



A New Light of Therapy for Non-Alcoholic Fatty Liver Disease: Symbiotic.

Prasandeeep Biswal^{1*}, Abhishek Pal¹, Alok Prasad Das²

¹School of Pharmaceutical Sciences, Siksha O Anusandhan University, Bhubaneswar, India.

²Centre of Bio-technology, Siksha O Anusandhan University, Bhubaneswar, India.

*Corresponding author's E-mail: prasandeeep123@gmail.com

Accepted on: 10-06-2015; Finalized on: 30-06-2015.

ABSTRACT

Non-alcoholic fatty liver disease (NAFLD) is the most common chronic liver disease in the world. Oral administration of symbiotic has been proposed as an effective treatment of NAFLD because of its modulating effect on the gut flora, which can influence the gut-liver axis. The incidence of obesity and its related conditions, including non-alcoholic fatty liver disease (NAFLD) has been dramatically increased in all age groups worldwide. Given the health consequences of these conditions and the subsequent economic burden on healthcare systems, their prevention and treatment have become major priorities. Symbiotic supplementation in addition to lifestyle modification is superior to lifestyle modification alone for the treatment of NAFLD, at least partially through attenuation of inflammatory markers in the body. Recent evidence suggests that the gut microbiota may play a role in the development of insulin resistance, hepatic steatosis, necro-inflammation and fibrosis. Because standard dietary and lifestyle changes and pathogenically-oriented therapies (e.g., antioxidants, oral hypoglycaemic agents, and lipid-lowering agents) often fail due to poor compliance and lack of efficacy, novel approaches directed toward a safe and effectively alter patho-mechanism is in way of study. The immune-regulatory effects of probiotics may be beneficial in NAFLD treatment as they modulate the intestinal microbiota, improve epithelial barrier function, strengthen the intestinal wall and decreases its permeability, reduce bacterial translocation and endotoxemia though improves intestinal inflammation, reduce oxidative and inflammatory liver damage.

Keywords: NAFLD, Probiotics, prebiotics, Symbiotics

INTRODUCTION

Symbiotic are the combinational outcome of prebiotics and probiotics that benefits the host by improving the live micro-organisms survivability and growth stimulating activity in gastro-intestinal tract. Due to perceived health boosting aid of both probiotic and prebiotic they can be administered in combination or as separate units in the form of microcapsule.¹

Therefore, symbiotic unveils several benefits which include reduction in serum cholesterol, anti-carcinogenic activity, immune-stimulating activity, antimicrobial activity and inflammatory liver damage. A true health beneficial effect has been proven by abundant incorporation of prebiotics into probiotic. Likewise prebiotic serves as a food for probiotics which modulates the gut flora.

In addition *Lactobacillus acidophilus*, *Bifidobacterium longum* and *Lactobacillus casei* consuming foods predominantly showing significant health benefits after adding prebiotics. Thus these fallouts have been amplifying the concept of symbiotic.²

Probiotics are living microorganisms which boosts the immune-system and shows beneficial effects for human health by producing antibodies. These are called as "Good bacteria" present inside the body that can protect the gut from harmful bacteria and inhibits their growth. Thus diarrhoea, colon cancer, vaginal and urogenital infection, high serum cholesterol and resistance to infectious disease happen due to those harmful bacterial growths in

gut. Hence probiotics are helpful to overcome these infectious conditions. The most common probiotic bacteria are belong to lactobacillus and bifido-bacteria groups and this has been shown proficient benefits against bacterial infection.

Now a days the heightening interest in low fat products encouraging for probiotic product development and there are different approaches has been studied for enhancement, survivability of probiotic bacteria in acidic and enteric medium including physical defence granted by food system.^{3,4}

Prebiotics are basically non-digestible food ingredients occur from natural sources that encourages the growth of healthy bacteria in gut but In reality the term prebiotics was found by replacing 'pro' into 'pre' which means 'before' or 'for'.^{5,6}

In recent years, Inulin and oligosaccharides are the most demonstrated prebiotics. The concept of development of prebiotics from different sources has been in demand because of arrival of various functional foods in health conscious consumers.⁷

Ban on the use of antibiotics and hormones as feed additives, consumer awareness, and strict quality control actions are the driving factors for intense research and development in the areas of functional food, especially the prebiotic oligosaccharides.⁸

The aim of this review is to study on prebiotics, probiotics, symbiotic and symbiotic formulations by



following different microencapsulation techniques which is a great link to cure the non-alcoholic fatty liver disease.

Role of Gut Microbiota in Liver

In general gut is an alimentary tube which not only involves in transfer of food to digestive organs but also contains gut flora that termed as the largest reservoir of live micro-organism and consist of various species of micro-organism. *E.coli* is one of the common species of bacteria that mainly found in the human gut.

Therefore gut micro-organism shows most of useful effects by assembling the energy from the fermentation of undigested carbohydrate and by absorption of (SCFAs) short-chain fatty acids. These are most essential fatty acids such as butyrate and propionates which mainly absorbed by colonic epithelium and another by the liver where butyrate is energy sources. But acetate found in peripheral tissues that mainly enter through systemic circulation which is also uses as a premier element for biosynthesis.^{9,10}

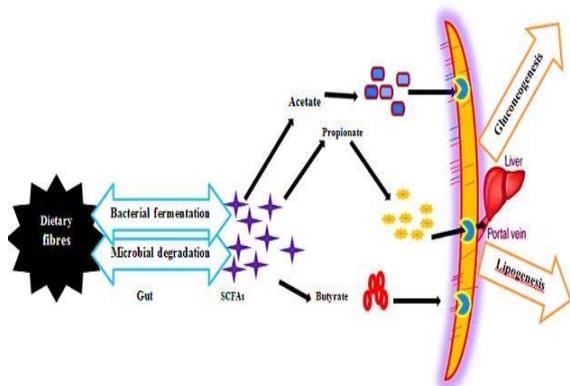


Figure 1: Schematic representation of Functional interaction between the gut microbiota and host metabolism¹⁰

Figure-1 explains that Acetate, propionate, and butyrate are the basic SCFAs mainly found in gut lumen of humans, but due to bacterial fermentation of carbohydrates inside the lumen that leads to production of propionate which mainly absorbed into the portal vein and rapidly removed from blood by the liver that could be a chief factor in the hepatic gluconeogenesis regulation.¹⁰

Liver glucose production and food intake regulates energy and glucose homeostasis thus, the upper duodenal lipids activates an intestine-brain-liver neural axis to control glucose homeostasis by inhibiting glucose production.

Liver is a prime part of body system for filtration and shows first line defence for the host simultaneously.¹¹

The research suggests that there is a dense link between gut micro biota and liver, this relation shows that helpful matters generated by the liver are absorbed by the gut that means liver accepts nearly about 70% of blood stream from intestine and signifies a defence contrary to antigens derived from gut. Therefore intestinal microbes

plays vibrant protagonist in preservations of gut-liver axis.¹²

Due to bare display towards gut, liver is a major spot for bacterial phagocytosis which involves in procurement of nutrients and leads to exclusion of pathogens therefore ‘Kupffer-cell’ so-called as permanent macrophages of liver regulates the passage of endotoxin and phagocyte bacteria through portal vein thus it may cause systemic bacterial infection. As a result K-cell is like an engine, once got activated, that will run the damage of liver in sharp speed therefore Gut liver axis shares a supreme part in pathogenesis of non-alcoholic fatty liver disease.¹³

NAFLD (Non-Alcoholic Fatty Liver Disease)

Non-alcoholic fatty liver disease (NAFLD) is a severe liver disease that the large proportion of a population found to have a condition of growing in incidence by means of the global epidemic of fattiness and resistance towards insulin for over the past few years and also both are accompanied with alteration in gastric micro biota.^{14,15}

On the other hand understanding of the previous studies was, how the micro biota subsidizes to the pathophysiology of non-alcoholic fatty liver disease is still a big task due to intestinal bacterial overgrowth which may leads to inflammation and liver injury.^{16,22}

Even though the diversity of reasons, behind the liver damage may be viral, toxic and metabolic, but pathogenic mechanisms are responsible for various kinds of liver injury from inflammation to liver cirrhosis. Thus changes in intestinal flora look like to show a key protagonist in stimulation and the liver damage progression therefore Probiotics may constructively powers several functions of the colonic micro biota and helps to make pathogenic adjustments in chronic liver disease.^{23,25}

Epidemiology of NAFLD

Table 1: The evaluation of degree of steatosis in percentage (%)^{29,30}

(%) of involvement of steatotic hepatocytes	Steatosis level
0% - 30%	Mild
33% - 66 %	Moderate
> 66 %	Severe

Both Non-alcoholic fatty liver and alcoholic liver have quite similar mechanism but NAFLD take place to people having no past of alcohol abuse. NAFLD mainly due to increase in influx of free fatty acids to liver that rises fat synthesis and indirectly helps in opening of inflammatory pathways.

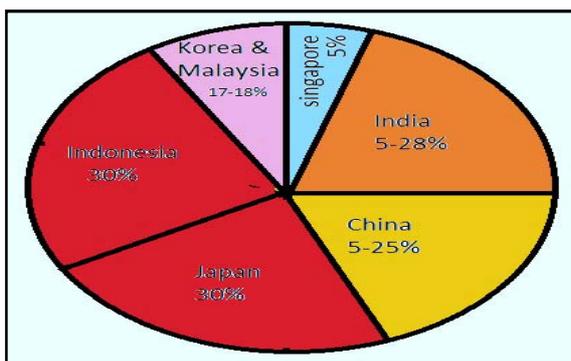
Therefore hepatocytes lipid deposition plays main anchor for creating fatty liver and that increases the consequence NAFLD world population.²⁶⁻²⁸

To diagnose Fatty Liver, there is a technique called Liver ultrasonography most commonly used. But a diagnosis of

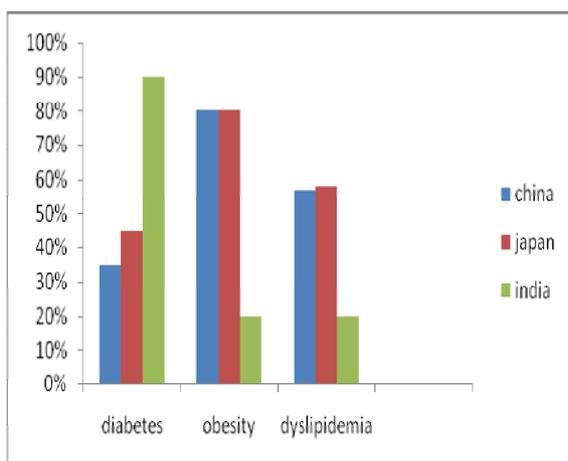
non-alcoholic fatty liver should be performed if steatosis reached more than 5% of hepatocyte that means more than 10% of liver is fatty and Table-1 below shows the steatosis level of mild, moderate and severe depending (%) of hepatocytes.²⁹⁻³¹

However, the prevalence of NAFLD differs depending on weight, gender, age, obesity and insulin resistance. The Study conveyed that US adults of 30% and 25% Italian adults are more exposed towards NAFLD Likewise, 80–90% in overweight adults, 30–50% diabetes also more open and importantly up to 90% with hyperlipidemia is mostly prone to NAFLD.³²⁻³⁵

In the year 2011 studied 400 patients (African American; Caucasians, Hispanic) of age range 28-70 from Brooked Army medium centre. As a result 46% found having NAFLD But among 54 diabetic patients, 74% Susceptible towards NAFLD whereas 22.2% shown NASH. But another study revealed that, 3 major countries of Asia like china, India, Japan shows apparent added prevalence of Non-Alcoholic Liver Disease up to 5 to 30% as part of others. Whereas in India high-risk citizenry like adipose 5-25%, dyslipidaemia 20% and (30-90%) diabetic are more open towards NAFLD shown in Graph-1 & 2.³⁶⁻⁴⁰



Graph 1: Incidence of non-alcoholic fatty liver disease within the adult citizens in Asian countries.^{38,39}



Graph 2: Incidence of non-alcoholic fatty liver disease between the high-threat citizenry (diabetes, obesity, dyslipidemia) in major Asian countries.³⁸⁻⁴⁰

Now the occurrence of having non-alcoholic fatty liver disease are growing globally and estimated to be 20-30%

in western countries like a current survey by National Health and Nutrition Examination (NHANE) predicted that about 30% prevalence of NAFLD in the United States between two conjugative year 2011 and 2012 where as in Asian countries the chance is within 15%.^{41,42}

One work from Taiwan had shown the prevalence of NASH in 80% obese patients when undergoing bariatric surgery. Now also children are going through diabetes therefore a research unveils that zone-3 steatosis and Diabetes are less common in children's than adults. But still it's a challenge for pathologist to diagnose NAFLD in children's.⁴³⁻⁴⁵

Therefore incidence of NAFLD between children is (3–10%), while it is mounting up to 40–70% with children having obesity. Moreover, pediatric NAFLD increased about 5% now with male-to-female ratio of 2:1.^{46,47}

Mainly the Clinico-pathological studies had been performed by employing a mathematical algorithm of mutilates by subsequent evaluation of 100 overweight children's and biopsy their liver resulted in type-1, type-2 diabetes, zone-3 steatosis and ballooning.^{48,49}

Pathogenesis of NAFLD

Non-Alcoholic Fatty Liver Disease is an ultimate communal pathological condition in western countries but now Asian countries are more dispose towards this disease due to two leading factor 1st is their rich-food intake and 2nd sedative lifestyle, that prompting in the direction of obesity and insulin resistance.⁵⁰

But previously experimental outcomes shown that obesity linked with NAFLD are relating to change in colonic penetrability and translocation of bacterial nutrients to liver.⁵¹

During obesity and insulin resistance (IR), there is rise in of free fatty acids (FFA) inflow and converts into triglycerides by beta oxidation or esterification with glycerol and stored in the form of lipid droplets that leads to increases fat deposition in liver.⁵²

Obesity is mainly due to gut micro flora variation and maximal intestinal permeability. Alteration in gut flora invokes release and overgrowth of bacterial endotoxin like lipo-polysaccharides and lipoprotein leads to endotoxemia which initiates liver damage. Then the next stage leads to activation of kupffer cell that indicates the liver damage by 1st hit i.e. by build-up fats in hepatocytes and the 2nd hit activates the (toll like receptors) TLRs along with CD-14 (clusters of differentiation -14) that releases lipo-polysaccharides abundantly and activates TNF- α (tumour necrosis factor- α) which gives a clue of liver injury (NAFLD), then the severity transforms into steatosis, NASH, fibrosis and cirrhosis.⁵³

But the key stage is steotisis which once has developed means that will sensitize the liver which leads to inflammation because of different stimuli. TNF- α intervened hepatocyte injury, Hyper-insulinemia,

oxidative stress, altered lipid partitioning and apoptosis all are due to second hit causing NASH.⁵⁴

TLRs (toll free receptors) are the key players in inflammation which identifies PAMPs (pathogen associated molecular patterns) such as lipopolysaccharides, lipoproteins to detect the presence of pathogen. The analysis betrayed that there are eleven Toll-like receptors (TLRs) have been identified in humans but TLRs 2, 4 and 9 are now the most importantly useful for recognition and signalling of Gram-positive and negative bacteria.⁵⁵ However TLRs Stimulation leads to the prompting of nuclear factor kappa B (NFκ-B) that results in antimicrobial gene with inflammatory cytokines and chemokine transcription.⁵⁶

But ethanol, ammonia and acetaldehyde production by the colonic micro flora are largely processed by liver and have capacity to control cytokine building and K- cell activity.⁵⁷

Histopathology

The histological field of NAFLD contains varied types of minor and ample atom macro-vesicular steatosis, which is mainly termed by means of inflammation with cell injury that leads to NASH and Evolution of fibrosis.⁵⁸

The capital histological feature of NAFLD is the accession of fat i.e. triglycerides accumulations in hepatocytes.^{59,60}

Masson's trichrome & Verhoeff stain of liver has shown a macro vesicular steatosis, balmy fibrosis and the hepatocytes however Macro-vesicular steatosis is basically due to the build-up of lipid that leads to alter the cell's nucleus.⁶¹

A previous report unveils that, Liver biopsy specimens of NAFLD patients are classified into four types. Type-1: steatosis alone, type 2: steatosis with lobular inflammation, type 3: steatosis with hepatocyte ballooning and type 4: combination of both type 3 and fibrosis which are new histological finding of NAFLD.⁶²

Selection of Prebiotic and Probiotic for Micro-Encapsulation

Prebiotics are usually belongs to hetero-groups, these are basically carbohydrates or polymeric carbohydrate molecules mainly isolated from dietary fibres, thus for microencapsulation the maximal food load of fibres should be a convincing step which helps to maintain a healthy environment in gut.

Prebiotics like fructo-oligosaccharides (FOS), Galactomannan, Short chain fatty acids, inulin, fructans, polysaccharides, mono-saccharides, and oligosaccharides are abundantly available in nature and also inexpensively isolated from plant roots, seeds, fruits, vegetables, marine herbs.

But the use of prebiotics for microencapsulation consists of its improvement of survivability, adherence and resistibility in GIT. Now most case studies had been

shown in improvisation of gut flora which is due to more use of symbiotic, which means use of prebiotics as substrate with probiotics. For example Inulin is the most widely accepted prebiotic from chicory roots because of its solubility, fermentability which leads to maximize bacterial count and modulates the gut flora.⁶³

Other prebiotics like glucans which is a polysaccharide of D-glucose monomer isolated from mushroom has shown potential activity by increasing in bacterial count in gut and provides an improvise immunity against pathogens and previously glucans are also used for diabetes and high cholesterol.⁶⁴⁻⁶⁶

Prebiotic action mainly depends on advance amount of bacillus growth and their count in large intestine because by metabolic agitator will not able to digest prebiotics like beta glucans, inulin, oligosaccharides, fructans but they are metabolized by beneficial bacilli known as (probiotics) and are existing in GIT that leads to access in bacterial advance and number.⁶⁷

Probiotics are mainly living cells which controls intestinal pathogen by modifying gut flora, constructing antibacterial activity in gut and up turning antibody level. Selection of probiotics for microencapsulation should include such parameters like survivability, adherence, anti-toxic activity and pathogen blocking activity which provides a shield against harmful bacteria.⁶⁸

Generally the main isolation source of probiotics are from milk, vegetables, fruits, yogurts, fishes which offers a vigorous health benefits by improvising the microbial balance. But in India 90% or more cultured products manufactured are from curd due to easy availability of milk like buffalo milk, cow milk, goat milk etc.⁶⁹

Lactic acid bacteria and bifido bacteria are most popular probiotic species which vastly isolated from numerous sources that offers better intestinal balance. According to JordiCun, lactic acid bacteria has shown auspicious properties when used as probiotics for improving oral health. But now bifido-bacteria are evolving groups of colonic bacteria helpful for human health. However Yogurts with bifido-bacteria or in combination with other lactobacillus species have been commonly useful for the fermented milks production.⁷⁰⁻⁷²

Symbiotic preparations are mainly for increasing endurance of probiotics in gastrointestinal tract which is a combination of prebiotics and probiotics. According to research fructo-oligosaccharides have shown more growth rate with these bacterial species *B. longum*, *B. animalis* Whereas in case of inulin growth rate little less as compared to FOS when treated in combination with same *bifido*-bacterial species.⁷³

Poor survivability of probiotics in GIT encourages a new technique called symbiotic which getting more popular day by day because of their highly usefulness in cholesterol lowering property and non-alcoholic fatty liver disease (NAFLD).⁷⁴



It has been recorded that patients with dissimilar stages of Non-alcoholic fatty liver disease i.e. from steatosis to cirrhosis are more open up towards endotoxemia and that leads increase in frequency of bacterial growth in small intestine and increase of intestinal permeability. Thus symbiotic could be a therapy to control and protect against intestinal alteration.^{75,76}

Microencapsulation: Purpose of this Technique

In recent years a fastest growing technology in pharmaceuticals called, microencapsulation is in report because of its demand on target drug delivery. In case probiotics, micro-encapsulation technique provides a barrier to these living cells throughout adverse environment until a drug reach the site of action.⁷⁶

Therefore this technology opens up a new working tool for pharmaceutical and biotechnological sector that helps to enhance their survivability and release profile but there are several factors that plays leading role in probiotics survivability, which includes PH, acidification, hydrogen peroxide production, oxygen toxicity, storage temperature, poor growth in GI system.^{77,78}

The main purpose of encapsulation is to provide not only environmental protection and controlling drug release characteristics but also to help in altering colloidal and surface properties, converting liquid into solid, reducing gastric irritation and to give stabilization to oxidation.⁷⁹

But practically this could be a process which involves by means of spreading a tinny coating layer to small solid particles or/ liquid droplets or/ dispersion. Where a hydrocolloid bead entraps the cells within bead matrix and reduces cell loss that helps probiotics to hit the target site. Likewise the bead matrix includes polymeric materials for example (chiton, xanthan, k-carageenan, cellulose acetate thalot, and alginate) for efficient target release in gut.⁸⁰

Lipids have shown beneficial approaches for encapsulation because of their lesser amount of solubility in water and fatty acids. Moreover lipids remain capable of holding several bioactive molecules during encapsulation for specific site delivery.⁸¹

Proteins, peptides, carbohydrates are called bioactive molecules which acts as a strategic growth factor likewise protein helpful in biodegradation in GIT due to its larger sensitivity and hetero groups like carbohydrates generally isolated from dietary fibres though helpful in cholesterol lowering.⁸²

Microencapsulation with maximum food load of fibres without compromising product quality is a convincing step forward which helps to reach high health benefits.

The uniqueness of microencapsulation is the smallness of the coated particle which indirectly depends on selection core material and coating material that leads to improvise the release properties. Therefore during microencapsulation, particle size adjustment always been

influenced by methods of encapsulation and core material.^{83,84}

Methods for Microencapsulation of Probiotics

Method of microencapsulation includes Air suspension, Multi-orifice centrifugal, Pan coating, Solvent evaporation, spray drying, emulsion method and extrusion method.⁸⁵

Extrusion Technique

Extrusion is the most useful and conjoint methodology to form capsules with the help of hydrocolloid beads. So, this is a most widely accepted technique due to its lesser cost, biocompatibility and formulation environment which offers maximal cell holding capacity and minimal cell damage.⁸⁶⁻⁸⁹

As a result, this methodology involves in preparing a hydrocolloid solution then adding microbes to form cell suspension, then take the suspension with the help of a syringe needle and allow the drops to fall into setting bath. The beads size and shape depend on the needle diameter and free-fall the distance, respectively.^{90,91}

The common material called alginate is mostly used for extrusion, which is a linear hetero-polysaccharide pull out from different algae species to formulate micro-beads. The size of the beads depends on viscosity of sodium alginate that means if viscosity raises the size of bead decreases.^{92,93}

To frame micro-beads all cell suspension is alloyed with a solution of sodium alginate and the admixture is dropped into a solution of calcium chloride (CaCl_2) to make gel spheres quickly by capturing the cells in a three-dimensionally by the lattice of alginate but for mass manufacturing of beads can either accomplished by nozzle system (figure-2) and centrifugal extraction process.

However Centrifugal extraction process is an aqueous extraction process, where the extruded rods separated into a droplet that forms capsules.^{93,94}

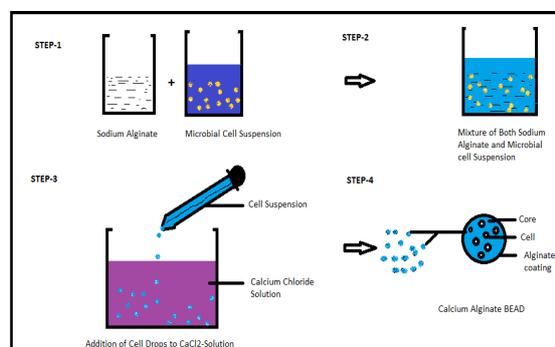


Figure 2: Schematic representation of extrusion methodology to frame micro-beads.^{90,91}

Emulsion Technique

This is a finer acclimated method and has been magnificently employed to encapsulate lactic acid

bacteria.⁹⁵ here, the addition of lesser bulk of the cell to polymer solution leads to formation of cell polymer suspension which emulsified by adding to an ample volume of a herbal oils (soybean oil, corn oil) then, by adding calcium chloride solution indicated that the formation of gel beads due to breakdown. Where, vegetable oils are continuous phase and cell polymer is alternative phase.

Most of the intellectual report gave an idea on the technique of encapsulation of probiotic-bacilli which have followed the method of emulsion to yield small bulk of capsules shell or beads. The beads or small bulk of capsules are made in a two-step action where first step involves in hardening and dispersion. Then followed by the homogenization procedure, admixture is to frame a water-in-oil emulsion. Once the water-in-oil emulsion made, the water soluble polymers cross-linked to formulate small gel units in between the oil phase (Fig. 3) however lesser the particle size of internal segment the emulsion, the lesser the concluding micro units will be.

The rapidity of agitation leads to controlling the size of the beads and can differ in the middle of 2 mm and 25 mm. In most of the cases shown that the addition emulsifiers leads to make an enhanced emulsion and this is due to lowering of the surface tension because of emulsifiers, hence leads to yield smaller spheres. Use of Tween 80 at 0.2% is most shared as emulsifier but use of Tween 80 and 0.5% sodium lauryl sulphate together helps to increase size of beads (i.e. produce a bead size of 25–35 mm)^{96,97}

K-carrageenan a natural polysaccharides from algae, cellulose acetate phthalate and many other supporting materials are used in the emulsion technique.⁹⁸

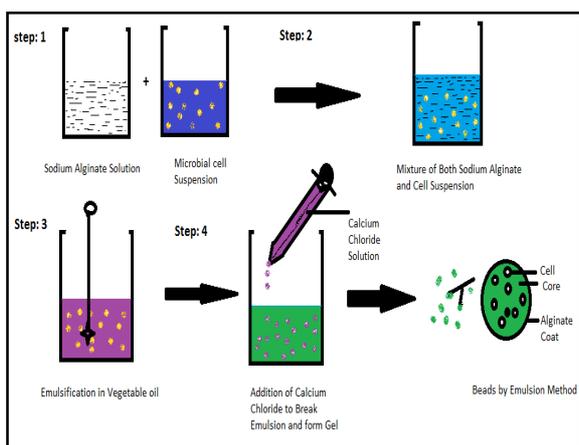


Figure 3: Schematic representation of emulsion methodology to frame micro-beads.^{96,97}

Spray Drying

Spray drying is the most popular microencapsulation technique because of rapid solidification of coating due to evaporation solvent in which coating material is dissolved (fig.-4). Even though spray drying of *Lactobacillus* cultures was first prepared in 1914 by Rogers, this thought was

not accepted due to very lesser rate of survivability and storage difficulty. So now it has most adopted method of food industry.^{99,100}

Microencapsulation by spray drying is showed by dispersing a core material in a coating solution, followed by atomization of mixture into air stream and the latent heat of vaporization mechanism helps to remove solvent from coating material thus construction of microcapsules as final product.¹⁰¹

This process produces spherical microcapsule size ranging from 5-600 micron and commonly employed in the microencapsulation of liquid flavours yielding dry and free flowing powders for industry due to its low bulk density and porous nature of coated particles.

This process might not be appropriate for encapsulating probiotic bacterial culture at high temperature like previous study shown probiotic bacteria count decreased by increasing inlet temperature but by adjusting inlet and outlet temperature viable encapsulated culture with desired particle size can be achievable.¹⁰²

However, according to previous study encapsulation at the rate of inlet temperature of 100°C with 45°C outlet temperature, bifido-bacterial cells were shown satisfactory results with gelatinised modified starch (as coating material).¹⁰³

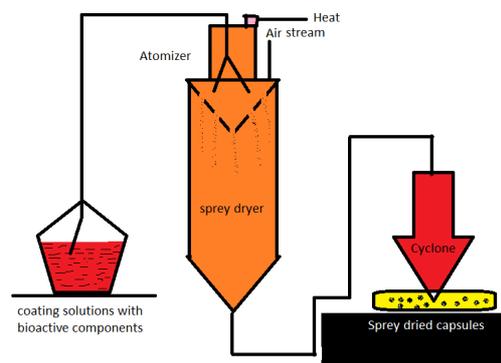


Figure 4: Schematic representation of spray microencapsulation technique.¹⁰¹

Air Suspension Technique

Air suspension is an altered technique resembles dispersion of solid where particles are suspended on upward moving air and this is due recirculation breeze mechanism.¹⁰⁴

This technique brings varied-ranging of coating candidate that gives a new aid for microencapsulation and also capable of smearing in the form of dispersion, emulsion and solvent solution.

However, firstly particles pass through the chamber of coating where polymer solution uses as a coating material and sprayed towards moving particle, then re-circulations mechanism helps to coat those particles in quick successions and product would dry by air stream. So, this procedure gives a proficient and effective

microencapsulation, even particle size of 37 micron can be effectively encapsulating by this method.¹⁰⁵

In the recent research evaluated that air-suspension fluidized-bed technique mostly useful for making of microcapsules core and shell having probiotic *Lactobacillus paracasei* cells as key ingredient. The procedure of air-suspension was accomplished in a Wurster coater system with help of a bottom-spraying atomizer where a solution containing probiotic cells with trehalose, maltodextrin was spray-coated and produces non-agglomerating dry coated particles with high probiotic cell viability (10^9 colony-forming units [cfu]/g particles) where microcrystalline cellulose an indolent carrier used as absorbent.¹⁰⁶

Casting of Materials and Coatings for Probiotics Microencapsulation

These are commonly used materials for microencapsulation of probiotics like Alginate, chitosan, starch, κ -Carrageenan, gelatin, milk, whey protein, inulin etc. Alginate is called a by itself acquired polysaccharide which isolated from several breed of algae and that mainly contains units of two monosaccharide such as α -L-guluronic acerbic and β -D-mannuronic acid where M/G ratios defines the alginate functionality and the gel backbone depends upon top admeasurement of block G. However for ensuring proper solubility water at High temperature (60°C to 80°C) is bare to dissolve alginate. Baffling concentration of alginate is common in acidic atmosphere so, the concentration range of 0.5–4% is mostly applicable.¹⁰⁷⁻¹¹²

A polysaccharide called Chitosan contains revoking charge which emerges from their amine groups acquired by deacetylation of chitin and further Chitosan can polymerize by means of polymer-polymer chain bonding accumulation in presence of anions and poly-anions. However insect cuticles and the membranes of fungi are the main isolation sources but Normally forms gel anatomy by ionic-tropic gelation and also soluble at pH <6. It is generally acclimated as coat/shell but not in the form of capsule and most of industries, this can be acclimated as self-healing polyurethane acrylic coating. In case of medicine, may be advantageous for terminating bleeding or/ as an antibacterial agent and also helpful in release drugs through the skin. Therefore chitosan could be the choice for probiotics encapsulation.¹¹³

Starch contains units of D-glucose which linked with glycosidic bonds. Most cases has been acclimated starch, High-amylose starch (HACS) and lyophilized starch (LCS) as an coating material for production of alginate capsules. However this has been reported that HACS and LCS decomposes when they exposed to pancreatic enzymes but resistant starch (RS) does not get decomposes at same exposure but it gives micro beads acceptable enteric supply appropriate as well gives them probiotic functionality so that they can be acclimated by the probiotic bacilli in the intestine. The assimilation of

Hi-Maize starch bacilli enhances sustainability of bacteria compared with the bacilli encapsulated without starch.¹¹⁴⁻¹¹⁸

Carrageenan is polysaccharides mainly isolated from seaweeds with anatomy consisting of D-galactose units containing α -(1–3) and β (1–4) bonds. This is commonly acceptable in industries due to their gelling properties. There are 3 different types of carrageenan such as kappa (κ), iota (ι), and lambda (λ) where κ carrageenan is the mono-sulfated but ι and λ are bi-sulfated and tri-sulfated. Carrageenan gelatin depends on change in temperature and that needs 60-80°C to make it properly dissolve. It got safe assurance by several government agencies including FDA, Codex-Alimentarius and the collective FAO/WHO food additive as well. Due to good acceptable gel anatomy that can allure the cell easily but the encapsulation of probiotic cell in κ -carrageenan chaplet keeps the bacilli in an applicable accompaniment however the formed gels are breakable and unable to bear pressures.¹¹⁹⁻¹²⁶

Other Coatings for Encapsulation: (Basically Proteins)

Gelatin is typically adapted by pharmaceutical industries because it is amphoteric in nature therefore this could be accomplished applicant to absorb with polysaccharides like gellan gums. Thermally reversible gelling cause in case of microencapsulation could be its tricky character. So, it is a protein acquired by partial hydrolysis of collagen and has highly viscous with water but forms gels at reduced temperature.¹²⁷

Milk proteins are familiarised carriers for probiotic cells but they can be acclimated as a drug delivery system due to biocompatibility with their physical and physicochemical assets. The after-effects of the readings are promising therefore using milk proteins as coating material would be an acceptable choice.¹²⁸

Whey protein seems to be good applicant as coating material because it is biodegradable and often get used by many nutriment products. The protein backgrounds receive altered cell release properties than fat based microencapsulation methods. Thus, Proteins-Based Coating in case of Encapsulation of Probiotics is a superior choice.¹²⁹

CONCLUSION

This review is based on types of methods, principles, use of different materials for encapsulating of probiotic cells and casting of materials for coatings of microencapsulated probiotics. The progresses in this area have been excellent with nutriment ingredients. However, microencapsulation of live bacteria or probiotics and the tools of symbiotic seem to be not established properly. The provision of probiotic bacteria and their improvement of viability in adverse environment will become vital incoming years. The challenges are to select the appropriate prebiotic sources, probiotic sources and encapsulation technique for



symbiotic. The chief assignment is to improvise the viability of probiotics in GIT.

REFERENCES

- Vivek KB, Use of encapsulated probiotics in dairy based foods, International Journal of Food, Agriculture and Veterinary Sciences, 3(1), 2013, 188-199.
- Gibson GR, Roberfroid MB, Dietary modulation of the human colonic microbiota: introducing the concept of prebiotics, Nutrition journal, 125, 1995, 1401-1412.
- Akalin S, Erisir D, Effects of inulin and oligofructose on the rheological characteristics and probiotic culture survival in low fat probiotic ice cream, Journal of Food Science, 73, 2008, 184-188.
- Flavia CAB, Castro IA, Saad SMI, Viability of *Lactobacillus acidophilus* in synbiotic guava mousses and its survival under *in vitro* simulated gastrointestinal conditions, International Journal of Food Microbiology, 137, 2010, 121-129.
- Schillinger U, Guigas C, and Heinrich W, *In vitro* adherence and other properties of lactobacilli used in probiotic yoghurt-like products, International Dairy Journal, 15, 2005, 1289-1297.
- Philippe RM, Michael DV, Christophe JC, Schrezenmeir J, Protection from gastrointestinal diseases with the use of probiotics, American Society for Clinical Nutrition, 73(2), 2001, 430s-436s.
- Aidaa FMNA, Shuhaimi MA, Yazid MB, Maaruf AGCA, Mushroom as a potential source of prebiotics, Trends in Food Science & Technology, 20, 2009, 567e-575.
- Samanta AK, Jayapal N, Senani S, Kolte AP, Sridhar M, Prebiotic inulin: Useful dietary adjuncts to manipulate the livestock gut microflora, Brazilian Journal of Microbiology, 44(1), 2013, 1-14.
- Cesaro C, Tiso A, Prete AD, Cariello R, Tuccillo C, Cotticelli G, Blanco CDV, Loguercio C, Gut microbiota and probiotics in chronic liver diseases, Digestive and Liver Disease journal, 43, 2011, 431-438.
- Tremaroli V and Backhed F, Functional interaction between the gut microbiota and host metabolism, nature, 489, 2012, 242-249.
- Wang PYT, Caspi L, Lam CKL, Chari M, Li X, Light PE, Juarez RG, Ang M, Schwartz GJ, Lam TKT, Upper intestinal lipids trigger a gut-brain-liver axis to regulate glucose production, Nature, 452, 2008, 1012-1016.
- Comparea D, Coccolia P, Roccoa A, Nardonea OM, Mariab SD, Cartenib M, Nardone G, Gut-liver axis: The impact of gut microbiota on non-alcoholic fatty liver disease, Nutrition, Metabolism and Cardiovascular Diseases, Department of Clinical and Experimental Medicine, Gastroenterology, 22(6), 2012, 471-476.
- Egmond MV, Garderen EV, Spriel ABV, Damen C A, Amersfoort ESV, Zandbergen GV, Hattum JV, Kuiper J, Winkel JGJVD Fc-RI-Positive liver Kupffer cells: Reappraisal of the function of immunoglobulin A in immunity, Nature Medicine, 6(6), 2000, 680-685.
- Wisniewsky AJ, Gaborit B, Dutour A, Clement K, Gut micro-biota and non-alcoholic fatty liver disease: new insights, European Society of Clinical Microbiology and Infectious Diseases, 19(4), 2013, 338-348.
- Large proportion of a population found to have a condition of growing Non-alcoholic fatty liver disease by means of the global epidemic of fattiness and resistance towards insulin, http://www.ucsfhealth.org/conditions/nonalcoholic_fatty_liver_disease/.
- Shanab A, Quigley EM, The role of gut microbiota in non-alcoholic fatty liver disease. Natures reviews, gastroenterol Hepatol, 7, 2010, 691-701.
- Henao MJ, Elinav E, Jin C, Hao L, Mehal WZ, Strowig T, Thaiss CA, Kau AL, Eisenbarth SC, Jurczak MJ, Camporez JP, Shulman GI, Gordon JI, Hoffman HM, Flavell RA, inflammation mediated dysbiosis regulates progression of NAFLD and obesity, Nature, 482, 2012, 179-185.
- Roy TL, Liopis M, Lepage P, Bruneau A, Rabot S, Bevilacqua C, Martin P, Philippe C, Walker F, Bado A, Perlemuter G, Doulier AMC, Philippe G, intestinal microbiota determines development of non-alcoholic fatty liver disease in mice, Gut, 2013, 62, 1787-1794.
- Greenblum S, Turnbaugh PJ, Borenstein E, Metagenomic systems biology of the human gut microbiome reveals topological shifts associated with obesity and inflammatory bowel disease, U S A, 109, 2012, 594-599.
- Dukowicz AC, Lacy BE, Levine GM. Small intestinal bacterial overgrowth: a comprehensive review, Gastroenterol Hepatol, 3, 2007, 112-122.
- Wood NJ, Microbiota: Dysbiosis driven by inflammasome deficiency exacerbates hepatic steatosis and governs rate of NAFLD progression, Natures Review Gastroenterol Hepatol, 9, 2012, 123.
- Solga SF, Diehl AM, Gut flora-based therapy in liver disease? The liver cares about the gut, Hepatology, 39, 2004, 1197-1200.
- Abt MC, Artis D, The intestinal microbiota in health and disease: the influence of microbial products on immune cell homeostasis. Current Opinion Gastroenterology, 25, 2009, 496-502.
- Vollaard EJ, Clasener HAL, Colonization resistance, Antimicrobial Agents and Chemotherapy, 38, 1994, 409-414.
- Loguercio C, Simone TD, Federico A, Terracciano F, Tuccillo C, Chicco MD, Carteni M, Gut-liver axis: a new point of attack to treat chronic liver damage?, The American Journal of Gastroenterology - Nature, 97, 2002, 2144-2146.
- Bellentani S, Saccoccio G, Masutti F, Croce LS, Brandi G, Sasso F, Cristanini G, Tiribelli C, Prevalence of and risk factors for hepatic steatosis in Northern Italy, Annals of Internal Medicine, 132, 2000, 112-117.
- Bellentani S, Bedogni G, Miglioli L, Tiribelli C, The epidemiology of fatty liver. European Journal of Gastroenterology & Hepatology, 16, 2004, 1087-1093.
- Matteoni CA, Younossi ZM, Gramlich T, Boparai N, Liu YC, McCullough AJ, Nonalcoholic fatty liver disease: a spectrum of clinical and pathological severity, Gastroenterology, 116(6), 1999, 1413-1419.
- Brunt EM, Tiniakos DG, Odze RD, Goldblum JR, Alcoholic and nonalcoholic fatty liver disease In Surgical Pathology of the GI Tract, Liver, Biliary Tract and Pancreas, 2nd edition, Philadelphia, 2009, 1007-1014.
- Araujo LM, Oliveira DA, Nunes DS, Liver and biliary ultrasonography in diabetic and non-diabetic obese women, Diabetes & Metabolism, 24, 1998, 458-462.
- Neuschwander-Tetri BA, Caldwell SH, Non-alcoholic-steatohepatitis: summary of an AASLD Single Topic Conference, Hepatology, 37, 2003, 1202-1219.
- Browning JD, Szczepaniak LS, Dobbins R, Nuremberg P, Horton JD, Cohen JC, Grundy SM, Hobbs HH, Prevalence of hepatic steatosis in an urban population in the United States: impact of ethnicity, Hepatology, 40, 2004, 1387-1395.
- Bedogni G, Miglioli L, Masutti F, Tiribelli C, Marchesini G, Bellentani S, Prevalence of and risk factors for nonalcoholic fatty liver disease: the Dionysos Nutrition and Liver Study, Hepatology, 42, 2005, 44-52.
- Bellentani S, Saccoccio G, Masutti F, Crocè LS, Brandi G, Sasso F, Cristanini G, Tiribelli C: Prevalence of and risk factors for hepatic



- steatosis in Northern Italy, *Annals of Internal Medicine*, 132, 2000, 112–117.
35. Browning JD, Statins and hepatic steatosis: perspectives from the Dallas Heart Study, *Hepatology*, 44, 2006, 466–471.
 36. Williams CD, Stengel J, Asike MI, Torres DM, Shaw J, Contreras M, Landt CL, Harrison SA, Prevalence of nonalcoholic fatty liver disease and non-alcoholic-steatohepatitis among a largely middle-aged population utilizing ultrasound and liver biopsy: a prospective study, *Gastroenterology*, 140(1), 2011, 124-131.
 37. Raji A, Seely EW, Arky RA, Simonson DC, Body fat distribution and insulin resistance in healthy Asian Indians and Caucasians, *Journal of Clinical Endocrinology & Metabolism*, 86, 2001, 5366–5371.
 38. Amarapurkar DN, Hashimoto E, Lesmana LA, Sollano JD, Chen PJ, Goh KL, Asia-Pacific Working Party on NAFLD, How common is non-alcoholic fatty liver disease in the Asia-Pacific region and are there local differences? *Journal of Gastroenterology Hepatology*, 22, 2007, 788–793.
 39. Jimba S, Nakagami T, Takahashi M, Wakamatsu T, Hirota Y, Iwamoto Y, Wasada T, Prevalence of non-alcoholic fatty liver disease and its association with impaired glucose metabolism in Japanese adults, *Diabetic Medicine*, 22, 2005, 1141–1145.
 40. Yoon KH, Lee JH, Kim JW, Cho JH, Choi YH, Ko SH, Zimmet P, Son YH, Epidemic obesity and type 2 diabetes in Asia, *The Lancet*, 368, 2006, 1681–1688.
 41. Ruhl CE, Everhart JE, Fatty liver indices in the multiethnic United States National Health and Nutrition Examination Survey, *Alimentary pharmacology & therapeutics*, 41(1), 2015, 65-76.
 42. Omagari K, Kadokawa Y, Masuda JI, Egawa I, Sawa T, Hazama H, Ohba K, Isomoto H, Mizuta Y, Hayashida K, Murase K, Kadota T, Murata I, Kohno S, Fatty liver in non-alcoholic non-overweight Japanese adults: incidence and clinical characteristics, *Journal of Gastroenterology Hepatology*, 17, 2002, 1098–1105.
 43. Huang HL, Lin WY, Lee LT, Wang HH, Lee WJ, Huang KC, Metabolic syndrome is related to non-alcoholic-steatohepatitis in severely obese subjects, *obese surgery*, 17, 2007, 1457–1463.
 44. Chen CH, Huang MH, Yang JC, Nien CK, Yang CC, Yeh YH, Yueh SK, Prevalence and risk factors of non-alcoholic fatty liver disease in an adult population of Taiwan: metabolic significance of non-alcoholic fatty liver disease in non-obese adult, *Journal of Clinical Gastroenterology*, 40, 2006, 745–752.
 45. Barshop NJ, Sirlin CB, Schwimmer JB, Lavine JE, Review article: epidemiology, pathogenesis and potential treatments of pediatric non-alcoholic fatty liver disease, *Alimentary Pharmacology & Therapeutics*, 28(1), 2008, 13–24.
 46. Rashid M, Roberts EA: Nonalcoholic steatohepatitis in children. *Journal of Pediatric Gastroenterology and Nutrition*, 30, 2000, 48–53.
 47. Tazawa Y, Noguchi H, Nishinomiya F, Takada G: Effect of weight changes on serum transaminase activities in obese children, *Acta Paediatrica Japonica*, 39, 1997, 210–214.
 48. Nonomura A, Mizukami Y, Unoura M, Kobayashi K, Takeda Y, Takeda R, Clinicopathologic study of alcohol- like liver disease in non-alcoholics; non-alcoholic steatohepatitis and fibrosis, *Journal of Gastroenterology*, 27, 1992, 521–528.
 49. Rashid M, Roberts EA, Nonalcoholic steatohepatitis in children, *Journal of Pediatric Gastroenterology and Nutrition*, 30, 2000, 48–53.
 50. Farhadi A, Gundlapalli S, Shaikh M, Frantzides C, Harrell L, Kwasny MM, Keshavarzian A, Susceptibility to gut leakiness: a possible mechanism for endotoxaemia in non-alcoholic steatohepatitis, *Liver international*, 28, 2008, 1026-33.
 51. Ritze Y, bardos G, Claus A, Ehrmann V, Bergheim I, Schwiertz A, Bischoff CS, *Lactobacillus rhamnosus GG* protects against non-alcoholic fatty liver disease in mice, *Plos one*, 9(1), 2014, e80169.
 52. Dowman JK, Tomlinson JW, Newsome PN, Pathogenesis of non-alcoholic fatty liver disease, *QJM: An International Journal of Medicine*, 103, 2010, 71–83.
 53. Cesaro C, Tiso A, Del Prete A, Cariello R, Tuccillo C, Cotticelli G, Del Vecchio Blanco C, Loguercio C. Gut microbiota and probiotics in chronic liver diseases, *Digestive and Liver Disease*, 43(6), 2011, 431-438.
 54. Anstee QM, Goldin RD, Mouse models in non-alcoholic fatty liver disease and steatohepatitis research, *International Journal of Experimental Pathology*, 87(1), 2006, 1-16.
 55. Neish AS, *Microbes in Gastrointestinal Health and Disease*, *Gastroenterology*, 136(1), 2009, 65-80.
 56. Zhang G, Ghosh S, Toll-like receptor–mediated NF-κB activation: a phylogenetically conserved paradigm in innate immunity, *Journal of Clinical Investigation*, 107(1), 2001, 13-19.
 57. Nagata K, Suzuki H, Sakaguchi S, Common pathogenic mechanism in development progression of liver injury caused by non-alcoholic or alcoholic steatohepatitis, the journal of toxicological sciences, 32(5), 2007, 453-468.
 58. Takaki A, Kawai D, Yamamoto K, Molecular Mechanisms and New Treatment Strategies for Non-Alcoholic Steatohepatitis (NASH), *International Journal of Molecular Sciences*, 15(5), 2014, 7352-7379.
 59. Farrell GC, Larter CZ, Nonalcoholic fatty liver disease: from steatosis to cirrhosis, *Hepatology*, 43, 2006, S99-S112.
 60. Paradis V, Zalinski S, Chelbi E, Guedj N, Degos F, Vilgrain V, Hepatocellular carcinomas in patients with metabolic syndrome often develop without significant liver fibrosis: a pathological analysis, *Hepatology*, 49(3), 2009, 851-859.
 61. Agarwal SR, Malhotra V, Sakhuja P, Sarin SK., Clinical biochemical and histological profile of non-alcoholic-steatohepatitis, *Indian Journal of Gastroenterology*, 20, 2001, 183–186.
 62. Matteoni CA, Younossi ZM, Gramlich T, Boparai N, Liu YC, McCullough AJ, Non-alcoholic fatty liver disease: a spectrum of clinical and pathological severity. *Gastroenterology*, 116(6), 1999, 1413-1419.
 63. Langlands SJ, Hopkins MJ, Coleman N, Cummings JH, Prebiotic carbohydrates modify the mucosa associated micro-flora of the human large bowel, *Gut*, 53, 2004, 1610-1616.
 64. Synytsya A, Mickova K, Synytsya A, Jablonsky I, Spevacek J, Erban V, Kovarikova E, opikova JC, Glucans from fruit bodies of cultivated mushrooms *Pleurotus ostreatus* and *Pleurotus eryngii*: Structure and potential prebiotic activity, *Carbohydrate Polymers*, 76, 2009, 548–556.
 65. Han XQ, Wu XM, Chai XY, Chen D, Dai H, Dong HL, Ma ZZ, Gao XM, Tu PF, Isolation, characterization and immunological activity of a polysaccharide from the fruit bodies of an edible mushroom *Sarcodon aspratus* (Berk.) S. Ito, *Food Research International*, 44, 2011, 489-93.
 66. Wasser SP, Medicinal mushrooms as a source of antitumor and immune-modulating polysaccharides, *Application of Microbiological Biotechnology*, 60, 2002, 258–274.
 67. Convertib A, de Souza RPO, Peregob P, de Oliveiraa MN, Effect of inulin as prebiotic and synbiotic interactions between probiotics to improve fermented milk firmness, *Journal of Food Engineering*, 107(1), 2011, 36–40.
 68. Evaluation of the various uses of microorganisms with emphasis on probiotics, *Journal of Microbial & Biochemical Technology*, R1, 004, 2011, 1-7.



69. Suganya K, Murugan T, Murugan M, Isolation and characterization of probiotic lactic acid bacteria from milk and curd samples, *International Journal of Pharma and Bio Sciences*, 4(1b), 2013, 317–324.
70. Fuentes MC, Lajo T, Carrion JM, Cune J, Cholesterol-lowering efficacy of *Lactobacillus plantarum* CECT 7527, 7528 and 7529 in hyper-cholesterolaemic adults, *British Journal of Nutrition*, 2012, 1-7.
71. Gilliland SE, Nelson CR & Maxwell C, Assimilation of cholesterol by *Lactobacillus acidophilus*, *Applied and Environmental Microbiology*, 49, 1985, 377–381.
72. Akin MB, Akin MS, Kirmaci Z, Effects of inulin and sugar levels on the viability of yoghurt and probiotic bacteria and the physical and sensory characteristics in probiotic ice-cream, *Food Chemistry*, 104(1), 2007, 93–99.
73. Bielecka M, Biedrzycka E, Majkowska A, Selection of bifidobacterial strains capable for colonisation of gastrointestinal tract, *Medical Science Monitor, International Medical Journal for Experimental and Clinical Research* 6, Suppl. 3, 2000, 123.
74. Kailasapathy K, Chin J, Survival and therapeutic potential of probiotic organisms with reference to *Lactobacillus acidophilus* and *Bifidobacterium* spp., *Immunology & Cell Biology - Nature*, 78, 2000, 80-88.
75. De Marzio DLH, Fenkel JM, Concepts and Treatment Approaches in Nonalcoholic Fatty Liver Disease, *Advances in Hepatology*, 2014, 2014, 7.
76. Crittenden R, Weerakkody R, Sanguansri L, Augustin MA, Synbiotic Microcapsules That Enhance Microbial Viability during Non-refrigerated Storage and Gastrointestinal Transit, *Applied and Environmental Microbiology*, 72(3), 2006, 2280–2282.
77. Wall A, Hernandez RIC, Parrilla EA, Mendoza JL, Rubio ARI, la Rosa LAD, Structural Stability and Viability of Microencapsulated Probiotic Bacteria: A Review, *Comprehensive Reviews in Food Science and Food Safety*, 12(6), 2013, 614-628.
78. Kailasapathy K, Microencapsulation of probiotic bacteria: technology and potential applications, *Current Issues in Intestinal Microbiology*, 3, 2002, 39–48.
79. Chen MJ, Chen KN, Applications of probiotic encapsulation in dairy products in Encapsulation and Controlled Release Technologies in Food Systems, 2007, 83–107.
80. Sheu TY, Marshall RT, Micro-encapsulation of *Lactobacilli* in calcium alginate gels, *Journal of Food Science*, 54, 1993, 557–561.
81. McClements DJ, Decker EA, Park Y, Controlling lipid bioavailability through physicochemical and structural approaches, *Critical Reviews in Food Science and Nutrition*, 49, 2009a, 48–67.
82. Redgwell RJ, Fischer M, Dietary fiber as a versatile food component: an industrial perspective, *Molecular Nutrition and Food Research*, 49, 2005, 521–535.
83. Adamson AW, Physical chemistry of surfaces, John Wiley & Sons, New York, 1982, 649.
84. Krasaekoopt W, Bhandari B, Deeth H, Evaluation of encapsulation techniques of probiotics for yoghurt, *International Dairy Journal*, 13(1), 2003, 3–13.
85. Solanki HK, Pawar DD, Shah DA, Prajapati VD, Jani GK, Mulla AM, Thakar PM, Development of Microencapsulation Delivery System for Long-Term Preservation of Probiotics as Biotherapeutics Agent, *BioMed Research International*, 21, 2013, 2013.
86. King AH, Encapsulation of Food Ingredients: A review of available technology focusing on hydrocolloids in Encapsulation and Controlled Release of Food Ingredients, *American Chemical Society*, 1995, 26-39.
87. Klein J, Stock J, Vorlop KD, Pore size and properties of spherical Ca-alginate biocatalysts, *European Journal Applied Microbiology Biotechnology*, 18(1), 1983, 86–91.
88. Kailasapathy K, Encapsulation technologies for functional foods and nutraceutical product development, *CAB Reviews*, 4(6), 2009, 1–19.
89. Melvik JE, Braek SG, Gaserod O, Klokke TI, Skaugrud F, Electrostatic bead generator for immobilisation of cells and macromolecules, *The 8th International Workshop on Bioencapsulation-Recent Progress in Research and Technology*, 1999, 13-15.
90. Tanaka H, Masatose M, Veleky IA, Diffusion characteristics of substrates in Ca-alginate beads, *Biotechnology and Bioengineering*, 26(1), 1984, 53–58.
91. Hainzen C, Microencapsulation by prilling and co-extraction, Workshop no.53, nutraceuticals and probiotics, technology training centre, basil, Germany, 2002, 26-28.
92. Smidsrod O, Haug A, Lian B Properties of poly (1,4- heuronates) in the gel state, Evaluation of a method for the determination of stiffness, *Acta Chemica Scandinavica*, 26(1), 1972, 71–78.
93. Krasaekoopt W, Bhandari B, Deeth H, The influence of coating materials on some properties of alginate beads and survivability of microencapsulated probiotic bacteria, *International Dairy Journal*, 14, 2004, 737–743.
94. Schlameus W, Centrifugal Extrusion Encapsulation, Encapsulation and controlled release of food ingredients, 590, *American Chemical Society*, San Antonio, 1995, 96–103.
95. Lacroix C, Paquin C, Arnaud JP, Batch fermentation with entrapped growing cells of *Lactobacillus casei*. Optimization of the rheological properties of the entrapment, *Applied Microbiology and biotechnology*, 32(4), 1990, 403-408.
96. Kebary KMK, Hussein SA, Badawi RM, Improving viability of *Bifidobacteria* and their effect on frozen ice milk, *Egyptian Journal of Dairy Science*, 26(2), 1998, 319–337.
97. Sheu TY, Marshall RT, Heymann H, Improving survival of culture bacteria in frozen desserts by micro-entrapment, *Journal of Dairy Science*, 76(7), 1993, 1902–1907.
98. Rao AV, Shivanarain N, Maharaj I, Survival of microencapsulated *Bifidobacterium pseudolongum* in simulated gastric and intestinal juices, *Canadian Institute of Food Science and Technology Journal*, 22(4), 1989, 345–349.
99. Porubcan RS & Sellers RL, Stabilized dry cultures of lactic acid producing bacteria, *US Patent* 3, 897, 1975, 307.
100. Dziezak JD, microencapsulation and encapsulated ingredients, *Food Technology*, 42, 1988, 36-151.
101. Jackson LS and Lee K, Microencapsulation the food industry, *Food science technology*, 24, 1991, 289-297.
102. Mauriello G, Aponte M, Andolfi R, Moschetti G, Villani F, Spray-drying of bacteriocin producing lactic acid bacteria, *Journal of Food Protection*, 62, 1999, 773-777.
103. Riordan OK, Andrews D, Buckle K, Conway P, Evaluation of microencapsulation of a *Bifidobacterium* strain with starch as an approach to prolonging viability during storage. *Journal of Applied Microbiology*, 91, 2001, 1059-1066.
104. Champagne CP, fustier P, microencapsulation for improved delivery of bioactive compounds into foods, *current opinion in biotechnology*, 18(2), 2007, 184-190.
105. Bakan JA, The theory and practice of industrial pharmacy, *Microencapsulation In: Lachman L, Lieberman HA, Kanig JL ed., 3rd ed. Ch. 13, Part III, Varghese Publishing House, Bombay, 1991, 419.*
106. Semyonov D, Ramon O, Kovacs A, Shani LF, Shimoni E, Air-Suspension Fluidized-Bed Microencapsulation of Probiotics, *Drying Technology*, 30(16), 2012, 1918-1930.



107. Dong Z, Wang Q, Du Y, Alginate/gelatin blend films and their properties for drug controlled release, *Journal of Membrane Science*, 280(1-2), 2006, 37–44.
108. Draget KI, Steinsvag K, Onsoyen E, Smidsrod O, Na⁺ and K⁺alginate; effect on Ca²⁺-gelation, *Carbohydrate Polymers*, 35(1-2), 1998, 1–6.
109. Hood SK, Zottola EA, Effect of low pH on the ability of Lactobacillus acidophilus to survive and adhere to human intestinal cells, *Journal of Food Science*, 53(5), 1988, 1514–1516.
110. Lee K, Heo T, Survival of Bifidobacterium longum immobilized in calcium alginate beads in simulated gastric juices and bile salt solution, *Applied and Environmental Microbiology*, 66(2), 2000, 869–873.
111. Harnsilawat T, Pongsawatmanit R, McClements DJ, Characterization of -lactoglobulin-sodium alginate interactions in aqueous solutions: a calorimetry, light scattering, electrophoretic mobility and solubility study, *Food Hydrocolloids*, 20(5), 2006, 577–585.
112. Hansen LT, Allan-Wojtas PM, Jin YL, Paulson AT, Survival of Calcium alginate microencapsulated Bifidobacterium spp. in milk and simulated gastrointestinal conditions, *Food Microbiology*, 19(1), 2002, 35–45.
113. Klein J, Stock J, Vorlop KD, Pore size and properties of spherical Calcium alginate biocatalysts, *European Journal of Applied Microbiology and Biotechnology*, 18(2), 1983, 86–91.
114. Lian W, Hsiao H, Chou C, Survival of bifidobacteria after spray-drying, *International Journal of Food Microbiology*, 74(1-2), 2002, 79–86.
115. Fanta GF, Knutson CA, Eskins KS, Felker FC, Starch microcapsules for delivery of active agents, US patent, 6, 2001, 238-677.
116. Haralampu SG, Resistant starch—a review of the physical properties and biological impact of RS3, *Carbohydrate Polymers*, 41(3), 2000, 285–292.
117. Malm CJ, Emerson J, Hiatt GD, Cellulose acetate phthalate as an enteric coating material, *Journal of the American Pharmaceutical Association*, 40(10), 1951, 520–525.
118. Kailasapathy K, Masondole L, Survival of free and microencapsulated Lactobacillus acidophilus and Bifidobacterium lactis and their effect on texture of feta cheese, *Australian Journal of Dairy Technology*, 60(3), 2005, 252–258.
119. Gaaloul S, Turgeon SL, Corredig M, Influence of shearing on the physical characteristics and rheological behaviour of an aqueous whey protein isolate-appa-carrageenan mixture, *Food Hydrocolloids*, 23(5), 2009, 1243–1252.
120. Yuguchi Y, Thuy TTT, Urakawa H, Kajiwarra K, Structural characteristics of carrageenan gels: temperature and concentration dependence, *Food Hydrocolloids*, 16(6), 2002, 515–522.
121. Mangione MR, Giacomazza D, Bulone D, Martorana V, san Biagio PL, Thermo-reversible gelation of -Carrageenan: relation between conformational transition and aggregation, *Biophysical Chemistry*, 104(1), 2003, 95–105.
122. Sarett HP, Safety of carrageenan used in foods, *The Lancet*, 1(8212), 1981, 151–152.
123. Doleyres Y, Fliiss I, Lacroix C, Continuous production of mixed lactic starters containing probiotic using immobilized cell technology, *Biotechnology Progress*, 20(1), 2004, 145–150.
124. Klien J, Vorlop DK, Immobilisation technique cells, *Comprehensive Biotechnology*, Ed. M. Moo-Yong, C. L. Cooney, and A. E. Humphery, Pergamon Press, Oxford, UK, 1985, 542–550.
125. Dinakar P, Mistry VV, Growth and viability of Bifidobacterium bifidum in cheddar cheese, *Journal of Dairy Science*, 77(10), 1994, 2854–2864.
126. Chen MJ, Chen KN, Applications of probiotic encapsulation in dairy products, *Encapsulation and controlled release technologies in food systems*, ed J. M. Lakkis, Blackwell publishing, Ames, Iowa, USA, 2007, 83–107.
127. Rokka S, Rantam R, Protecting probiotic bacteria by microencapsulation: challenges for industrial applications, *European Food Research and Technology*, 231(1), 2010, 1–12.
128. Livney YD, Milk proteins as vehicles for bio-actives, *Current Opinion in Colloid and Interface Science*, 15(1-2), 2010, 73–83.
129. Reid A, Champagne CP, Gardner N, Fustier P, Vuilleumard JC, Survival in food systems of Lactobacillus rhamnosus R011 microentrapped in whey protein gel particles, *Journal of Food Science*, 72(1), 2006, M31–M37.

Source of Support: Nil, Conflict of Interest: None.

