellular GSH for several hours, in case of intracellular GSH depletion the complexes undergo the rapid transformation to receive metabolites, this depletion seems to be the prime factor that permits lipid peroxidation and impair antioxidant enzymes. Nephroprotectant by the exogenous selenite might be directly related to its antioxidant activity.

CONCLUSION

Taking into account the results of this study, it is concluded that HgCl₂ administration produces severe nephrotoxicity in rats, increase in the serum: urea, creatinine and uric acid levels, and activity of antioxidant enzymes and GSH level were decreased in kidney along with increase the lipid peroxidation level. Co-administration with sodium selenite, show significant modification in the activity of antioxidant enzymes and histological damage caused by HgCl₂ might be achieved by the use of sodium selenite. We suggest that the use of sodium selenite may offer a beneficial strategy against HgCl₂ toxicity.

REFERENCES

- Fowler BA, Woods JS. Ultrastructural and Biochemical changes in renal mitochondria during chronic oral methylmercury exposure: the relationship to renal function. *Exp Mol pathol*, 27, 1977, 403-412.
- Lund BO, Miller DM, Woods JS. Mercury-induced H₂O₂ production and lipid peroxidation in vitro in rat kidney mitochondria. *Biochem.Pharmacol.* 42, 1991, S181-S187.
- Paller MS. Free radical scavengers in mercuric chloride-induced acute renal failure in the rat. *J. lab.Clin.Med.* 105, 1985, 459-463.
- Girardi G, Elias MM. Mercuric chloride effects on rat renal redox enzymes activities: SOD protection. *Free radic Biol Med.* 18, 1995, 61-66.
- Karin A and Elias ASJ. Rapid induction of cell death by seleniul-compromised thioredoxinreductase I but not by the fully active enzyme containing selenocysteine. *J Biol Chem.* 278, 2003, 15966-15972.
- Cabanero AI, Madrid Y, C'amra C. Selenium and mercury bioaccessibility in fish samples, on in vitro digestion method. *Anal Chim Acta.* 526,2004, 51-61.
- Stadtman, T.C. Selenocysteine. *Annu. Rev. Biochem.* 65, 1996, 83-100.
- Combs, G.F. and Gray, W.P. Chemopreventive agents: selenium. *Pharmacol. Ther.* 79, 1998, 179-192.
- Weckbercker G, Cory JG. ribonucleotidereductase activity and growth of glutathione-depended mouse leukaemia L 1210 cells in vitro. *Cancer let.* 40, 1988, 257-264.
- Buege JA, Aust SD. Microsomal lipid peroxidation. *Methods enzymol.* 105, 1984, 302-10.
- Flohe L, Gunzler WA. analysis of glutathione peroxidase. *Methods enzymol.* 105, 1984, 114-21.
- Habig WH, Pabst Jakoby WB. Glutathione-S-transferase the first step in mercapturic acid formation. *J. Biol Chem.* 249, 1974, 7130-9.
- Bradford MA. Rapid and sensitive method for the quantities of microgram quantities of protein utilizing the principale of protein-dye binding. *Anal biochem.* 72, 1976, 248-54.
- Sener G, Sehirli O, Tozan A, Velioglu-ovuç A, Gedik N, Omrntag GZ. Gingobiloba extract protects against mercury (II)-induced oxidative tissue damage in rats. *food chem Toxicol.* 45, 2007, 543-550.
- Stohs SJ, Bagchi D. Oxidative mechanisms in the toxicity of metal-ions. *Free Radic Biol Med.* 18, 1995, 321-336.
- Pokorny J. Major factors affecting auto-oxidation. In: Chan, H.W.(Ed.), *Autooxidation of unsaturated lipids.* Academic Press. London, 1987, 141-207.
- Yee S, Choi B. Methylmercury poisoning induces oxidative stress in the mouse-brain. *Exp Mol Pathol.* 60, 1994, 188-196.
- Hussain S, Rodgers D, Duhart HA, Ali SF. Mercuric chloride-induced reactiveoxygen species and its effect on antioxidant enzymes in different regions of rat brain. *J Environ Sci Health,* 32B, 1997, 395-409.
- Sasakura C, Suzuki KT. Biological interaction between transition metals (Ag, Cd and Hg), selenide/sulfide and selenoprotein P. *J Inorg Biochem.* 71, 1998, 159-162.
- Chandienne J and Tappel AL. Interaction of gold(I)with the active site selenium-glutathione peroxidase. *J Inorg Biochem.* 20, 1984, 313-325.
- Perottoni J, Rodrigues OED, Paixao MW, Zeni G, Labato LP, Braga AL, Rocha JBT, Emanuelli T. Renal and hepatic ALA-D activity and selected oxidative stress parameters of rats exposed to inorganic mercury and organoselenium compounds. *Food Chem Toxicol.* 42, 2004b, 17-28.
- Yoneda S, Suzuki KT. Detoxification of mercury by selenium by binding of equimolar Hg-Se complex to a specific plasma protein. *Toxicol Appl Pharmacol.* 143, 1997, 274-280.
- Hoffman DJ, Heinz GH. Effects of mercury and selenium on glutathione metabolism and oxidative stress in mallard ducks. *Environ. Toxicol. Chem.* 17, 1998, 161-166.
- Magos L, Clarkson TW, Spanow S, Hudson AR. Comparison of the protection given by selenite, selenomethionine and biological selenium against the renotoxicity of mercury. *Arch Toxicol.* 60, 1987, 422-426.
- Boya P, Pena A, Beloqui O, Larrea E, Conchillo M, and Castelmiz Y. Antioxidant status and glutathione metabolism in peripheral blood mononuclear cells from patients with chronic hepatitis C. *J Hepatol.* 31, 1990, 808-814.
- Sorg O, Schilter B, Howegger P, Monnet-Tschudi F. Increased vulnerability of neurons and glial cells to low concentrations of methylmercury in a prooxidant situation. *Acta neuropathol.* 96, 1998, 621-627.
- Sheen N and Ajjih TA. Prevention of nephrotoxicity induced by antioxidant drug cisplatin, using ganodermalucidum, a medicinal mushroom occurring in south india. *Curr Sci.* 85, 2003, 478-482.
- Seppanen K, Soininen P, Salonen JT, Lotjonen S, Laatikainen R. Does mercury promote lipid peroxidation? An in vitro study concerning mercury, copper, and iron in peroxidation of low densitylipoprotein. *Biol Trace Elem Res.* 101, 2004, 117-132.
- Valko M, Morris H, Cronin MT. Metals, Toxicity and oxidative stress. *Curr Med Chem.* 12, 2005, 1161-1208.

Source of Support: Nil, Conflict of Interest: None.

