



Hemophilia and Its Treatment: Brief Review

Nikisha G. N.^{1*}, Menezes G. A.²

¹CRRI, ²Scientist, Sree Balaji Medical College & Hospital (Bharath University), Chromepet, Chennai, Tamil Nadu, India.

*Corresponding author's E-mail: nikimol@yahoo.co.in

Accepted on: 18-03-2014; Finalized on: 30-04-2014.

ABSTRACT

Hemophilia is a genetic bleeding disorder. Hemophilia A or B is treated by recombinant clotting factor VIII or factor IX and immunosuppressives to prevent formation of alloantibodies and inhibitors. Formation of inhibitors to these factors poses a challenge in treating hemophilia. Plasma derived activated prothrombin complex concentrate and activated recombinant factor VII are used to treat patients with inhibitors. Treatment also varies with situations such as, pregnancy, surgery, and malignancy, as these trigger increased risk of bleeding. This review illustrates the various etiologies of hemophilia, methods of diagnosis, newer treatment options available for hemophilia, treatment in special circumstances and the future of hemophilia treatment. In the present times, long lasting and permanent cure for hemophilia is not available. Replacement by factors provides a temporary cure for hemophilia. Permanent cure may lie in the direction of gene therapy or stem cell therapy which is under development. Few successes are achieved by gene therapy in previous studies providing cure lasting for many months without need for recombinant factor replacement. Further modifications of therapy are required to achieve long lasting and permanent cure for hemophilia.

Keywords: Hemophilia, Bleeding disorders, Treatment of Hemophilia.

INTRODUCTION

Hemophilia derived from the Greek *haima* (Blood) and *philia* (Love), is a genetic bleeding disorder caused by the deficiency of clotting factor-VIII (Hemophilia-A) or factor-IX (Hemophilia-B) or factor-XI (Hemophilia-C). Hemophilia A (also known as classic hemophilia) and B (also known as Christmas disease) are X-linked recessive trait with defective *F8* and *F9* genes in long arm of X chromosome with incidence of one in 5,000 and one in 30,000 males respectively while Hemophilia C (also known as plasma thromboplastin antecedent (PTA) deficiency or Rosenthal syndrome) has autosomal recessive inheritance with defective gene in chromosome 4 and has incidence of one in 1,00,000 males and is more common in Jews of Ashkenazi.

Hemophilia can also be acquired due to the development of autoantibodies directed against the clotting factors. Acquired hemophilia is very rare with incidence of one in 1 million persons.¹⁻³

In Hemophilia there is internal or external bleeding which may be spontaneous or with trivial trauma. It can lead to complications like chronic anaemia, haemarthrosis, intracranial hemorrhage and compartment syndrome. Early diagnosis and management is required to prevent the development of these complications.⁴ The management varies in special situations such as, pregnancy, major surgeries, and malignancy. This review aims in presenting the current advances in diagnosis and treatment of hemophilia and the recent recommendations in the management of special conditions or procedures in hemophiliacs.

CAUSES OF HEMOPHILIA

When there is injury to the blood vessel, the platelets are activated at the site of injury which leads to activation of the clotting factors and formation of fibrin blood clot by the 'Intrinsic pathway' of coagulation. Factor VIII and factor IX are required to activate factor X which in turn activates prothrombin activator which converts prothrombin to thrombin. Thrombin helps in conversion of fibrinogen to fibrin which traps the platelets and forms clot. Factor XI is needed to activate factor IX.⁵

The factors VIII, IX or XI are genetically transferred to the offspring through X chromosome (*F8* and *F9*) and chromosome 4 (*F11*). Any defect in these genes causes absence or reduced production of these factors. Sometimes antibodies labeled as 'inhibitors' to these factors may develop. Inhibitors may develop as a response to therapy with the clotting factors in hemophiliacs or idiopathically in normal subjects without any genetic defect. In these patients, even after therapy with clotting factor, activated partial thromboplastin time (aPTT) and prothrombin time (PT) are prolonged.⁶ Various mechanisms are suggested in the genesis of genetic defect or inhibitor formation.

1. F8 gene mutation

Hemophilia A is caused by mutations in the *F8* gene. In hemophilia A, there are large DNA mutations like intron 22 and intron 1 inversion and many small mutations like missense mutations, nonsense mutations and frameshift mutations.⁷ Factor 8 consists of 2332 amino acids and has domains A1, A2, A3, B, C1, and C2. Severity of the disease is correlated with the type of mutation in these domains.⁸ According to Ryan *et al*, a point mutation causing amino acid substitution N1922S in A3 domain of *F8* gene leads to



defective folding of A3 which ultimately stops the production of factor VIII.⁹ There may also be Arg2150His substitution in C1 domain of factor 8 which results in reduced binding of factor 8 to Von Willebrand factor which leads to reduced stability of factor VIII.¹⁰

2. Lack of f8 mRNA

In some hemophiliacs no defects were found in DNA. There is defective mRNA or lack of mRNA in some of them which prevents the relay of message.¹¹⁻¹³ In these patients, RT-PCR did not give any mRNA corresponding product which is specific for factor 8. This can be due to swift degradation of mRNA due to an unidentifiable mutation in f8 intron.¹⁴

3. F8 inhibitors

Inhibitor to factor VIII develops as a complication to therapy of hemophilia A, mostly in patients with deletion or nonsense mutation in *F8* gene. Alloantibodies develop against the replaced factor VIII and bind to A2 and C2 domain of F8 and inactivate F8 completely.⁶ The incidence of inhibitor development is directly proportional to age of the individual. This may be due to decline of immune regulation function in old age.¹⁵ According to Viel *et al*, mismatched factor VIII replacement by giving H1 and H2 types of factor VIII (present in white population) to black population in whom H3, H4 and H5 are present increases the incidence of developing alloantibodies to factor VIII in black population.¹⁶ These patients don't respond or respond poorly to therapy.

In Acquired Hemophilia, autoantibodies mostly of IgG class are present against factor VIII in subjects who have normal factor VIII gene. It can be idiopathic or can accompany other conditions like autoimmune diseases, cancer and drug ingestion.¹⁷ They bind to A2, A3 or C2 domain of factor VIII and inactivate factor VIII incompletely.⁶ This results in reduced function of factor VIII and increased aPTT even after therapy.

4. Defect in tissue factor pathway

Tissue factor (TF) can initiate an alternate pathway of coagulation in the absence of factor VIII, IX or XI by forming TF-factor VII complex. This becomes TF-Factor VIIa complex which activates factor X to Xa. Absence of factors VIII, IX or XI is more significant when there is deficient TF concentration.^{1,18}

DIAGNOSIS OF HEMOPHILIA

Diagnosis of hemophilia is made on the basis of clinical suspicion and laboratory tests. Mutation in a particular gene may be detected which helps in development of newer modalities of treatment like gene therapy.

a. Clinical diagnosis

Clinical conditions, such as hemarthrosis, intracranial bleeding, excessive bleeding in trivial trauma, prolonged bleeding after surgery and menorrhagia point strongly

towards a diagnosis of hemophilia.¹⁹ Bleeding can be spontaneous in severe cases or only as a response to trauma in mild cases and these are correlated with the laboratory tests.

b. Lab diagnosis

Hemophilia can be diagnosed by coagulation factor assays or activated partial thromboplastin time. Bleeding time, prothrombin time and thrombin time are normal in Hemophilia. Bleeding time is not affected as it shows only platelet function. Prothrombin time is not affected as it depends only on extrinsic pathway of coagulation and factors I, II, V, VII and X. Thrombin time is normal because it depends on fibrinogen.

Activated partial thromboplastin time

This measures the integrity of intrinsic pathway and common pathway.²⁰ In Hemophilia, there is prolongation of aPTT. As factors VIII, IX and XI are part of intrinsic pathway of coagulation along with other factors, aPTT is prolonged in all 3 types of Hemophilia.²¹

Coagulation factors F8/F9 assays

The type of hemophilia and degree of factor activity can be assessed by factor assay. The normal value of factor VIII is 50-150% and factor IX is 50-150%.²² In hemophilia these values are reduced. Knowledge of the exact amount of activity of clotting factors help in grading the severity of the disease and its precise management.¹⁹

Thrombin generation assay

This measures the ability of blood to form thrombin. This is valuable in assessing the response to therapy in hemophiliacs with inhibitors as the conventional coagulation profiles are not useful in them.²³ It shows the overall assessment of hemostasis while aPTT shows only the time taken to form a clot.²⁴ Thrombin generation (TG) maximum peak and lag-phase time are used to find the severity of hemophilia.^{25,26} Severity determined by conventional methods show discrepancies as many severe hemophiliacs may not have severe symptoms and bleeding.²⁷ This is because other pro-thrombotic factors play a role in bleeding in hemophiliacs which can be detected by TG assay.²⁸

Thromboelastography

The integrity of coagulation can be assessed by thromboelastography. R-time and K-time shows the integrity of clotting factors.²⁹ R- time shows the time taken for onset of clotting. PT and aPTT show the integrity of coagulation till this point only. K time is the time taken from end of R until the clot reaches 20mm and it shows the speed of clot formation.³⁰ This helps in monitoring of patients on therapy.

Thrombodynamics test

It is also called spatial clot growth assay.³¹ It has clotting phase and elongation phase which can be activated by intrinsic or extrinsic pathway. In hemophilia, clotting



phase (initiation phase) when activated by intrinsic pathway is delayed whereas activation by extrinsic pathway is normal. Elongation phase (which indicates clot thickening) and rate of clot growth are poorer. Thin friable clots are formed which results in bleeding. These friable clots are formed because of weak positive feedback mechanism involving thrombin.³²

c. Genetic diagnosis

Genetic testing helps in confirmation of diagnosis and identification of carriers. This can increase the index of suspicion and aid in early prenatal diagnosis of fetuses which is crucial in management during delivery.³³

Carrier detection

This is imperative in genetic counseling and in vigilant care of pregnant mother and possibly hemophilic child. Hemophilia A carrier women have wide variation in levels of FVIII and rarely may show mild bleeding tendencies.¹⁹ This is because of X-inactivation in women during embryonic life.³⁴ Hemophilia carrier women are at risk for post partum hemorrhage and hemophilic child is at risk of intracranial hemorrhage. Inverse PCR (I-PCR) or inverse shifting PCR (IS-PCR) can be used to detect intron 1 and intron 22 inversions.^{35,36}

Prenatal diagnosis

Prenatal diagnosis of fetuses with hemophilia is crucial during management of labour. It is done by assessing chorionic villous sampling at 11-14 weeks of gestation or amniocentesis after 15 weeks of gestation or cordocentesis after 20 weeks of gestation.³⁷ This is beneficial in fetuses with a strong family history of moderate to severe hemophilia. In early 1980s hemophilia was prenatally diagnosed mainly by immunoradiometry, factor VIII:CAg and factor VIIIIR:Ag assays.³⁸

1. Amniocentesis

This is done between 15 to 18th week of gestation. Under ultrasound guidance, through maternal abdominal wall, a needle is inserted into the amniotic sac and amniotic fluid containing amniocytes (Foetal cells) is obtained. Direct mutation detection or linkage analysis is used to find affected fetuses.³⁹

2. Cordocentesis

It is also called percutaneous umbilical blood sampling. This is done if the results of other tests are uncertain. Umbilical cord blood is taken using ultrasound guided needle and factors VIII and IX in fetal blood are measured.⁴⁰

3. Preimplantation genetic diagnosis

In this, in-vitro fertilization is done and affected male embryos are identified and only healthy embryos are returned to uterus. It is done by linkage analysis to detect F8 intron 22 inversion in blastomeres obtained by biopsy

of embryo.⁴¹ Cell sample is lysed and PCR is done followed by direct genotyping and mutation analysis.

4. Chorionic villus sampling

It is the most common method of prenatal diagnosis. It is done between 11 and 14th week of gestation.⁴⁰ Chorionic villus is taken through transcervical or transabdominal route under direct ultrasound guidance and indirect or direct mutation analysis or linkage analysis is done to diagnose affected foetus.³⁹

5. Non invasive tests-digital PCR

Invasive tests always carry a risk of foetal loss. To prevent that, non invasive tests are done by analyzing foetal DNA circulating in maternal plasma.^{42,43} Cell free foetal DNA present in maternal plasma has amplified Y chromosomes (Y-PCR) which can be tested.³⁷ Digital PCR known as relative mutation dosage approach can be used for the detection of hemophilia.^{44,37}

Genetic diagnosis in adults

In adults with clinical signs and symptoms of bleeding disorder, genetic testing is done to confirm genetic or other etiology of hemophilia and its appropriate management. It can be done through many methods like direct mutation detection, targeted mutation analysis or inverse PCR.^{19,45}

MANAGEMENT OF HEMOPHILIA

Initially in early 1960's concentrates of antihemophilic globulin⁴⁶ and glycine precipitated factor VIII were used in the treatment of hemophilia.⁴⁷ Then in 1970's and 80's plasma derived products and cryoprecipitate were used to treat hemophilia.⁴⁸ But this increased the incidence of HIV and Hepatitis A and B in hemophiliacs and increased the development of antibodies to factors VIII and IX.⁴⁹⁻⁵¹ By the beginning of 1990, recombinant- DNA derived antihemophilic factors were used to treat hemophilia.^{52,53} Following that desmopressin and immunosuppressive therapy were used.^{54,55} Activated prothrombin complex concentrate was used to treat patients with inhibitors.⁵⁶ In patients with liver cirrhosis or hepatitis due to transfusion, liver transplantation was done which provided good results and cured factor VIII and factor IX deficiencies.⁵⁷⁻⁵⁹

ADVANCES IN MANAGEMENT

In recent times, many advances are made in the treatment of hemophilia. They include modification of time old therapy or breakthrough of new drugs.

a. Immunosuppressives

Steroids alone or steroids with cyclophosphamide or rituximab or cyclosporine are used as first line therapy. People who did not have complete remission with first line are given second line therapy with steroids, cytotoxics and rituximab. According to Collins *et al*, first line therapy with combination of steroids and cyclophosphamide gives a stable remission.⁶⁰



b. Gene therapy

Gene therapy may provide complete lifelong cure for hemophilia. Gene therapy for hemophilia B using adeno-associated viral vector shows promising results in some studies. It provides remission temporarily and further studies are required to achieve permanent cure.^{61,62}

There are some limiting factors to its use like humoral and cellular immune response, toxicity and safety issues which must be rectified in future.⁶³ There is development of antibodies to recombinant proteins in hemophilia A. This can be prevented by giving adenoviral vector expressing factor VIII to neonates as their immune system is immature.⁶⁴

c. Therapy under development

Many new treatment options are tried in the recent times. Bone marrow transplantation and hematopoietic stem cell transplantation are being tried in mice.^{65,66} In vitro studies are done using solulin to increase clot stability in whole blood.⁶⁷ Further studies are required to achieve long lasting and permanent cure for hemophilia.

MANAGEMENT IN SPECIAL SITUATIONS

Management of hemophilia is challenging in special situations like pregnancy, newborn, and surgeries. They require special attention and a need to watch for and prevent undue hemorrhage and complications.

a. Management in pregnancy

Management in carriers of hemophilia during pregnancy requires prenatal diagnosis of hemophilia by chorionic villous sampling or amniocentesis or cordocentesis and careful watch for post partum hemorrhage (PPH).³⁷ According to study by Kadir *et al* 48% of hemophilia carriers had primary PPH. Pregnancy in carriers should be managed by multidisciplinary approach in a tertiary care center.⁶⁸ In case of PPH, fibrinogen supplementation in the form of fresh frozen plasma, cryoprecipitate or fibrinogen concentrate or antifibrinolytic agents like tranexamic acid or recombinant activated factor VII can be given.⁶⁹

In neonates born of carriers, there is increased risk of intracranial or extracranial hemorrhage during delivery.⁷⁰ This can be prevented by prenatal diagnosis of affected foetus and carrier and taking proper precautions. The best possible mode of delivery is under debate. According to James *et al*, the optimal mode of delivery is planned caesarian delivery before labor and this will reduce 85% risk of intracranial hemorrhage.⁷¹ According to Ljung *et al*, planned vaginal delivery without use of instruments like vacuum or forceps is the optimal mode of delivery in hemophilia carriers.⁷² Mode of delivery should be decided based on maternal and fetal factors and individualized.

b. Management in neonate

If hemophilia is suspected in neonate, it should be diagnosed and confirmed by analysis of cord blood. Recombinant factor VIII or factor IX concentrates or fresh

frozen plasma are given to affected children. If intracranial bleed is suspected, factor should be given immediately before confirming the diagnosis by cranial ultrasound or CT or MRI scan.⁷³

c. Management in acute hemarthrosis

In acute hemarthrosis, there is bleeding into the joint which results in rapid joint swelling, pain and reduced movements. Radiological confirmation is not indicated routinely. Therapy with recombinant factors VIII or IX is the first line of treatment. It is followed by resting the joint, applying ice, compression bandage, leg elevation and physiotherapy.^{74,75} In profuse hemarthrosis, arthrocentesis is done after complete correction of factor deficit.⁷⁴ In case of repeated hemarthrosis, synovectomy or angiographic embolization is done.⁷⁶

d. Management in minor/major surgical procedures

Hemorrhage is a common complication during surgery and postoperatively increasing the mortality and morbidity. During surgical procedures in hemophiliacs, activated recombinant factor VII (rFVIIa) is used to achieve hemostasis.^{77,78} In major cardiac surgeries, rFVIIa is used customarily during surgery and postoperatively.⁷⁹ Desmopressin is used prophylactically in open heart surgery patients with hemophilia.⁸⁰

e. Management in malignancy

Cancer in hemophiliacs should be treated in same way as non hemophiliacs. But the risk of bleeding is higher due to therapy induced thrombocytopenia. Replacement therapy should be given as continuous prophylaxis during chemotherapy and radiotherapy when accompanied by thrombocytopenia. If cancer surgery is done, low molecular weight heparin should be given postoperatively to prevent thrombosis.⁸¹

f. Management in patients with inhibitors

Development of inhibitors is a complication of treatment of hemophilia with replacement therapy. It is because of the formation of alloantibodies to factor VIII. This reduces the efficacy of therapy by recombinant factors VIII or IX.⁸² Patients with inhibitors need a bypassing agent to control bleeding. Plasma derived activated prothrombin complex concentrate and activated recombinant factor VII are used as bypassing agent to aid in hemostasis.⁸³

CONCLUSION

In conclusion, early diagnosis of hemophilia and early management helps in improving the quality of living of the patient and prevents early development of complications. Replacement of factors provides effective control of hemophilia for short term. Newer research based options are required to provide long lasting and complete cure for hemophilia which may be possible by gene therapy in the near future.



REFERENCES

- Ataullakhanov FI, Dashkevich NM, Negrier C, Panteleev MA, Factor XI and traveling waves: the key to understanding coagulation in hemophilia?, *Expert Rev. Hematol.*, 6(2), 2013, 111–113.
- Athale AH, Marcucci M, Iorio A, Immune tolerance induction for treating inhibitors in people with congenital haemophilia A or B (Protocol), *Cochrane Database of Systematic Reviews*, Issue 6, 2013, Art. No.: CD010561. DOI: 10.1002/14651858.CD010561.
- Asakai R, Chung DW, Davie EW, and Seligsohn U, Factor XI Deficiency in Ashkenazi Jews in Israel, *N Engl J Med.*, 325, 1991, 153-158
- Sahu S, Lata I, Singh S, and Kumar M, Revisiting hemophilia management in acute medicine, *J Emerg Trauma Shock*, Apr-Jun, 4(2), 2011, 292–298.
- Gailani D, Renné T, The intrinsic pathway of coagulation: a target for treating thromboembolic disease?, *J Thromb Haemost.*, 5, 2007, 1106–1112.
- Franchini M, Gandini G, Paolantonio TD, Mariani G, Acquired Hemophilia A: A Concise Review, *Am J Hematol.*, 80, 2005, 55–63.
- Rossetti LC, Radic CP, Candela M, Pérez Bianco R, de Tezanos Pinto M, Goodeve A, Sixteen novel hemophilia A causative mutations in the first Argentinian series of severe molecular defects, *Haematologica*, Jun, 92(6), 2007, 842-5.
- Shen BW, Spiegel PC, Chang CH, Huh JW, Lee JS, Kim J, Larripa IB, De Brasi CD, The tertiary structure and domain organization of coagulation factor VIII, *Blood*, 111(3), 2008, 1240-7.
- Summers RJ, Meeks SL, Healey JF, Brown HC, Parker ET, Kempton CL, Doering CB, Lollar P, Factor VIII A3 domain substitution N1922S results in hemophilia A due to domain-specific misfolding and hyposecretion of functional protein, *Blood*, 117(11), 2011, 3190-3198
- Jacquemin M, Lavend'homme R, Benhida A, Vanzieleghe B, d'Oiron R, Lavergne JM, Brackmann HH, Schwaab R, VandenDriessche T, Chuah MK, Hoylaerts M, Gilles JG, Peerlinck K, Vermeylen J, Saint-Remy JM, A novel cause of mild/moderate hemophilia A: mutations scattered in the factor VIII C1 domain reduce factor VIII binding to vonWillebrand factor, *Blood*, 96(3), 2000, 958-965
- Naylor J, Brinke A, Hassock S, Green PM, Giannelli F, Characteristic mRNA abnormality found in half the patients with severe haemophilia A is due to large DNA inversions, *Hum Mol Genet*, Nov, 2(11), 1993, 1773-8.
- Naylor JA, Green PM, Rizza CR, Giannelli F, Analysis of factor VIII mRNA reveals defects in everyone of 28 haemophilia A patients, *Hum Mol Genet.*, Jan, 2(1), 1993, 11-7.
- Castaman G, Giacomelli SH, Mancuso ME, Sanna S, Santagostino E, Rodeghiero F, F8 mRNA studies in haemophilia A patients with different splice site mutations, *Haemophilia*, Sep 1, 16(5), 2010, 786-90.
- El-Maarri O, Singer H, Klein C, Watzka M, Herbiniaux U, Brackmann HH, Schröder J, Graw J, Müller CR, Schramm W, Schwaab R, Haaf T, Hanfland P, Oldenburg J, Lack of F8 mRNA: a novel mechanism leading to hemophilia A, *Blood*, Apr 1, 107(7), 2006, 2759-65.
- Hay CR, Palmer B, Chalmers E, Liesner R, Maclean R, Rangarajan S, Williams M, Collins PW, United Kingdom Haemophilia Centre Doctors' Organisation (UKHCDO), Incidence of factor VIII inhibitors throughout life in severe hemophilia A in the United Kingdom, *Blood*, Jun 9, 117(23), 2011, 6367-70.
- Viel KR, Ameri A, Abshire TC, Iyer RV, Watts RG, Lutcher C, Channell C, Cole SA, Fernstrom KM, Nakaya S, Kasper CK, Thompson AR, Almasy L, Howard TE, Inhibitors of factor VIII in black patients with hemophilia, *N Engl J Med.*, Apr 16, 360(16), 2009, 1618-27
- Bouvry P, Recloux P, Acquired hemophilia, *Haematologica*, Nov-Dec, 79(6), 1994, 550-6.
- Repke D, Gemmell CH, Guha A, Turitto VT, Broze GJ Jr, Nemerson Y, Hemophilia as a defect of the tissue factor pathway of blood coagulation: Effect of factors VIII and IX on factor X activation in a continuous-flow reactor, *Proc Natl Acad Sci U S A*, 87(19), 1990, 7623-7.
- Tantawy AA, Molecular genetics of hemophilia A: Clinical perspectives, *The Egyptian Journal of Medical Human Genetics*, 11, 2010, 105–114
- Kamal AH, Tefferi A, Pruthi RK, How to Interpret and Pursue an Abnormal Prothrombin Time, Activated Partial Thromboplastin Time, and Bleeding Time in Adults, *Mayo Clin Proc.*, 82(7), 2007, 864-73.
- Bowyer AE, Van Veen JJ, Goodeve AC, Kitchen S, Makris M. Specific and global coagulation assays in the diagnosis of discrepant mild hemophilia A, *Haematologica*, 98(12), 2013, 1980-7.
- Madan R, Gupt B, Saluja S, Kansra UC, Tripathi BK, Guliani BP, Coagulation profile in diabetes and its association with diabetic microvascular complications., *J Assoc Physicians India*, 58, 2010, 481-4.
- Young G, Sørensen B, Dargaud Y, Negrier C, Brummel-Ziedins K, Key NS, Thrombin generation and whole blood viscoelastic assays in the management of hemophilia: current state of art and future perspectives, *Blood*, 121(11), 2013, 1944-50.
- Salvagno GL, Berntorp E, Thrombin generation testing for monitoring hemophilia treatment: a clinical perspective, *Semin Thromb Hemost*, 36(7), 2010, 780-90.
- Salvagno GL, Astermark J, Lippi G, Ekman M, Franchini M, Guidi GC, Berntorp E, Thrombin generation assay: a useful routine check-up tool in the management of patients with haemophilia?, *Haemophilia*, 15(1), 2009, 290-6.
- Zekavat OR, Haghpanah S, Dehghani J, Afrasiabi A, Peyvandi F, Karimi M, Comparison of Thrombin Generation Assay With Conventional Coagulation Tests in Evaluation of Bleeding Risk in Patients With Rare Bleeding Disorders, *Clin Appl Thromb Hemost.*, Feb 6, 2013
- Vila V, Aznar JA, Moret A, Marco A, Navarro S, Vila C, España F, Assessment of the thrombin generation assay in haemophilia, Comparative study between fresh and frozen platelet-rich plasma, *Haemophilia*, 19(2), 2013, 318-21
- van Dijk K, van der Bom JG, Fischer K, Grobbee DE, van den Berg HM, Do prothrombotic factors influence clinical phenotype of severe haemophilia? A review of the literature, *Thromb Haemost.*, 92(2), 2004, 305-10.
- Milind Thakur, Aamer B. Ahmed, A review of thromboelastography, *International Journal of Perioperative Ultrasound and Applied technologies*, January-April, 1(1), 2012, 25-29.
- Whitten CW, Greilich PE, Thromboelastography: Past, Present, and Future, *Anesthesiology*, 92(5), 2000, 1223-5.
- Soshitova NP, Karamzin SS, Balandina AN, Fadeeva OA, Kretchetova AV, Galstian GM, Panteleev MA, Ataullakhanov FI, Predicting prothrombotic tendencies in sepsis using spatial clot growth dynamics, *Blood Coagul Fibrinolysis*, 23(6), 2012, 498-507
- Ovanesov MV, Krasotkina JV, Ul'yanova LI, Abushinova KV, Plyushch OP, Domogatskii SP, Vorob'ev AI, Ataullakhanov FI, Hemophilia A and B are associated with abnormal spatial dynamics of clot growth, *Biochim Biophys Acta.*, 1572(1), 2002, 45-57.
- Shetty S, Ghosh K, Bhide A, Mohanty D, Carrier detection and prenatal diagnosis in families with hemophilia, *Natl Med J India*, 14(2), 2001, 81-3.



34. Migeon BR, The role of X inactivation and cellular mosaicism in women's health and sex-specific diseases, *JAMA*, 295(12), 2006, 1428-33.
35. He ZH, Chen SF, Chen J, Jiang WY, A Modified I-PCR to detect the factor VIII Inv22 for genetic diagnosis and prenatal diagnosis in haemophilia A, *Haemophilia*, 18(3), 2012, 452-6.
36. Ilić, Nina, Aleksandra Krstić, Miloš Kuzmanović, Dragan Mičić, Nada Konstantinidis, and Marija Guć-Ščekić, Identification of intron 1 and intron 22 inversions of factor VIII gene in Serbian patients with hemophilia A, *Genetika* 45, no. 1, 2013, 207-216.
37. Kadir RA, Davies J, Hemostatic disorders in women, *J Thromb Haemost.*, Jun, 11 Suppl 1, 2013, 170-9
38. Pecorara M, Casarino L, Mori PG, Morfini M, Mancuso G, Scrivano AM, Boeri E, Molinari AC, De Biasi R, Ciavarella N, Hemophilia A: carrier detection and prenatal diagnosis by DNA analysis, *Blood*, 70(2), 1987, 531-5.
39. Chi C and Kadir RA, Antenatal Diagnosis, *Inherited Bleeding Disorders in Women*, 2009, 99-123.
40. Lee CA and Kadir RA, Inherited Bleeding Disorders in Pregnancy: von Willebrand Disease, Factor XI Deficiency, and Hemophilia A and B Carriers, *Disorders of Thrombosis and Hemostasis in Pregnancy*, Springer London, 2012, pp. 115-130.
41. Laurie AD, Hill AM, Harraway JR, Fellowes AP, Phillipson GT, Benny PS, Smith MP, George PM, Preimplantation genetic diagnosis for hemophilia A using indirect linkage analysis and direct genotyping approaches, *J Thromb Haemost.*, 8(4), 2010, 783-9.
42. Avent ND, Refining noninvasive prenatal diagnosis with single-molecule next-generation sequencing, *Clin Chem.*, 58(4), 2012, 657-8
43. Galbiati S, Brisci A, Damin F, Gentilin B, Curcio C, Restagno G, Cremonesi L, Ferrari M, Fetal DNA in maternal plasma: a noninvasive tool for prenatal diagnosis of beta-thalassemia, *Expert Opin Biol Ther.*, Jun, 12 Suppl 1, 2012, S181-7.
44. Lench N, Barrett A, Fielding S, McKay F, Hill M, Jenkins L, White H, Chitty LS, The clinical implementation of non-invasive prenatal diagnosis for single-gene disorders: challenges and progress made, *Prenat Diagn.*, Jun, 33(6), 2013, 555-62.
45. Rossetti LC, Radic CP, Larripa IB, De Brasi CD, Genotyping the hemophilia inversion hotspot by use of inverse PCR, *Clin Chem.*, Jul, 51(7), 2005, 1154-8.
46. Pool JG, Shannon AE, Production of high-potency concentrates of antihemophilic globulin in a closed-bag system: assay in vitro and in vivo, *N Engl J Med.*, Dec 30, 273(27), 1965, 1443-7.
47. Abildgaard CF, Simone JV, Corrigan JJ, Seeler RA, Edelstein G, Vanderheiden J, Schulman I, Treatment of hemophilia with glycine-precipitated Factor VIII, *N Engl J Med.*, Sep 1, 275(9), 1966, 471-5.
48. Biggs R, Haemophilia treatment in the United Kingdom from 1969 to 1974, *Br J Haematol.*, Apr, 35(4), 1977, 487-504.
49. Biggs R, Jaundice and antibodies directed against factors VIII and IX in patients treated for haemophilia or Christmas disease in the United Kingdom, *Br J Haematol.*, Mar, 26(3), 1974, 313-29.
50. Lewis JH, Maxwell NG, Brandon JM, Jaundice and hepatitis B antigen/antibody in hemophilia, *Transfusion*, May-Jun, 14(3), 1974, 203-11.
51. Ragni MV, Winkelstein A, Kingsley L, Spero JA, Lewis JH, 1986 update of HIV seroprevalence, seroconversion, AIDS incidence, and immunologic correlates of HIV infection in patients with hemophilia A and B, *Blood*, Sep, 70(3), 1987, 786-90.
52. White GC 2nd, McMillan CW, Kingdon HS, Shoemaker CB, Use of recombinant antihemophilic factor in the treatment of two patients with classic hemophilia, *N Engl J Med.*, Jan 19, 320(3), 1989, 166-70.
53. Schwartz RS, Abildgaard CF, Aledort LM, Arkin S, Bloom AL, Brackmann HH, Brettler DB, Fukui H, Hilgartner MW, Inwood MJ, Human recombinant DNA-derived antihemophilic factor (factor VIII) in the treatment of hemophilia A, *N Engl J Med.*, Dec 27, 323(26), 1990, 1800-5.
54. Rodeghiero F, Castaman G, Mannucci PM, Prospective multicenter study on subcutaneous concentrated desmopressin for home treatment of patients with von Willebrand disease and mild or moderate hemophilia A, *Thromb Haemost.*, Nov, 76(5), 1996, 692-6.
55. Shaffer LG, Phillips MD, Successful treatment of acquired hemophilia with oral immunosuppressive therapy, *Ann Intern Med.*, Aug 1, 127(3), 1997, 206-9.
56. Leissingner CA, Use of prothrombin complex concentrates and activated prothrombin complex concentrates as prophylactic therapy in haemophilia patients with inhibitors, *Haemophilia*, Sep, 5 Suppl 3, 1999, 25-32.
57. Delorme MA, Adams PC, Grant D, Ghent CN, Walker IR, Wall WJ, Orthotopic liver transplantation in a patient with combined hemophilia A and B, *Am J Hematol.*, Feb, 33(2), 1990, 136-8.
58. Bontempo FA, Lewis JH, Gorenc TJ, Spero JA, Ragni MV, Scott JP, Starzl TE, Liver transplantation in hemophilia A, *Blood*, Jun, 69(6), 1987, 1721-4.
59. Gordon FH, Mistry PK, Sabin CA, Lee CA, Outcome of orthotopic liver transplantation in patients with haemophilia, *Gut*, May, 42(5), 1998, 744-9.
60. Collins P, Baudo F, Knoebl P, Lévesque H, Nemes L, Pellegrini F, Marco P, Tengborn L, Huth-Kühne A: EACH2 registry collaborators, Immunosuppression for acquired hemophilia A: results from the European Acquired Haemophilia Registry (EACH2), *Blood*, Jul 5, 120(1), 2012, 47-55.
61. Nathwani AC, Tuddenham EG, Rangarajan S, Rosales C, McIntosh J, Linch DC, Chowdhary P, Riddell A, Pie AJ, Harrington C, O'Beirne J, Smith K, Pasi J, Glader B, Rustagi P, Ng CY, Kay MA, Zhou J, Spence Y, Morton CL, Allay J, Coleman J, Sleep S, Cunningham JM, Srivastava D, Basner-Tschakarjan E, Mingozi F, High KA, Gray JT, Reiss UM, Nienhuis AW, Davidoff AM, Adenovirus-associated virus vector-mediated gene transfer in hemophilia B, *N Engl J Med.*, Dec 22, 365(25), 2011, 2357-65
62. Ponder KP, Hemophilia gene therapy: a Holy Grail found, *Mol Ther.*, Mar, 19(3), 2011, 427-8
63. High KA, The gene therapy journey for hemophilia: are we there yet?, *Blood*, Nov 29, 120(23), 2012, 4482-7.
64. Hu C, Cela RG, Suzuki M, Lee B, Lipshutz GS, Neonatal helper-dependent adenoviral vector gene therapy mediates correction of hemophilia A and tolerance to human factor VIII, *Proc Natl Acad Sci U S A*, Feb 1, 108(5), 2011, 2082-7.
65. Follenzi A, Raut S, Merlin S, Sarkar R, Gupta S, Role of bone marrow transplantation for correcting hemophilia A in mice, *Blood*, Jun 7, 119(23), 2012, 5532-42.
66. Ramezani A, Hawley RG, Correction of murine hemophilia A following nonmyeloablative transplantation of hematopoietic stem cells engineered to encode an enhanced human factor VIII variant using a safety-augmented retroviral vector, *Blood*, Jul 16, 114(3), 2009, 526-34.
67. Foley JH, Petersen KU, Rea CJ, Harpell L, Powell S, Lillicrap D, Nesheim ME, Sørensen B, Solulin increases clot stability in whole blood from humans and dogs with hemophilia, *Blood*, Apr 12, 119(15), 2012, 3622-8.
68. Huq FY, Kadir RA, Management of pregnancy, labour and delivery in women with inherited bleeding disorders, *Haemophilia*, Jul, 17 Suppl 1, 2011, 20-30.
69. Kadir RA, Davies J, Hemostatic disorders in women, *J Thromb Haemost*, 11 (Suppl. 1), 2013, 170–9.



70. Richards M, Lavigne Lissalde G, Combesure C, Batorova A, Dolan G, Fischer K, Klamroth R, Lambert T, Lopez-Fernandez M, Pérez R, Rocino A, Fijnvandraat K, European Haemophilia Treatment and Standardization Board, Neonatal bleeding in haemophilia: a European cohort study, *Br J Haematol.*, Feb, 156(3), 2012, 374-82
71. James AH, Hoots K, The optimal mode of delivery for the haemophilia carrier expecting an affected infant is caesarean delivery, *Haemophilia*, May, 16(3), 2010, 420-4.
72. Ljung R, The optimal mode of delivery for the haemophilia carrier expecting an affected infant is vaginal delivery, *Haemophilia*, May, 16(3), 2010, 415-9.
73. Chalmers E, Williams M, Brennand J, Liesner R, Collins P, Richards M, Paediatric Working Party of United Kingdom Haemophilia Doctors' Organization, Guideline on the management of haemophilia in the fetus and neonate, *Br J Haematol.*, Jul, 154(2), 2011, 208-15.
74. Simpson ML, Valentino LA, Management of joint bleeding in hemophilia, *Expert Rev Hematol.*, Aug, 5(4), 2012, 459-68.
75. Forsyth AL, Zourikian N, Valentino LA, Rivard GE, The effect of cooling on coagulation and haemostasis: Should "Ice" be part of treatment of acute haemarthrosis in haemophilia?, *Haemophilia*, Nov, 18(6), 2012, 843-50.
76. Hermans C, De Moerloose P, Fischer K, Holstein K, Klamroth R, Lambert T, Lavigne-Lissalde G, Perez R, Richards M, Dolan G, European Haemophilia Therapy Standardisation Board, Management of acute haemarthrosis in haemophilia A without inhibitors: literature review, European survey and recommendations, *Haemophilia*, May, 17(3), 2011, 383-92.
77. Boadas A, Fernández-Palazzi F, De Bosch NB, Cedeño M, Ruiz-Sáez A, Elective surgery in patients with congenital coagulopathies and inhibitors: experience of the National Haemophilia Centre of Venezuela, *Haemophilia*, May, 17(3), 2011, 422-7.
78. Shapiro A, Cooper DL, US survey of surgical capabilities and experience with surgical procedures in patients with congenital haemophilia with inhibitors, *Haemophilia*, May, 18(3), 2012, 400-5.
79. Warren O, Mandal K, Hadjianastassiou V, Knowlton L, Panesar S, John K, Darzi A, Athanasiou T, Recombinant activated factor VII in cardiac surgery: a systematic review, *Ann Thorac Surg.*, Feb, 83(2), 2007, 707-14.
80. Cattaneo M, The use of desmopressin in open-heart surgery, *Haemophilia*, Jan, 14 Suppl 1, 2008, 40-7.
81. Mannucci PM, Schutgens RE, Santagostino E, Mauser-Bunschoten EP, How I treat age-related morbidities in elderly persons with hemophilia, *Blood*, 114(26), 2009, 5256-63.
82. Kempton CL, White GC 2nd, How we treat a hemophilia A patient with a factor VIII inhibitor, *Blood*, Jan 1, 113(1), 2009, 11-7.
83. Ingerslev J, Sørensen B, Parallel use of by-passing agents in haemophilia with inhibitors: a critical review, *Br J Haematol.*, Oct, 155(2), 2011, 256-62.

Source of Support: Nil, Conflict of Interest: None.

