



Isolation and Characterisation of Phosphate Solubilising Bacteria from Different Soil

S.Anbuselvi*, L. Jeyanthi Rebecca, Guddu Kumar, Jitendra Kumar
 Department of Industrial Biotechnology, Bharath University, Chennai, Tamil Nadu, India.
 *Corresponding author's E-mail: anbuselvichennai@yahoo.com

Accepted on: 15-05-2015; Finalized on: 30-06-2015.

ABSTRACT

Soil microbes play a significant role in maintaining the nitrogen and phosphorus in soil. The phosphate content was found to be low in soil and get phosphates from rocks and decaying matter. In order to improve the phosphate in soil, is applying phosphor bacteria as inoculants to get sufficient amount of phosphorus in soil. Deficiency of phosphate leads to restrict the plant growth. The phosphate solubilising bacteria was isolated from different types of soil and the solubility characteristics of phosphate solubilising microbes were analyzed. Plants were grown with PSB Inoculants and morphological features were studied.

Keywords: PSB, phosphate, inoculants, biofertilizer

INTRODUCTION

Phosphorus is important for growth and maintenance of plants. It shows many physiological functions such as cell division, photosynthesis and development of root system¹. Plants absorb phosphate only in soluble form. The transfer of insoluble phosphate into soluble form is carried out by variety of microbes in the soil. Microbes can dissolve insoluble inorganic phosphates present in the soil and make them available to the plants².

Plants require phosphorus from the soil solution as phosphate anion. It increases the strength of plants, promotes flower formation and fruit production. It also increases their resistance to diseases and adverse conditions³. Phosphorus deficiency is found to be common in soil and phosphorus fertilizers are required to solve it and maintain the crop production. Only small portion is utilized by plants and remaining is converted into insoluble phosphate⁴.

Phosphorus can supply to the plants through biological ways. Phosphate solubilising microbes have been reported in conversion of insoluble phosphate to soluble primary and secondary orthophosphate ions⁵. This phosphate bio fertilizer helps to increase the accessibility of accumulated phosphates for plant growth^{6,7}. PSM improved phosphate uptake by plants and activated phosphorus in crop plants⁸.

PSM are best to be present more abundant in the rhizosphere of soil. One gram of fertile soil contains thousands of bacteria and Fungi. They can be divided into phosphate solubilising bacteria and phosphate solubilising fungi⁹. The solubilisation of phosphates takes place through processes or mechanisms of organic acid production and proton extrusion¹⁰.

Phosphate solubilising bacteria are found to be more common in fertile soil. Soil bacteria are in cocci, rod or

spiral shape. Bacillus and Pseudomonas secrete organic acids and lower the pH in their vicinity to bring about the dissolution of bound phosphate in soil. The present study was aimed at the isolation and characterisation of phosphate solubilising bacteria from different types of soil and its impact on the growth of plants.

MATERIALS AND METHODS

Soil samples were collected from different areas. Samples were air dried powdered and check the pH and NPK content of the soil.

10gm of soil sample was dissolved in 100ml distilled water sterilized water and mix the sample well and considered the diluted the soil sample in sterilized distilled water up to 10^{-7} dilution (each test tube containing 9ml of sterilized distilled water) then 10^{-5} 10^{-6} 10^{-7} dilutions taken for spread plate technique. Sterilized nutrient agar prepared and poured into petri dishes after solidification of the medium 0.1ml sample was poured into agar medium plate by using L-Rod spread the sample evenly over the agar surface and then incubated at 37°C for 24 hours.

Isolation of PSB

Pikovskaya's agar medium was found to be as selective media for the isolation of phosphate solubilising bacteria. The composition of pikovskaya medium was maintained in¹¹. The sterilized pikovskaya medium was prepared and poured into Petri dishes. After solidification of the medium, 0.1ml sample was poured into agar medium plate by using L-Rod spread the sample evenly over the agar surface and then incubated at 37°C for 24 hours³.

Detection of PSB

0.1 ml of PSB were isolated from each sample was subjected into pikovskaya agar medium containing insoluble tri calcium phosphate and incubated at 27-30°C for 7 days. Insoluble tri calcium phosphate is present in



the medium used for halo zone formation at 37°C in two weeks.

The morphological features of isolated bacteria via shape, size elevation, surface margins, surface texture and colour were observed¹² and characterized by gram staining.

Solubilisation Index

10 ml of each PSB culture was preserved in sterile distilled water. This was placed in pikovskaya agar medium at 28°C for seven days. Solubilisation index was measured by Edipremono¹³.

Quantitative analysis of phosphorus solubilisation by PSB

The determination of available phosphorus in sample was determined by phosphor molybdate method¹⁴.

Preparation of liquid inoculants

The pikovskaya 's broth incubated with water in 250ml conical flask . It was allowed to multiply by incubating at 32°C in a incubator cum shaker at 100 rpm for 72 hours. The broth containing approximately 25×10^{11} cfu/ml was used as a starter culture for the production of liquid inoculants. This can be used as nutrient enhancer for growth of plants.

RESULTS AND DISCUSSION

The isolation of phosphate solubilising bacteria from different types of soil were carried out and exhibited halozone formation (Figure 1).

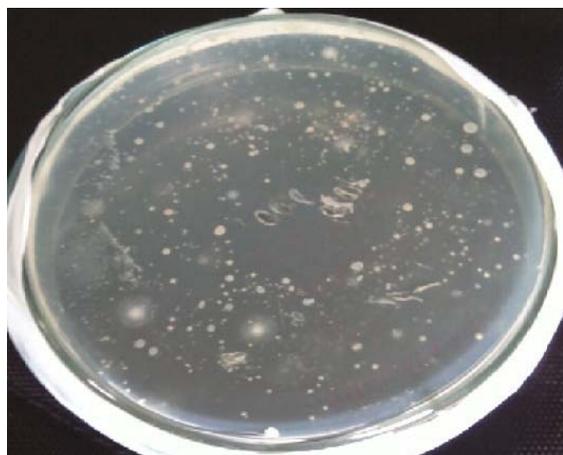


Figure 1: Clear zone formation from garden soil

It showed the capability of PSB secrete organic acids. The isolate showed colorless colonies which donot produce pigment. Cells were gram-negative, rod shaped and identified based on biochemical tests. The isolate were identified as *Bacillus*, *Bacillus substilis*, *Diphtheroids* and *arthobacteia* and *Actinomyces*.

High density of *bacillus* and *bacillus substitis* were found in fertile soil with the range of 2×10^7 and 3×10^6 cfu/g when compared with garden soil. *Bacillus* and *actinomyces* were observed in garbage soil. In non fertile

soil, *bacillus* was found to be very less in the concentration of 1×10^2 with 3 zone of clearance. The *Arthobacter* and *Diphtheroids* were also found (Table1). Six zone of clearance was found in *bacillus* of fertile soil. The maximum solubilisation of phosphate was observed in 5% of TCP (Table 2). Similar results showed that solubilisation index of *bacillus* ranged from 3.0 to 4.5¹⁵.

Table 1: Different kinds of phosphate solubilising bacteria in soil samples

S. No	Soil sample	Microbes	Colony Count (cfu/g)	Average zone of clearance
1	Fertile soil	<i>Bacillus</i>	2×10^7	10
		<i>Bacillus substilis</i>	3×10^6	9
2	Garden soil	<i>Bacillus</i>	4×10^6	6
		<i>Bacillus substilis</i>	3×10^5	5
3	Garbage soil	<i>Bacillus</i>	2×10^5	5
		<i>Actinomyces</i>	3×10^6	4
4	Non fertile soil	<i>Arthobacter</i>	2×10^4	3
		<i>Diphtheroids</i>	7×10^4	0

Table 2: Solubilisation zone of inhibition (mm)of different PSB

S. No	Organism	2.5% TCP	3%TCP	5%TCP	7%TCP
1	<i>Bacillus</i>	45	10.8	5.5	3.2
2	<i>Bacillus substilis</i>	80	10.6	10.2	2.5
3	<i>Actinomyces</i>	3.0	0.2	1.0	-

The amount of phosphate solubilisation activity f soil sample ranged from 16.3 µg to150 µg/ml. According to samiran, *Bacillus* TRSB16 showed high rate of solubilisation of calcium phosphate (144µg /ml) and *Arthobacter* showed low amount of phosphate solubilisation¹⁶.

The PSB based inoculants were used as nutrient enriched form of biofertilizer for the growth of green gram plant. The rapid growth was observed in inoculants treated plants than normal plants(Figure 2).



Figure 2: Morphological features of green gram plant (control and PSB treated)

CONCLUSION

The isolation and characterization of phosphate solubilising bacteria were identified in different range of

soil. The morphological features of PSB were analyzed based on degree of phosphate solubilisation and phosphorus uptake was measured.

Thus phosphate solubilising bacteria are predominantly in fertile and garden soil. This phospho bacteria can be used as biofertilizer for improving the yield of plants.

REFERENCES

1. Kannaiyan S, Kumar K and Govindarajan, Role of phosphate solubilising microbes in farming, Scientific pub, Jodhpur, 2004.
2. Yosef BB, Rogers RD, Wolfram JH and Richman, E.J. Soil. Sci of America, 1999, 1703-1708.
3. Bisen PS and Verma K, In Handbook of Microbiology, CBS Publishers and distributors.
4. Rodriguez C, Martinez-Molima E and E Velazquez, Effect of inoculation of a phosphate solubilising strain from *Pseudomonas jessenii* on growth of barley and chickpea plants, In proceedings of international congress of bacteriology and applied microbiology, 2002, 27.
5. Chabot RH, Antoun and Cescas MP, Stimulation of growth of maize and lettuce by inorganic phosphate solubilising microorganisms, Canadian. J. Micro.Biol, 39, 1993, 941-947.
6. Goldstein AH, Bacterial phosphate solubilisation, Historical perspective and future prospects, Am.J.Alt.Agri, 1, 1996, 57-65.
7. Gyaneshwar PG, Naresh kumar and LJ Parekh, Effect of buffering on the phosphate solubilising ability of microorganisms, World.J.Microbiol.Biotech, 14, 2002, 669-673.
8. Rodgers RD and JH Wolfram, Phosphorus sulphur and silicon related elements, 17, 1993, 1-4.
9. Guar AC, Phosphate solubilising microbes as biofertilizers, Omega scientific publishers, New Delhi, 1990, 76.
10. Nahas E, Factors determining rock phosphate solubilisation by microorganisms isolated from soil, World J.Microbiol.Biotechnol., 12, 1996, 18-23.
11. Pikovskaya RI, Mobilization of phosphorus in soil in connection with vital activity of some microbial species. Microbiologiya, 17, 1948, 362-370.
12. Lal L, In, Agritech.pub.Academy, Udaipur, 2002, 224.
13. Edi; Premona MA, Moawa and PLG Vleck, Effect of phosphate solubilising bacteria on the growth of maize and its survival in rhizosphere, Indonesian J of crop science, 11, 1996, 13-23.
14. Murphy JP, Riley, A modified single solution method for the determination of phosphate in natural waters, Analytica chimia acta, 27, 1962, 31-36.
15. Rashid MS, Khalil N, Ayush S Alam and F Latif, Organic acids production and phosphate solubilisation by PSM under *in vitro* conditions, Pak.J.Biolo.Sci, 7, 2004, 187-196.
16. Samiran B, Patil R, Sengupta C and D Standing, Stress induced phosphate solubilisation by *Arthobacter* and *Bacillus* from tomato rhizosphere, Australian.J. Crop science, 4(6), 2010, 376-383.

Source of Support: Nil, Conflict of Interest: None.