



Biomaterials from Sponges, Ascidians and Other Marine Organisms

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ABSTRACT

Biomaterial, one of the most interesting fields of modern science deals with the biologically derived materials or substances that are used within a biological system. Although according to the sources two types are there yet in terms of several advantageous properties natural biomaterials are far more important than synthetic biomaterials. Among the natural sources the newest one and also the most potent one identified to be is the marine environment. The most undiscovered part of the earth, this marine environment is the powerhouse of millions of undiscovered species generating the greatest biodiversity zone. Here in these article biomaterials from various marine organisms like sponges, ascidians, crustaceans, sessile organisms, corals, actinobacteria, seaweeds, fungi have been reported. The uses of some of the important biomaterials are also discussed. For future work first an appropriate characterisation method should be developed for isolating the sample organisms from marine environments. Extreme environments underwater like hydrothermal vent, hyper saline region etc and also various thermocline and halocline environment provide unusual microorganisms producing uncommon bioactive compounds. Development of new strategies coupled with chemical synthesis method could pave the way for future discovery in this actively growing field.

Keywords: Ascidians, Biomaterials, seaweeds, sponges.

INTRODUCTION

Biomedical engineering is an area that deals with traditional engineering approaches to improve the fundamental quality of life by solving various problems in the fields of life science and medicine. Interestingly the subject of biomaterial science tends to be the answer for the majority of problems associated with biomedical engineering. Biomaterial may be defined as 'a material intended to interface with biological systems to evaluate, treat, augment or replace any tissue, organ or function of the body'.¹ Although there exists a debate within the scientific community regarding the exact definition of biomaterials, whether it is materials those interact with the biological systems or these are the materials which have been derived from biological organism, yet it's of no confusion that biomaterial science is concerned about the interaction between the biological metabolism and these substances. An ideal biomaterial should exhibit some properties like non toxicity, it should produce appropriate host response, should avoid adverse tissue reactions and rejections, biological or corrosion resistance to degradation, should possess sufficient amount of mechanical and rheological strength etc. Biomaterial industry is the emerging industry of 21st century. It has been observed that this domain accounts for 2-3% of the overall health expenses in developed countries.² Significant industrial growth is expected within the next 20 years ultimately generating a multi-billion dollar industry.

Types of Biomaterials

Biomaterials can be derived either from nature or synthesised artificially in the laboratory using metallic

components, polymers, ceramics or composite materials. These materials are used for various biomedical applications. Some of their uses have been listed in table 1.

Table 1: Medical applications of synthetic biomaterials

Category	Material	Application
Metallic components	Stainless steel	Fracture fixation ³
	Nickel-Titanium	Bone plates ⁴
	Gold alloy	Dental restoration ⁵
	Co-Cr-Mo-Ni alloy	Bone and joint replacement ⁶
	Hg-Ag amalgum	Dental restoration ⁷
Polymers	Polyethylene	Joint replacement ⁸
	Polyethylene terephthalates	Vascular prosthesis ⁹
	Polypropylene	Sutures ¹⁰
	Polyesters	Drug delivery system ¹¹
	Poly tetra fluoro ethylene	Soft-tissue augmentation ¹²
	Silicones	Soft tissue replacement ¹³
	Hydrogel	Ophthalmology ¹⁴
Ceramics	Zirconia	Joint replacement ¹⁵
	Alumina	Dental implant ¹⁶
	Calcium phosphate	Bone repair ¹⁷
Composites	Bisphenol A-glycidyl-quartz/ silica filler	Dental restorations ¹⁸



Natural

When the biomaterial comes from the natural sources, then it is termed as natural biomaterial. Throughout the human civilization biological structures and natural substances have always been a model system for solving critical challenges in engineering, building and material science. Nature has always provided a wide array of substances with significant diversity in structure and functions.¹⁹ The bio mimetic potentiality of these substances can also be applied to the biomaterial research arena. Although synthetic biomaterials have been commercialised for various biomedical application they have certain disadvantages including toxicity and reduced ability of tissue remodelling. The major advantage of nature derived biomaterials is the increased chances of biocompatibility, biodegradability, tissue remodelling, cell adhesion, proliferation, differentiation and less toxicity. Furthermore wide variety of organisms of earth provides a great opportunity to discover their biomaterial potentiality. Among the natural sources the most predominant one is marine environment. From the very first day of its existence, the earth contains marine environment. The age of terrestrial organism is much younger in comparison to marine organisms. Which is why there exists much higher biochemical diversity in the marine realm and this diversity is directly proportional to the probability of getting novel biomaterials.

Diversity of Marine Organisms

Aquatic environment is the main component of natural sources. Aquatic organisms can be divided into two types fresh water organisms and marine organisms. 72% of earth has been covered by aquatic systems. Among these 97% of earth's water content is within the oceans. Life begun at sea and afterwards many of the species of marine system were unable to make the transition to the terrestrial life. Consequently marine organisms have higher genetic diversity than freshwater and terrestrial species.²⁰ As an example, all except one of the 35 animal phyla are found in the sea and surprisingly half of these are primarily marine.²¹ A comparative study showed that average heterozygosity was considerably less in freshwater fish subspecies with respect to marine population.²² Elliott et al.,²³ showed that genetic diversity of the Orange Roughly needs only 200 migrants per year to be maintained which infers that marine organisms probably exchange up to 100 times more migrants than freshwater species in every generation. Several theories suggest that there may be 5 million²⁴ to 10 million²⁵ under scribed deep sea species. Various groups of the sea include Echinodermata (starfish and sea urchins), Brachiopoda (lamp shells), Bryozoa (moss animals), Sipunculida (Peanut worms), Polychaeta (bristle worms), Pycnogonida (Sea spiders), Tunicata (sea squirts) and Ctenophora (comb jellies) and the three large groups that are confined to the marine environment only are Cnidaria (anaemon, corals), Crustaceae (crabs, barnacles etc.) and the Mollusca (snails, slugs etc.).

Marine Organisms: A Source of Different Biomaterials

The incredible diversity of the marine organisms has become the reason of attraction for the entire scientific community in the field of biomaterial research. Starting from the marine sponges and ascidians up to the mussels, barnacles, crustaceans etc. have been already reported as potential sources of commercially important novel biomaterials. According to size they can be broadly classified into two categories macro-organisms and micro-organisms.

a) Macro-organisms

Sponges

Sponges are the most primitive of all the multi cellular organisms that have been existing 700-800 million years and approximately among 15000 sponge species, only 1% lives in the freshwater, remaining in the marine habitat.²⁶ Sponges are excellent research subjects due to their fibrous skeletons²⁷ and mineralized spicules, containing amorphous silica²⁸ or calcium carbonate. The skeletal formation of *demospongiae*, *hexactinellida*, *calcareae* etc. can be attributed as natural bio composite material based on rigid glass or calcium carbonate consequently increasing the possibility of developing a novel bone replacement biomaterial. Recently production of active metabolites by various species of sponges has gained attention. Sponges produce various toxins to compete for space with other species, to repel and escape the predators and for intra community communication. Discovery of various bioactive compounds^{29,30} including anticancer, antichemotactic³¹ and antifouling agents³² suggests production of potential pharmaceutical agents.

Ascidians

These are the marine invertebrate organisms which are filter feeders. Their habitat includes all over the world generally in shallow water having salinity more than 2.5%. Thousands of natural marine products have been found in ascidians. In a similar manner as that of sponge they also require synthesis of chemical substances for survival in highly dangerous predation environments. Identification of these compounds has led to the discovery of many potent drugs. Bioactive compounds of different types of polyketides, hydrocarbons, enediynes, peptides, alkaloids, terpenes, tubercidins etc. have been isolated from ascidians.³³ A composite skeletal tissue has been reported from ascidian which consists of amorphous and crystalline calcium carbonate in two separated domains having an organic sheath in between. The calcitic layer consists of characteristic magnesium.³⁴ Moreover the therapeutic potential of glycol amino glycans has been proved. One of the current materials of importance chondroitin sulphate has been isolated from marine ascidians.³⁵ Ascidians also provide the source for important bioactive compounds like depsipeptide with anti tumorigenic property,³⁶ lamellarian³⁷ and imidazoles alkaloids³⁸ and cyclic peptides.³⁹



Table 2: Biomaterials from marine sponges and ascidians

Sponges	Ascidians
Silica ⁴⁰	Cellulose ³⁴
Aragonite ⁴⁰	Calcite ³⁴
Chitin ⁴⁰	Calcium Carbonate ³⁴
Collagen ⁴¹	Chondroitin Sulfate ³⁵
Stevensin ⁴²	Antitumor depsipeptide ³⁶
3-Akylpyridinium ²⁹	Lamellarian alkaloids ³⁷
3-Alkylpyridine ²⁹	Cyclic peptides ³⁹
Terpenes ⁴³	Imidazole alkaloids ³⁸
Sterols ⁴³	Polyketides ³³
Cyclic/ linear peptides ⁴³	Odd chain hydrocarbons ³³
Alkaloids ⁴³	Enediynes ³³
Fatty acids ⁴³	Peptides ³³
Peroxides ⁴³	Alkaloids ³³
Amino acid derivatives ⁴³	Terpenes ³³
Antibiotics ⁴³	Tuberidins ³³

Crustaceans

Marine crustaceans such as crabs, shrimps, krill's, lobsters, prawns etc. provide a great source of two important polysaccharides chitin and chitosan. Chitin is the second most naturally abundant polymer after cellulose.⁴⁴ Chitin forms the crystalline micro fibrillar structure of the exoskeleton of arthropods and fungi and some yeast.⁴⁵ Although fungal chitin is more uniform than animal one yet isolation of fungal chitin is difficult due to its association with various other polysaccharides e.g. mannan, poly galactosamine etc.⁴⁶ Structurally a linear chain of (1-2) linked 2-acetamide-2-deoxy- β -D-glucopyranose units.⁴⁷ The chain arrangement of crude extracted chitin produces two forms α and β .⁴⁸ Chitosan is none other than another form of chitin which is produced by achieving a certain a degree of deacetylation. It is the only cationic pseudo natural polymer as a result of which it is widely used in many applications due to its unique characteristics.⁴⁹ In aquaculture sector mainly shrimp and prawn are predominant. Shrimp chitin antibacterial activity has also been reported.⁵⁰

Sessile Organisms

These are the incredible class of marine organisms including mussels, barnacles, sea anemones, urchins, starfishes, tube worms, limpets etc. They have the unique ability for the production of adhesive substances for attachment to the substratum of the rocks or clinging to the coastlines against the turbulent flow of water of the sea or oceanic environment.⁵¹ This attachment is particularly important for the purpose of reproduction, signal for food supply, escape from the predators etc. Mussels generate a byssal thread and an adhesion plaque thereby attaching via specialised adhesive proteins consist of hydroxyproline and dihydroxy phenylalanine.⁵² On specialised high energy surfaces mussels sticks with a

strength of almost 300 Kilopascals.⁵³ The threads are composed of collagen resembling proteins whereas the plaques are of cross linked protein matrix. One major constituent of these adhesives is that 3,4-dihydroxyphenylalanine (DOPA) an uncommon amino acid produced by the post translational modification of tyrosine.⁵⁴ Absence of this amino acid causes the loss of adhesion ability of the proteins.⁵⁵ The property of adhesion ability along with special characteristics of biocompatibility and biodegradability makes it suitable candidate for industrial and medical adhesives.⁵⁶

Corals

The oceanic environment provides a wonderful combination of calcified sessile and free living organisms containing a wide array of micro scale organised skeletal materials. These structural materials are usually made of calcite and aragonite which are nothing but the crystalline form of CaCO_3 and silicate materials. In biomedical applications coralline calcite or aragonite has been successfully applied for replacement of fractured bone due to their ability of forming strong chemical bond with in vivo soft tissue and bones.⁵⁷ The specialised advantage of using coralline apatite is increased chances of resorption by the attack of enzymes like carboanhydrase.⁵⁸ Secondly its porous crystalline structure permits the blood supply for the newly formed bones by allowing in growth of blood vessels ultimately infiltrating the implant.⁵⁹ Use of porous coral apatite has also been established for the *in vitro* culture of prokaryotic and eukaryotic cells. Interestingly coralline (*Goniopora*, *Millepora*) calcium carbonate converted hydroxy apatite constructs also show the ability of bone differentiation.^{59,60}

Seaweeds

Loosely the term seaweed indicates the class of marine algae comprising red, brown and green algae. From the biomaterial science perspective marine algae is an excellent source of commercially important biomaterials. Polysaccharides like agar, alginate, fucoidan and carrageenan are obtained from algae. Agar is a typical linear copolymer of hydrophobic basic alternating repeating units of 1,3-linked β -D-galactopyranose and 1,4-linked 3,6-anhydro- α -L-galactopyranose.⁶¹ This structural polysaccharide is found to be present in the cell wall of red algae especially within the genera *Gelidium*⁶² and *Gracilaria*.⁶³ Along with these two *Ceramium*, *Acanthopeltis* and *Pterocladia* are the main sources of commercial agar.⁶⁴ Alginate is another kind of commercially important biopolymer having its predominant existence in the cell wall of brown algae *Laminariapallida*, *Laminaria japonica*, *laminariadigitata*, *Ascophyllum* and *Macrocystis*.⁶⁵ The component of polysaccharide in the sea weed has been used for the production of bioplastics.⁶⁶ For various biomedical applications and enzyme immobilization purposes alginate has been proved to be worthwhile. Carrageenans are high molecular weight polysaccharides composed of



D-galactose backbone. Among the 15 different structures, the major sources of industrially relevant κ -carrageenan, ι -carrageenan and λ -carrageenan are red seaweed *Kappaphycus alvarezii*, *Eucheumaspinosum* and *Gigartina species*.⁶⁷ Another specialised sulphated polysaccharide fucoidans can be extracted from brown algae. Several adventitious properties of algal fucoidans over the marine invertebrates like higher anticoagulant activity⁶⁸ resulted in the use of marine brown algae such as Komby, hijiki, derwrack, mozuku as its major source. Moreover marine algae of Rhodophyta division has been successfully implemented as a starting material for the extraction of calcium carbonate to produce hydroxyapatite.⁶⁹

Micro-organisms

Actinobacteria

Actinobacteria, is one of the largest bacterial phyla consisting of mainly organisms with high G+C content.⁷⁰ Although primarily thought of as a soil bacteria, their presence in more numbers has been established in freshwater⁷¹ and marine sediments.⁷² This class of bacteria consists of most economically significant prokaryotic organisms which produce almost half of the bioactive compounds in the Antibiotic Literature Database.⁷³ The members of this group show excellent physiological, morphological and metabolic diversity as evident by the production of various secondary metabolites and extracellular enzymes.⁷⁴ Extreme environment of the marine actinobacteria habitat starting from temperature below 0°C, extreme pressure (up to 1100 atm approximately) at the deep sea floor to highly acidic conditions with extremely hot temperature (100°C) near the hydrothermal vents may be the possible reason for the production of wide classes of bioactive materials.⁷⁵

Fungi

Meristematic black yeast, these are the fungi that resides in hyper saline waters, represented by halophilic *Hortaea werneckii*, *Phaeothecatriangularis*, *Trimmatostroma*, halotolerant *Aureobasidium pullulans*,⁷⁶ and different species of the genus *Cladosporium*, taxonomically and phylogenetically closely related to black yeasts.⁷⁷ Cellular dehydration due to extracellular freezing and hyper saline stress can accumulate many solutes, which can be a good cryoprotectants and osmolytes. Glycerol and mycosporine like amino acids (MAAs) are the potent substances which functions as water soluble UV-absorbing (310-320nm) compound.⁷⁸ The pure compound from *Collema cristatum* prevented pyrimidine dimer formation and cell destruction by absorption of UV-B radiation.

Uses of marine biomaterials

In recent years biomolecules from marine sources have gained wide attention due to their bio mimetic potentiality. With their incredible structural similarity as

well as biocompatibility, they have found significant usage in various biomedical applications.

Table 3: Biomaterials obtained from other marine organisms

Organisms name	Biomaterial obtained	References
Crustaceans		
Crab	Chitin	79
	CaCO ₃	80
Shrimp	Chitin	81
	Chitosan	82
Oyster	Proteolytic enzyme	83
	Chitin	84
Squid pen	Chitin	85
	Hydroxyapatite	86
	Chitosan	87
	Chitin Chitosan	88 89
Sessile organisms		
Mussels	Adhesive protein	90
Barnacles	Barnacle cement	91
Sea anemones	Silk like protein	92
	Adhesion protein	93
Sea urchins	Adhesive protein	94
	Calcite	95
	Calcium Carbonate	96
Starfishes	Adhesion protein	97
	Collagen	98
	Calcium Phosphate	99
Tube worms	Chitin	100
	cement protein	101
Corals		
Bamboocoral	Calcium carbonate	102
Octocoral	Calcite	103
Actinobacteria		
<i>Streptomyces sp.</i>	Polyketide	104
	Pyrrroloiminoquinone	105
	Pyrrrolizidine	106
<i>Actinomycete sp.</i>	Indolocarbazole	107
	Isoprenoid	108
<i>Actinomadura sp.</i>	Indolocarbazole	109
	Phenazine	110
Seaweed		
Red algae	Agar	62
	Carrageenan	67
	Calcium Carbonate	69
Brown algae	Alginate	111
	Fucoidans	68
Green algae	Bioplastics	66
Marine fungi		
Black yeast	Glycerol	112

a) Calcium carbonate and hydroxyapatite

Calcium and phosphate composite material such as hydroxyapatite has some special importance in the biomaterial science because of their analogy with the mineral components of the bone. Although calcium carbonate is the more abundant form of calcium in the marine environment yet calcium phosphate composites like hydroxy apatite is of more importance from the application point of view. Calcium carbonate can be found in the crystalline form of calcite or aragonite in octocorals, bamboocorals etc. Though there is variety of sources yet the unique properties of coral calcium carbonate like porosity, architecture, pore interconnectivity etc. made them the primary candidate for orthopaedics and dentist applications.¹¹³ These criteria are important for bone regeneration purpose. Several different study indicated one major limitation of calcium carbonate for using it as a possible bone substituent because of its faster resorption.¹¹⁴ For this purpose now the focus has been shifted to the production of calcium phosphate compounds such as hydroxy apatite which has all the properties of calcium carbonate but with increased efficiency in terms of resorption. Several synthetic methods such as hydrothermal,¹¹⁵ sol-gel,¹¹⁶ chemical precipitation,¹¹⁷ reverse microemulsion¹¹⁸ and polymer assisted method¹¹⁹ etc. have been reported already. Coral is now also widely used for the hydroxy apatite production. Porous hydroxy apatite microstructure produced from coralline carbonate showed the advantage of circulation of body fluids and the capability of firm attachment with the tissue substratum.¹²⁰ Hydroxyapatite ceramic carrier has been successfully applied in repairing tibial gaps in sheep model by autologous transplantation of bone marrow osteoprogenitor cells.¹²¹ For dentist application biomimetic nanohydroxyapatite toothpastes have been found to be useful for the remineralisation of enamel surface.¹²² Coralline hydroxyapatite implant also has the ability of cosmetic reconstruction without any risk of infection.¹²³

b) Biosilica

Bio derived silica, commonly termed as biosilica, is made up of amorphous silica and is produced in many marine organisms such as sponges, diatoms, choanoflagellates and radiolarians. Among all these bio silicifying organisms' sponges and diatoms are the two most important sources. The process of biosilica formation is mediated by the enzyme silicatein through the formation of various concentric layers.¹²⁴ The naturally occurring silica has been identified as a bio composite with high flexibility and toughness which could be credited to their layer based structural organisation and hydrated nature.¹²⁵ Silica based biomaterials have additional properties of biocompatibility and adventitious reaction product formation after implantation.¹²⁶ The bioactive glasses based on silica has a wide range of applications involving bone tissue replacement, soft tissue augmentation, maxillofacial reconstruction, urological

tissue augmentation, ossicle replacement etc. Biosilica can offer the properties of nanotoxicity, high stability and a hydrophilic and porous nanoscale structure useful for applying in the encapsulation of drugs.¹²⁷ Biosilica induces the expression of the important mediator BMP 2 which is responsible for inducing the differentiation of bone forming progenitor cells and also inhibits the function of osteoclasts, thereby acting as a promising candidate for treatment of the disease of osteoporosis.¹²⁸ Recently silicon substituted hydroxyapatites are being developed which increases the bioactivity and mechanical properties of bone substituted material. The increased bioactivity leads to excellent osteo integration by promoting the reaction between bone and implant owing to increase in solubility of the material.¹²⁹

c) Alginate

One of the most important biomaterials which has found number of applications in biomedical engineering due to its unique properties of gelation and biocompatibility. The alginate hydrogels are especially the subject of interest. The extraordinary property of structural mimicking of extracellular matrices of the tissues has led to an extensive use of alginate hydrogels for the purpose of wound healing, drug delivery and tissue engineering. Hydrogels are basically hydrophilic polymeric networks which have the capacity to accept water thousands of times of their dry weight. This hydrogel can play a significant role for devising a controlled drug delivery strategy.¹³⁰ Polycaprolactone,¹³¹ chitosan¹³² and carbon nanotube¹³³ have been successfully applied for drug delivery with alginate hydrogels. Specific tissue engineering systems have been produced based on the alginates such as artificial pancreas where alginate is used for islet cell encapsulation, alginate delivery vehicle mediated bone regeneration system, bio artificial liver etc.¹³⁴ Moreover the alginate has been found to be a good candidate for skeletal mussel regeneration. The hydrophilic property of alginate allows it to retain a moist environment by adsorption and desorption process and subsequently alginate hydrogel can be used efficiently for the purpose of wound dressing.¹³⁵ Cell immobilization is another promising application of alginate gel systems. Entrapment of the cells within the gel allow them to be cultivated within different types of bioreactors to obtain high cell densities.¹³⁶

d) Chitosan

Chitosan, a linear polysaccharide comprise of randomly distributed glucosamine residues, can be obtained from chitin by enzymatic or chemical method. Chitosan is one of the major sources of surface pollution in coastal areas. Recently it has been established that due to its excellent coagulating properties it can be used for the purpose of wastewater treatment.¹³⁷ Important properties of biocompatibility, non-toxicity with the specialised advantage of antimicrobial activity has lead to the implementation of chitosan based films for food packaging which can be an appropriate alternative of



commercially available packaging materials which hampers the environment.¹³⁸ Repeated reuse of these biomaterials provide an attractive route for waste management. These chitosan films have also been established as a potential local drug delivery system.¹³⁹ Uses of chitosan include preparation of an immobilizing and permeabilizing matrix for microorganisms, delivery system for nucleic acids,¹⁴⁰ hollow fiber membranes for removal of ions,¹⁴¹ matrix for artificial skin, tablet binder, plant cell culture and surgical sutures.¹⁴² Chitosan is also used as a composite material by blending with hydroxyapatite or producing hybrid with alginate for bone tissue engineering.¹⁴³

e) Fucoidan

One of the most important biomaterials found mainly in the marine brown algae is the sulphated polysaccharide fucoidan. It has a significant role in controlling the acute and chronic inflammatory response by the mechanism of enzyme and complement cascade inhibition. Fucoidan has been found to contain several interesting properties that may lead to prevention of the disease of cancer. Evidence has been discovered that this sulphated polysaccharide can inhibit proliferation and induce apoptotic cell death of human lung carcinoma cells,¹⁴⁴ human breast cancer cells by the activation of caspase 8.¹⁴⁵ Enzyme digested fucoidan extracts suppress the expression and secretion of various angiogenesis factors thereby produce inhibitory effect on angiogenesis of tumor cells.¹⁴⁶ Furthermore immunomodulating activity of fucoidan empowers it to act as a potential mitogen for lymphocyte and macrophage activation.¹⁴⁷ Fucoidan can be used for the treatment of osteoarthritis. Oral administration of seaweed extract containing fucoidan inhibited the symptoms of osteoarthritis.¹⁴⁸ For biomedical applications fucoidan – chitosan micro complex has been produced as a carrier for controlled release of specialised growth factor and the findings suggested growth factor containing fucoidan-chitosan hydrogel can be used for the treatment of ischemic disease.¹⁴⁹ Another blended hydrogel consisting of chitosan, alginate and fucoidan has found successful application in healing-impaired wound dressing.¹⁵⁰ Low molecular weight fucoidan is in use for bone extracellular matrix formation in 3D culture.¹⁵¹

CONCLUSION

All the aforementioned evidences suggest that marine organisms have the capacity to generate a wide range of useful biomaterials. The major area of concern for the scientific community is the enormous diversity of marine ecosystems and exploitation of this feature for the generation of novel biomaterials. Already nature derived biomaterials have taken a giant leap beating the synthetic biomaterials in terms of its several beneficial properties. With the advent of marine derived biomaterials there will be a new horizon in the field of biomedical science. The first and foremost step should be proper isolation method. Marine environment is entirely different from

the terrestrial one having great differences in pressure, temperature etc. So, application of suitable isolation technique along with laboratory based chemical approach should be developed for research, development and commercialisation of the biomaterials. At the time of birth the earth had extreme environment which are still present in large amount in marine environment in the form of hydrothermal vent, hot springs etc. Also thermocline and halocline environment demand special mention in this context. These are all sources of unique microorganisms which can be investigated for novel bioactive compounds. Therefore in conclusion we can say that by the help of intense research and discovery, strategic approach should be taken for the exploration of one of the finest attractions of modern science.

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REFERENCES

- Williams DF, On the nature of biomaterials, *Biomaterials*, 30, 2009, 5897-5909.
- Aribo S, Natural products: A Minefield of biomaterials, *ISRN Materials Science*, 2012.
- Porter DA, Melissa D, Susan JFM, Fifth Metatarsal Jones Fracture Fixation With a 4.5-mm Cannulated Stainless Steel Screw in the Competitive and Recreational Athlete A Clinical and Radiographic Evaluation, *The American Journal of Sports Medicine*, 33, 2005, 726-733.
- Ryhänen JM, Kallioinen J, Tuukkanen P, Lehenkari J, Junila E, Niemelä P, Sandvik, W. Serlo, Bone modeling and cell-material interface responses induced by nickel-titanium shape memory alloy after periosteal implantation, *Biomaterials*, 20, 1999, 1309-1317.
- Knosp H, Richard JH, Christopher WC, Gold in dentistry: Alloys, uses and performance, *Gold Bulletin*, 36, 2003, 93-102.
- Okazaki Y, Emiko G, Metal release from stainless steel, Co-Cr-Mo-Ni-Fe and Ni-Ti alloys in vascular implants, *Corrosion Science*, 50, 2008, 3429-3438.
- Pleva J, Dental mercury - A Public health hazard, *Reviews on Environmental Health*, 10, 1994, 1-28.
- Benz EB, Micheline F, John JG, Benjamin EB, Thomas ST, Myron S, Transmission electron microscopy of intracellular particles of polyethylene from joint replacement prostheses: size distribution and cellular response, *Biomaterials*, 22, 2001, 2835-2842.
- Chandy T, Gladwin SD, Robert FW, Gundu HRR, Use of plasma glow for surface-engineering biomolecules to enhance blood compatibility of Dacron and PTFE vascular prosthesis, *Biomaterials*, 21, 2000, 699-712.
- Dobrin PB, Some mechanical properties of polypropylene sutures: relationship to the use of polypropylene in vascular surgery, *Surgical Research*, 45, 1988, 568-573.
- Padilla DJ, Omayra L, Henrik RI, Lucie G, Jean MJF, Francis CS, Polyester dendritic systems for drug delivery applications: in vitro and in vivo evaluation, *Bioconjugate Chemistry*, 13, 2002, 453-461.
- Maas C, Robert S, Soft tissue augmentation apparatus, U.S. Patent No. 5,607,477, 4 Mar, 1997.



13. Middleton MS, Magnetic resonance evaluation of breast implants and soft-tissue silicone, *Topics in Magnetic Resonance Imaging*, 9, 1998, 92.
14. Lai JY, Biocompatibility of chemically cross-linked gelatine hydrogels for ophthalmic use, *Journal of Materials Science: Materials in Medicine*, 21, 2010, 1899-1911.
15. Katti KS, Biomaterials in total joint replacement, *Colloids and Surfaces B: Bio interfaces*, 39, 2004, 133-142.
16. Kawahara H, Masaya H, Takuji S, Single crystal alumina for dental implants and bone screws, *Journal of Biomedical Materials Research*, 14, 1980, 597-605.
17. Tien YC, Chih TT, Lin JHC, Ju CP, Lin SD, Augmentation of tendon-bone healing by the use of calcium-phosphate cement, *Journal of Bone & Joint Surgery*, 86, 2004, 1072-1076.
18. Ferracane JL, Current trends in dental composites, *Critical Reviews in Oral Biology & Medicine*, 6, 1995, 302-318.
19. FratzlIP, Biomimetic materials research: what can we really learn from nature's structural materials, *Royal Society Interface*, 4, 2007, 637-642.
20. Gray JS, Marine biodiversity: patterns, threats and conservation needs, *Biodiversity & Conservation*, 6, 1997, 153-175.
21. Snelgrove PVR, Getting to the bottom of marine biodiversity: Sedimentary habitats: Ocean bottoms are the most widespread habitat on earth and support high biodiversity and key ecosystem services, *BioScience*, 49, 1999, 129-138.
22. Ward RD, Woodwark M, Skibinski DOF, A comparison of genetic diversity levels in marine, freshwater, and anadromous fishes, *Journal of Fish Biology*, 44, 1994, 213-232.
23. Elliott NG, Ward RD, Enzyme variation in orange roughy, *Hoplostethus atlanticus* (Teleostei: Trachichthyidae), from southern Australian and New Zealand waters, *Marine and Freshwater Research*, 43, 1992, 1561-1571.
24. May RM, Bottoms up for the oceans, *Nature*, 357, 1992, 278-279.
25. Grassle JF, Nancy JM, Deep-sea species richness: regional and local diversity estimates from quantitative bottom samples, *American Naturalist*, 1992, 313-341.
26. Belarbi EH, Gómez AC, Chisti Y, Camacho FG, Grima EM, Producing drugs from marine sponges, *Biotechnology Advances*, 21, 2003, 585-598.
27. Ehrlich H, Maldonado M, Spindler KD, Eckert C, Hanke T, Born R, Goebel C, Simon P, Heinemann S, Worch H, First evidence of chitin as a component of the skeletal fibers of marine sponges. Part I. Verongidae (Demospongia: Porifera), *Journal of Experimental Zoology Part B: Molecular and Developmental Evolution*, 308, 2007, 347-356.
28. Müller WEG, Wang X, Cui FZ, Jochum KP, Tremel W, Bill J, Schröder HC, Natalio F, Schloßmacher U, Wiens M, Sponge spicules as blueprints for the biofabrication of inorganic-organic composites and biomaterials, *Applied Microbiology and Biotechnology*, 83, 2009, 397-413.
29. Turk T, Sepčić, K, Mancini I, Guella G, 3-Akylpyridinium and 3-alkylpyridine compounds from marine sponges, their synthesis, biological activities and potential use. *Studies In Natural Products Chemistry*, 35, 2008, 355-397.
30. Richelle-Maurer E, Gomez R, Braekman JC, Van de Vyver G, Van Soest RW, Devijver C, Primary cultures from the marine sponge *Xestospongia muta* (Petrosiidae, Haplosclerida), *Journal of Biotechnology*, 100, 2003, 169-176.
31. Monks NR, Lerner C, Henriques AT, Farias FM, Schapoval EES, Suyenaga ES, Rocha AB, Schwartzmann G, Mothes B, Anticancer, antichemotactic and antimicrobial activities of marine sponges collected off the coast of Santa Catarina, southern Brazil, *Journal of Experimental Marine Biology and Ecology*, 281, 2002, 1-12.
32. Armstrong E, Douglas McKenzie J, Goldsworthy GT, Aquaculture of sponges on scallops for natural products research and antifouling, *Progress in Industrial Microbiology*, 35, 1999, 163-174.
33. Schmidt EW, Donia MS, Life in cellulose houses: symbiotic bacterial biosynthesis of ascidian drugs and drug leads, *Current Opinion in Biotechnology*, 21, 2010, 827-833.
34. Aizenberg J, Lambert G, Weiner S, Addadi L, Factors involved in the formation of amorphous and crystalline calcium carbonate: a study of an ascidian skeleton, *Journal of the American Chemical Society*, 124, 2002, 32-39.
35. Karim S, Pereira J, Gueniche F, Delbarre-Ladrat C, Sinquin C, Ratskol J, Godeau G, Fischer AM, Helley D, Collic-Jouault S, Marine polysaccharides: A source of bioactive molecules for cell therapy and tissue engineering, *Marine Drugs*, 9, 2011, 1664-1681.
36. Cruz LJ, Luque-Ortega JR, Rivas L, Albericio F, Kahalalide F, an antitumor depsipeptide in clinical trials, and its analogues as effective antileishmanial agents, *Molecular Pharmaceutics*, 6, 2009, 813-824.
37. Lindquist N, Fenical W, Van Duyne GD, Clardy J, New alkaloids of the lamellarin class from the marine ascidian *Didemnum chartaceum* (Sluiter, 1909), *The Journal of Organic Chemistry*, 53, 1988, 4570-4574.
38. Appleton DR, Page MJ, Lambert G, Berridge MV, Copp BR, Kottamides AD: Novel Bioactive Imidazolone-Containing Alkaloids from the New Zealand Ascidian *Pycnoclavellakottae*, *The Journal of Organic Chemistry*, 67, 2002, 5402-5404.
39. Salomon CE, Faulkner DJ, Localization Studies of Bioactive Cyclic Peptides in the Ascidian *Lissoclinum patella*, *Natural Products*, 65, 2002, 689-692.
40. Ehrlich H, Simon P, Carrillo-Cabrera W, Bazhenov VV, Botting JP, Ilan M, Ereskovsky AV, Muricy G, Worch H, Mensch A, Born R, Springer A, Kummer K, Vyalikh DV, Molodtsov SL, Kurek D, Kammer M, Paasch S, Brunner E, Insights into chemistry of biological materials: newly discovered silica-aragonite-chitin bio composites in demosponges, *Chemistry of Materials*, 22, 2010, 1462-1471.
41. Pallela R, Bojja S, Janapala VR, Biochemical and biophysical characterization of collagens of marine sponge, *Ircinia fusca* (Porifera: Demospongiae: Irciniidae), *International Journal of Biological Macromolecules*, 49, 2011, 85-92.
42. Andrade P, Willoughby R, Pomponi SA, Kerr RG, Biosynthetic studies of the alkaloid, stevensine, in a cell culture of the marine sponge *Teichaxinellamorchella*, *Tetrahedron Letters*, 40, 1999, 4775-4778.
43. Joseph B, Nair VM, Sujatha S, Pharmacological perception of peptides from marine sponge: A Review, *International Pharmaceutical Science & Research*, 3, 2012, 4689-4696.
44. Dutta PK, Dutta J, Tripathi VS, Chitin and chitosan: Chemistry, properties and applications, *Science and Industrial Research*, 63, 2004, 20-31.
45. Horisberger M, Vonlanthen M, Location of mannan and chitin on thin sections of budding yeasts with gold markers, *Archives of Microbiology*, 115, 1977, 1-7.
46. Peniche C, Argüelles-Monal W, Goycoolea FM, Chitin and chitosan: major sources, properties and applications, *Monomers, Polymers and Composites from Renewable Resources*, 2008, 517.
47. Ravi Kumar MNV, A review of chitin and chitosan applications, *Reactive and Functional Polymers*, 46, 2000, 1-27.
48. Kurita K, Chitin and chitosan: functional biopolymers from marine crustaceans, *Marine Biotechnology*, 8, 2006, 203-226.



49. Rinaudo M, Chitin and chitosan: properties and applications, *Progress in Polymer Science*, 31, 2006, 603-632.
50. Varadharajan D, Ramesh S, Antibacterial activity of commercially important aquaculture candidate shrimp chitin extracts against estuarine and marine pathogens from Parangipettai coast, south east coast of India, *Journal of Microbiology & Biotechnology Research*, 2, 2012, 632-640.
51. Kamino K, Inoue K, Maruyama T, Takamatsu N, Harayama S, Shizuri Y, Barnacle cement proteins importance of disulfide bonds in their insolubility, *Journal of Biological Chemistry*, 275, 2000, 27360-27365.
52. Waite JH, Evidence for a repeating 3, 4-dihydroxyphenylalanine- and hydroxyproline-containing decapeptide in the adhesive protein of the mussel, *Mytilusedulis L*, *Journal of Biological Chemistry*, 258, 1983, 2911-2915.
53. Burkett JR, Wojtas, JL, Cloud JL, Wilker JJ, A method for measuring the adhesion strength of marine mussels, *The Journal of Adhesion*, 85, 2009, 601-615.
54. Deming TJ, Mussel byssus and biomolecular materials, *Current Opinion in Chemical Biology*, 3, 1999, 100-105.
55. Yu M, Hwang J, Deming TJ, Role of L-3, 4-dihydroxyphenylalanine in mussel adhesive proteins, *Journal of the American Chemical Society*, 121, 1999, 5825-5826.
56. Grande DA, Pitman MI, The use of adhesives in chondrocyte transplantation surgery, Preliminary studies, *Bulletin of the Hospital for Joint Diseases Orthopaedic Institute*, 48, 1987, 140-148.
57. Ben-Nissan B, Natural bio ceramics: from coral to bone and beyond, *Current Opinion in Solid State and Materials Science*, 7, 2003, 283-288.
58. Guillemain G, Patat, JL, Fournie J, Chetail M, The use of coral as a bone graft substitute, *Journal of Biomedical Materials Research*, 21, 1987, 557-567.
59. Vago R, Plotquin D, Bunin A, Sinelnikov I, Atar D, Itzhak D, Hard tissue remodeling using biofabricated coralline biomaterials, *Journal of Biochemical and Biophysical Methods*, 50, 2002, 253-259.
60. Ripamonti U, Crooks, J, Khoali L, Roden L, The induction of bone formation by coral-derived calcium carbonate/hydroxyapatite constructs, *Biomaterials*, 30, 2009, 1428-1439.
61. Lahaye M, Rochas C, Chemical structure and physico-chemical properties of agar, *International Workshop on Gelidium*, Springer Netherlands, 1991.
62. Guerrero P, Etxabide A, Leceta I, Peñalba M, de la Caba K, Extraction of agar from *Gelidium sesquipedale* (Rhodophyta) and surface characterization of agar based films, *Carbohydrate Polymers*, 99, 2014, 491-498.
63. Li H, Yu X, Jin Y, Zhang W, Liu Y, Development of an eco-friendly agar extraction technique from the red seaweed *Gracilaria lemaneiformis* is *Bioresource Technology*, 99, 2008, 3301-3305.
64. Duckworth M, Yaphe W, The structure of agar: Part I. Fractionation of a complex mixture of polysaccharides, *Carbohydrate Research*, 16, 1971, 189-197.
65. McHugh DJ, Worldwide distribution of commercial resources of seaweeds including *Gelidium*, *International workshop on Gelidium*, Springer Netherlands, 1991.
66. Rajendran N, Puppala S, Raj MS, Angeeleena BR, Rajam C, Seaweeds can be a new source for bio plastics, *Journal of Pharmacy Research*, 5, 2012, 1476-1479.
67. Jiao G, Yu G, Zhang J, Ewart HS, Chemical structures and bioactivities of sulphated polysaccharides from marine algae, *Marine Drugs*, 9, 2011, 196-223.
68. Mourão PAS, Mariana SP, Searching for alternatives to heparin: sulphated fucans from marine invertebrates, *Trends in Cardiovascular Medicine*, 9, 1999, 225-232.
69. Felício-Fernandes G, Mauro L, Calcium phosphate biomaterials from marine algae, *Hydrothermal synthesis and characterisation*, *Química Nova*, 23, 2000, 441-446.
70. Ventura M, Canchaya C, Tauch A, Chandra G, Fitzgerald GF, Chater KF, van Sinderen D, Genomics of Actinobacteria: tracing the evolutionary history of an ancient phylum, *Microbiology and Molecular Biology Reviews*, 71, 2007, 495-548.
71. Ghai R, Rodríguez-Valera F, McMahon KD, Toyama D, Rinke R, de Oliveira TCS, Garcia JW, de Miranda FP, Henrique-Silva F, Metagenomics of the water column in the pristine upper course of the Amazon river, *PLoS one*, 6, 2011, e23785.
72. Stach EM, Alan TB, Estimating and comparing the diversity of marine actinobacteria, *Antonie van Leeuwenhoek*, 87, 2005, 3-9.
73. Lazzarini A, Cavaletti L, Toppo G, Marinelli F, Rare genera of actinomycetes as potential producers of new antibiotics, *Antonie van Leeuwenhoek*, 78, 2000, 399-405.
74. Gao B, Gupta RS, Phylogenetic framework and molecular signatures for the main clades of the phylum Actinobacteria, *Microbiology and Molecular Biology Reviews*, 76, 2012, 66-112.
75. Lam, KS, Discovery of novel metabolites from marine actinomycetes, *Current Opinion in Microbial*, 9, 2006, 245-251.
76. Gunde-Cimerman N, Zalar P, Hoog S, Plemenitaš A, Hypersaline waters in salterns–natural ecological niches for halophilic black yeasts, *FEMS Microbiology Ecology*, 32, 2000, 235-240.
77. De Hoog GS, Guého E, Masclaux F, Gerrits van den Ende AHG, Kwon-Chung KJ, McGinnis MR, Nutritional physiology and taxonomy of human-pathogenic *Cladosporium-Xylohypha* species, *Medical Mycology*, 33, 1995, 339-347.
78. Diego L, Pérez P, Sommaruga R, del Carmen Diéguez M, Ferraro M, Brizzio S, Zagarese H, van Broock M, Constitutive and UV-inducible synthesis of photo protective compounds (carotenoids and mycosporines) by freshwater yeasts, *Photochemical & Photobiological Sciences*, 3, 2004, 281-286.
79. Ifuku S, Nogi M, Abe K, Yoshioka M, Morimoto M, Saimoto H, Yano H, Preparation of chitin nanofibers with a uniform width as α -chitin from crab shells, *Biomacromolecules*, 10, 2009, 1584-1588.
80. Boßelmann F, Romano P, Fabritius H, Raabe D, Epple M, The composition of the exoskeleton of two crustacea: The American lobster *Homarus americanus* and the edible crab *Cancer pagurus*, *Thermochimica Acta*, 463, 2007, 65-68.
81. Ciria LA, Huerta S, Hall GM, Shirai K, Pilot scale lactic acid fermentation of shrimp wastes for chitin recovery, *Process Biochemistry*, 37, 2002, 1359-1366.
82. Toan NV, Production of Chitin and Chitosan from Partially Autolyzed Shrimp Shell Materials, *The Open Biomaterials Journal*, 1, 2009.
83. Mekkes JR, Le Poole IC, Das PK, Bos JD, Westerhof W, Efficient debridement of necrotic wounds using proteolytic enzymes derived from Antarctic krill: a double-blind, placebo-controlled study in a standardized animal wound model. *Wound Repair and Regeneration*, 6, 1998, 50-57.
84. Nicol S, Graham WH, Chitin production by krill, *Biochemical Systematics and Ecology*, 21, 1993, 181-184.
85. Suzuki M, Sakuda S, Nagasawa H, Identification of chitin in the prismatic layer of the shell and a chitin synthase gene from the

- Japanese pearl oyster, *Pinctadafucata*, *Bioscience, Biotechnology and Biochemistry*, 71, 2007, 1735-1744.
86. Wu SC, Hsu HC, Wu YN, Ho WF, Hydroxyapatite synthesized from oyster shell powders by ball milling and heat treatment, *Materials Characterization*, 62, 2011, 1180-1187.
87. Zentz F, Bédouet L, Almeida MJ, Millet C, Lopez E, Giraud M, Characterization and quantification of chitosan extracted from nacre of the abalone *Haliotis tuberculata* and the oyster *Pinctada maxima*, *Marine Biotechnology*, 3, 2001, 36-44.
88. Lavall RL, Assis OB, Campana-Filho SP, β -Chitin from the pens of *Loligo* sp.: Extraction and characterization, *Bio resource Technology*, 98, 2007, 2465-2472.
89. Shepherd R, Reader S and Falshaw A, Chitosan functional properties, *Glycoconjugate Journal* 14, 1997, 535-542.
90. Wilker JJ, Marine bioinorganic materials: mussels pumping iron, *Current Opinion in Chemical Biology*, 14, 2010, 276-283.
91. Yule AB, Walker G, The adhesion of the barnacle, *Balanus balanoides*, to slate surfaces. *Journal of the Marine Biological Association of United Kingdom*, 64, 1984, 147-156.
92. Yang YJ, Choi YS, Jung D, Park BR, Hwang WB, Kim HW, Cha HJ, Production of a novel silk-like protein from sea anemone and fabrication of wet-spun and electro spun marine-derived silk fibers, *NPG Asia Materials*, 5, 2013, e50.
93. Reynolds WS, Schwarz JA, Weis VM, Symbiosis-enhanced gene expression in cnidarian-algal associations: cloning and characterization of a cDNA, sym32, encoding a possible cell adhesion protein, *Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology* 126, 2000, 33-44.
94. DeAngelis PL, Glabe CG, Polysaccharide structural features that are critical for the binding of sulphated fucans to bindin, the adhesive protein from sea urchin sperm, *Journal of Biological Chemistry*, 262, 1987, 13946-13952.
95. Stupp SI, Paul VB, Molecular manipulation of microstructures: biomaterials, ceramics, and semiconductors, *Science*, 277, 1997, 1242-1248.
96. Wilt FH, Bio mineralization of the spicules of sea urchin embryos, *Zoological Science*, 19, 2002, 253-261.
97. Hennebert E, Wattiez R, Flammang P, Characterisation of the carbohydrate fraction of the temporary adhesive secreted by the tube feet of the sea star *Asterias rubens*, *Marine Biotechnology*, 13, 2011, 484-495.
98. Lee KJ, Park HY, Kim YK, Park JI, Yoon HD, Biochemical characterization of collagen from the starfish *Asterias amurensis*. *Journal of the Korean Society for Applied Biological Chemistry*, 52, 2009, 221-226.
99. Lee SJ, Choi HS, Lee MH, Highly Sinterable Calcium Phosphate Fabricated by Using Starfish Bone. *Journal of Nanoscience and Nanotechnology*, 11, 2011, 1815-1817.
100. Ogawa Y, Kobayashi K, Kimura S, Nishiyama Y, Wada M, Kuga S, X-ray texture analysis indicates downward spinning of chitin micro fibrils in tubeworm tube. *Journal of Structural Biology*, 184, 2013, 212-216.
101. Zhao H, Sun C, Stewart RJ, Waite JH, Cement proteins of the tube-building polychaete *Phragmatopoma californica*, *Journal of Biological Chemistry*, 280, 2005, 42938-42944.
102. Ehrlich H, Etnoyer P, Litvinov SD, Olennikova MM, Domaschke H, Hanke T, Born R, Meissner H, Worch H, Biomaterial structure in deep-sea bamboo coral (Anthozoa: Gorgonacea: Isididae): perspectives for the development of bone implants and templates for tissue engineering, *Material wissenschaft und Werkstofftechnik*, 37, 2006, 552-557.
103. Born R, Ehrlich H, Bazhenov V, Shapkin NP, Investigation of nanoorganized biomaterials of marine origin, *Arabian Journal of Chemistry*, 3, 2010, 27-32.
104. Huang YF, Tian L, Fu HW, Hua HM, Pei YH, One new anthraquinone from marine *Streptomyces* sp. FX-58, *Natural product research*, 20, 2006, 1207-1210.
105. Hughes CC, MacMillan JB., Gaudêncio SP, Jensen PR, Fenical W, The Ammosamides: Structures of Cell Cycle Modulators from a Marine-Derived *Streptomyces* Species. *Angewandte Chemie International Edition*, 48, 2009, 725-727.
106. Bugni TS, Woolery M, Kauffman CA, Jensen PR, Fenical W, Bohemamines from a marine-derived *Streptomyces* sp, *Journal of Natural Products*, 69, 2006, 1626-1628.
107. Liu R, Zhu T, Li D, Gu J, Xia W, Fang Y, Liu H, Zhu W, Gu Q, Two indolocarbazole alkaloids with apoptosis activity from a marine-derived actinomycete Z2039-2, *Archives of Pharmacal Research*, 30, 2007, 270-274.
108. Kawasaki T, Kuzuyama TOMOHISA, Furihata K, Itoh N, Seto H, Dairi T, A relationship between the mevalonate pathway and isoprenoid production in actinomycetes, *The Journal of Antibiotics*, 56, 2003, 957.
109. Han XX, Cui CB, Gu QQ, Zhu WM, Liu HB, Gu JY, Osada H, ZHD-0501, a novel naturally occurring staurosporine analog from *Actinomadura* sp. 007, *Tetrahedron Letters*, 46, 2005, 6137-6140.
110. Lombó F, Velasco A, Castro A, De la Calle F, Braña AF, Sánchez-Puelles JM, Méndez C, Salas JA, Deciphering the biosynthesis pathway of the antitumor thiocoraline from a marine actinomycete and its expression in two *Streptomyces* species, *ChemBioChem*, 7, 2006, 366-376.
111. Jork A, Thürmer F, Cramer H, Zimmermann G, Gessner P, Hämel K, Hofmann G, Kuttler B, Hahn HJ, Josimovic-Alasevic O, Fritsch KG, Zimmerman U, Biocompatible alginate from freshly collected *Laminaria pallida* for implantation, *Applied Microbiology and Biotechnology*, 53, 2000, 224-229.
112. Petrovic U, Gunde-Cimerman N, Plemenitas A, Cellular responses to environmental salinity in the halophilic black yeast *Hortaea werneckii*, *Molecular Microbiology*, 45, 2002, 665-672.
113. Laine J, Labady M, Albornoz A, Yunes S, Porosities and pore sizes in coralline calcium carbonate, *Materials Characterization*, 59, 2008, 1522-1525.
114. Braye F, Irigaray JL, Oudadesse H, Weber G, Deschamps N, Deschamps C, Frayssinet P, Tourenne P, Tixier H, Terver S, Lefaivre J, Amirabadi A, Resorption kinetics of osseous substitute: natural coral and synthetic hydroxyapatite, *Biomaterials*, 17, 1996, 1345-1350.
115. Earl JS, Wood DJ, Milne SJ, Hydrothermal synthesis of hydroxy apatite, *Physics: Conference Series*, 26, IOP Publishing, 2006.
116. Liu DM, Troczynski T, Tseng WJ, Water-based sol-gel synthesis of hydroxy apatite: process development, *Biomaterials*, 22, 2001, 1721-1730.
117. Wang A, Liu D, Yin H, Wu H, Wada Y, Ren M, Jiang T, Cheng X, Xu Y, Size-controlled synthesis of hydroxy apatite nanorods by chemical precipitation in the presence of organic modifiers, *Materials Science and Engineering: C*, 27, 2007, 865-869.
118. Sun Y, Guo G, Tao D, Wang Z, Reverse microemulsion-directed synthesis of hydroxy apatite nanoparticles under hydrothermal conditions, *Journal of Physics and Chemistry of Solids*, 68, 2007, 373-377.
119. Tseng YH, Kuo CS, Li YY, Huang CP, Polymer-assisted synthesis of hydroxy apatite nanoparticle, *Materials Science and Engineering: C*, 29(3), 2009, 819-822.
120. Roy DM, Linnehan S, Hydroxy apatite formed from coral skeletal carbonate by hydrothermal exchange, *Nature*, 247, 1974, 220-222.

121. Kon E, Muraglia A, Corsi A, Bianco P, Maracci M, Martin I, Boyde A, Ruspantini I, Chistolini P, Rocca M, Giardino R, Cancedda R, Quarto R, Autologous bone marrow stromal cells loaded onto porous hydroxy apatite ceramic accelerate bone repair in critical-size defects of sheep long bones, *Biomedical Materials Research*, 49, 2000, 328-337.
122. Tschoppe P, Zandim DL, Martus P, Kielbassa AM, Enamel and dentine remineralisation by nano-hydroxy apatite toothpastes, *Journal of Dentistry*, 39, 2011, 430-437.
123. Dutton JJ, Coralline hydroxy apatite as an ocular implant, *Ophthalmology*, 98, 1991, 370-377.
124. Müller WEG, Wang X, Kropf K, Boreiko A, Schloßmacher U, Brandt D, Schröder HC, Wiens M, Silicatein expression in the hexactinellid *Crateromorphameyeri*: the lead marker gene restricted to siliceous sponges, *Cell and Tissue Research*, 333, 2008, 339-351.
125. Sarikaya M, Fong H, Sunderland N, Flinn BD, Mayer G, Mescher A, Gaiño E, Biomimetic model of a sponge-spicular optical fiber—mechanical properties and structure, *Journal of Materials Research*, 16, 2001, 1420-1428.
126. Hench LL, June W, Surface-active biomaterials, *Science*, 226, 1984, 630-636.
127. Sealy C, Nanoparticles target cancer cells in vivo, *Angew. Chem. Int. Ed.*, 45, 2006, 2238.
128. Wang X, Schröder HC, Wiens M, Ushijima H, Müller WE, Bio-silica and bio-polyphosphate: applications in biomedicine (bone formation), *Current Opinion in Biotechnology*, 23, 2012, 570-578.
129. Pietak AM, Reid JW, Stott MJ, Sayer M, Silicon substitution in the calcium phosphate bioceramics, *Biomaterials*, 28, 2007, 4023-4032.
130. Tønnesen HH, Karlsen J, Alginate in drug delivery systems, *Drug Development and Industrial Pharmacy*, 28, 2002, 621-630.
131. Colinet I, Dulong V, Mocanu G, Picton L, Le Cerf D, New amphiphilic and pH-sensitive hydrogel for controlled release of a model poorly water-soluble drug, *European Journal of Pharmaceutics and Biopharmaceutics*, 73, 2009, 345-350.
132. Lucinda-Silva RM, Salgado HRN, Evangelista RC, Alginate–chitosan systems: In vitro controlled release of triamcinolone and in vivo gastrointestinal transit, *Carbohydrate Polymers*, 81, 2010, 260-268.
133. Zhang X, Hui Z, Wan D, Huang H, Huang J, Yuan H, Yu J, Alginate microsphere filled with carbon nanotube as drug carrier, *International Journal of Biological Macromolecules*, 47, 2010, 389-395.
134. Christensen BE, Alginates as biomaterials in tissue engineering, *Carbohydrate Chemistry: Chemical and Biological Approaches*, 37, 2011, 227-258.
135. Balakrishnan B, Mohanty M, Umashankar PR, Jayakrishnan A, Evaluation of an in situ forming hydrogel wound dressing based on oxidized alginate and gelatin, *Biomaterials*, 26, 2005, 6335-6342.
136. d'Ayala GG, Malinconico M, Laurienzo P, Marine derived polysaccharides for biomedical applications: chemical modification approaches, *Molecules*, 13, 2008, 2069-2106.
137. No HK, Meyers SP, Application of chitosan for treatment of wastewaters, *Reviews of Environmental Contamination and Toxicology*, Springer New York, 2000, 1-27.
138. Dutta PK, Tripathi S, Mehrotra GK, Dutta J, Perspectives for chitosan based antimicrobial films in food applications, *Food Chemistry*, 114, 2009, 1173-1182.
139. Noel SP, Courtney H, Bumgardner JD, Haggard WO, Chitosan films: a potential local drug delivery system for antibiotics, *Clinical Orthopaedics and Related Research*, 466, 2008, 1377-1382.
140. Lai WF, Lin MCM, Nucleic acid delivery with chitosan and its derivatives, *Journal of Controlled Release*, 134, 2009, 158-168.
141. Liu C, Bai R, Adsorptive removal of copper ions with highly porous chitosan/cellulose acetate blend hollow fiber membranes, *Journal of Membrane Science*, 284, 2006, 313-322.
142. Upadrashta SM, Katikaneni PR, Nuessle NO, Chitosan as a tablet binder, *Drug development and Industrial Pharmacy*, 18, 1992, 1701-1708.
143. Venkatesan J, Kim SK, Chitosan composites for bone tissue engineering—An overview, *Marine Drugs*, 8, 2010, 2252-2266.
144. Boo HJ, Hyun JH, Kim SC, Kang JI, Kim MK, Kim SY, Cho H, Yoo ES, Kang HK, Fucoidan from *Undariapinnatifida* induces apoptosis in A549 human lung carcinoma cells, *Phytotherapy Research*, 25, 2011, 1082-1086.
145. Yamasaki-Miyamoto Y, Yamasaki M, Tachibana H, Yamada K, Fucoidan induces apoptosis through activation of caspase-8 on human breast cancer MCF-7 cells, *Journal of Agricultural and Food Chemistry*, 57, 2009, 8677-8682.
146. Ye J, Li Y, Teruya K, Katakura Y, Ichikawa A, Eto H, Hosoi M, Hosoi M, Nishimoto S, Shirahata S, Enzyme-digested fucoidan extracts derived from seaweed *Mozuku* of *Cladosiphon novae-caledoniae* inhibit invasion and angiogenesis of tumor cells, *Cytotechnology*, 47, 2005, 117-126.
147. Choi EM, Kim AJ, Kim YO, Hwang JK, Immuno modulating activity of arabinogalactan and fucoidan in vitro, *Journal of Medicinal Food*, 8, 2005, 446-453.
148. Irhimeh MR, Fitton JH, Lowenthal RM, Fucoidan ingestion increases the expression of CXCR4 on human CD34+ cells, *Experimental Hematology*, 35, 2007, 989-994.
149. Nakamura S, Nambu M, Ishizuka T, Hattori H, Kanatani Y, Takase B, Kishimoto S, Effect of controlled release of fibroblast growth factor-2 from chitosan/fucoidan micro complex-hydrogel on in vitro and in vivo vascularization, *Journal of Biomedical Materials Research Part A*, 85, 2008, 619-627.
150. Murakami K, Aoki H, Nakamura S, Nakamura SI, Takikawa M, Hanzawa M, Kishimoto S, Hydrogel blends of chitin/chitosan, fucoidan and alginate as healing-impaired wound dressings, *Biomaterials*, 31, 2010, 83-90.
151. Changotade SG, Korb J, Bassil B, Barroukh C, Willig SCJ, Patrick D, Godeau G, Senni K, Potential effects of a low-molecular-weight fucoidan extracted from brown algae on bone biomaterial osteoconductive properties, *Journal of Biomedical Materials Research Part A87*, 3, 2008, 666-675.

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