



EFFECTS OF MACRO-MINERAL ELEMENTS ON GROWTH AND L-GLUTAMIC ACID FERMENTATION BY A MUTANT *MICROCOCCUS GLUTAMICUS* AB₁₀₀

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ABSTRACT

An experimental study was carried out to examine the effects of different macro-mineral elements on growth and l-glutamic acid fermentation by a mutant *Micrococcus glutamicus* AB₁₀₀. It was required for improved production of l-glutamic acid by this mutant. K₂HPO₄, 0.15%; MgSO₄.7H₂O, 0.03% and CaCO₃, 0.04% were required for improved production of l-glutamic acid by this mutant. KH₂PO₄, NaH₂PO₄.H₂O, HCl, NaCl and CaCl₂ showed no effect on growth and l-glutamic acid accumulation, where as K₂B₄O₇.X H₂O showed negative impact on growth and the production. L-glutamic acid production was increased significantly (p<0.01) from 14.8 mg/ml to 18.8 mg/ml and dry cell weight was increased significantly (p<0.01) from 7.3 mg/ml to 9.8 mg/ml after addition of the necessary macro-mineral elements in the fermentation broth.

Keywords: Macro-mineral, growth, L-glutamic acid, mutant, *Micrococcus glutamicus*.

INTRODUCTION

L-glutamic acid, a non-essential amino acid has a wide spectrum of commercial use as flavor enhancer, food additive, infusion compounds etc¹. The industrial production of l-glutamic acid is mainly carried out by fermentation using bacterial species namely *Micrococcus glutamicus*, *Brevibacterium roseum*, *Brevibacterium flavum* etc.² For l-glutamic acid over production, *Micrococcus glutamicus* strains have been developed in our laboratory by induced mutation in our laboratory³. However, each bacterium has a defined range of growth conditions including requirements of macro-minerals elements⁴. Roy and Chatterjee (1989) reported the influence of culture conditions on l-glutamic acid production by *Arthrobacter globiformis*⁵. Lee et al (2006) claimed the requirements of different macro-mineral elements on growth and l-threonine production by *E.Coli* mutant⁶. Kase and Nakayama, Banik and Majumdar (1975), tani et al (1988) reported that microbial production of l-methionine required different macro elements⁷⁻¹¹.

Considering all these facts, the present study was intended to examine the effects of different macro-mineral elements on growth and l-glutamic acid fermentation by a biotin-auxotrophic mutant *Micrococcus glutamicus* AB₁₀₀.

MATERIALS AND METHODS

Microorganism: *Micrococcus glutamicus* AB₁₀₀, a biotin requiring auxotrophic mutant derived from a regulatory mutant derived from a regulatory mutant *Micrococcus glutamicus* AB₁ by induced mutation was used throughout this study³.

Minimal salt medium: Minimal salt medium contained glucose, 9.0%; (NH₄)₂HPO₄, 1.4%; MgSO₄.7H₂O, 0.025%; K₂HPO₄, 0.1%; biotin, 0.2 µg/ml. pH was adjusted to 6.5.

Fermentation conditions: Fermentation was carried out using shake-flask method on a rotary shaker rotating at 150 rpm, in 100 ml Erlenmeyer conical flask containing 20 ml minimal salt medium for 72h at 29°C. The medium was inoculated with 4.0% (v/v) of a 48h old seed culture (6.0 x 10⁷ cells) of *Micrococcus glutamicus* AB₁₀₀¹².

Addition of different macro mineral elements : Different macro-mineral elements (namely, K₂HPO₄, KH₂PO₄, NaH₂PO₄.H₂O, MgSO₄.7H₂O, KCl, NaCl, CaCl₂.2H₂O, CaCO₃ and K₂B₄O₇.XH₂O) at varying concentrations (0.02-0.2%) were added to the minimal salt medium one by one to assess their effects on growth and l-glutamic acid production by this mutant⁷.

Analysis of amino acid: Descending paper chromatography was used to detect l-glutamic acid in culture broth using a solvent system composed of n-butanol : acetic acid : water (2:1:1) which was run for 18h on a whatman no. 1 chromatography paper. The spots were visualized by spraying with a solution of 0.2% ninhydrin in acetone and quantitative estimation of l-glutamic acid was done using colorimetric estimation method^{13,14}.

Estimation of Dry Cell Weight: After proper centrifugation, 2 ml of 1.0 (M) HCl was poured into the precipitate of the bacterial cells and CaCO₃ to dissolve CaCO₃. The remaining bacterial cells were washed with water and derived at 100°C until cells weight remain constant¹⁵.

Statistical analysis: All data were expressed as mean ± SEM, where n = 6. The data were analyzed by one way ANOVA followed by Dunett's post-hoc multiple



comparison test using “prism 4.0” software (Graph pad Inc., USA). A “p” value less than 0.05 was considered significant and less than 0.01 as a highly significant.

RESULTS AND DISCUSSION

The effect of different macro-mineral elements on growth and l-glutamic acid production by the mutant *Micrococcus glutamicus* AB₁₀₀ were depicted in Fig 1-9. Production was

maximum with KH₂PO₄, 0.15%; MgSO₄.7H₂O, 0.03% and CaCO₃, 0.04% along with maximum dry cell weight KH₂PO₄, NaH₂PO₄.H₂O, KCl, NaCl, and CaCl₂.2H₂O did not have any significant impact on both cellular growth and l-glutamic acid production. But K₂B₄O₇.XH₂O was proved to be detrimental for both cellular growth and l-glutamic acid production.

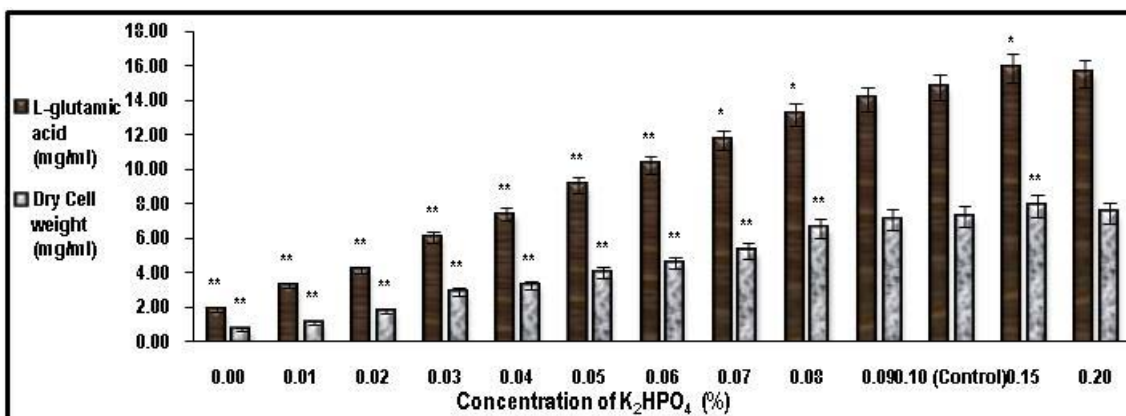


Figure 1: Effect of K₂HPO₄ on growth and production of L-glutamic acid by *Micrococcus glutamicus* AB₁₀₀. (values were expressed as mean ± SEM, where n=6; *p<0.05; **p<0.01 when compared to control)

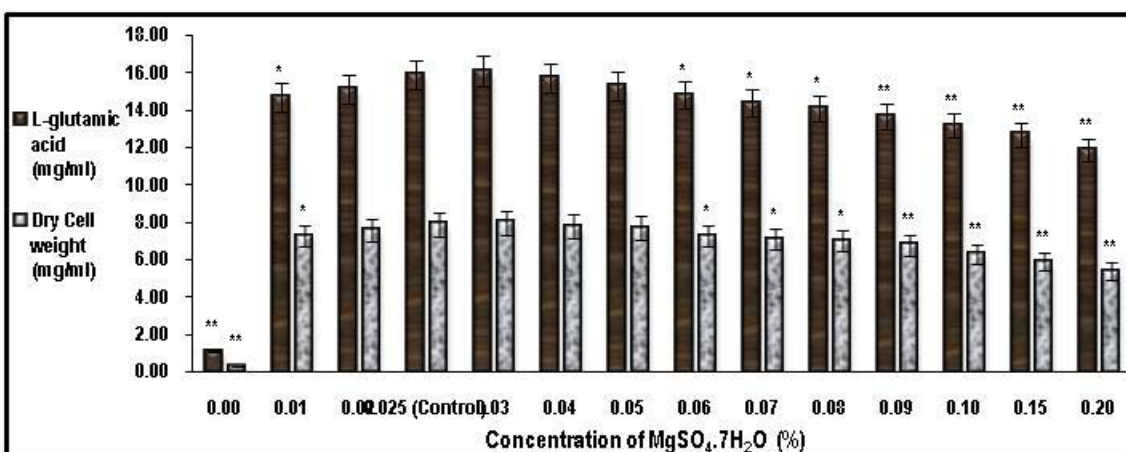


Figure 2: Effect of MgSO₄.7H₂O on growth and production of L-glutamic acid by *Micrococcus glutamicus* AB₁₀₀. (values were expressed as mean ± SEM, where n=6; *p<0.05; **p<0.01 when compared to control)

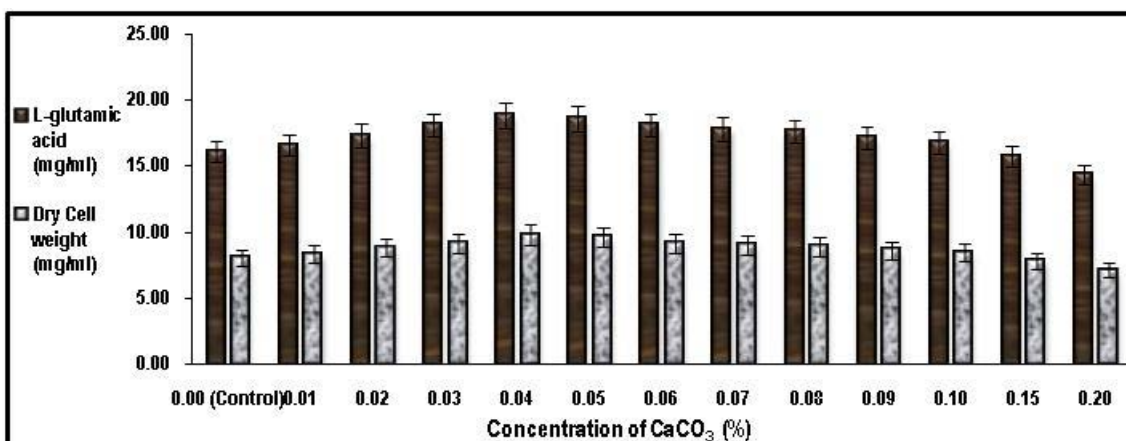


Figure 3: Effect of CaCO₃ on growth and production of L-glutamic acid by *Micrococcus glutamicus* AB₁₀₀. (values were expressed as mean ± SEM, where n=6; *p<0.05; **p<0.01 when compared to control)



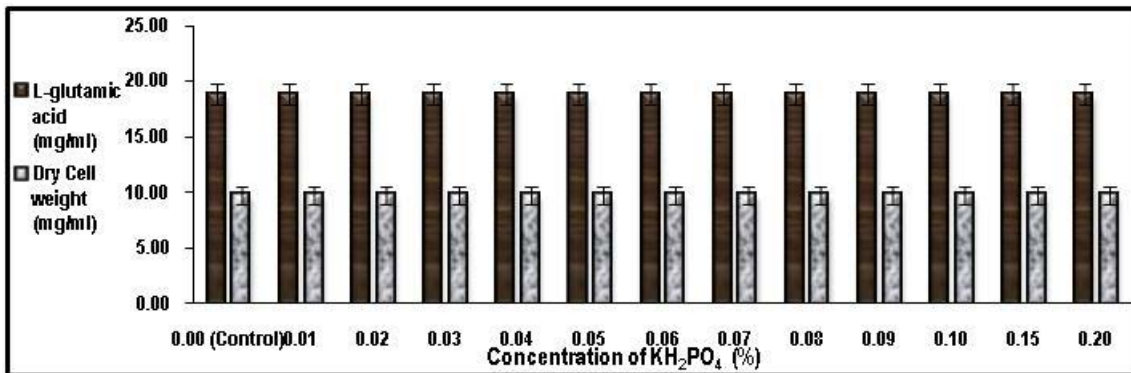


Figure 4: Effect of KH₂PO₄ on growth and production of L-glutamic acid by *Micrococcus glutamicus* AB₁₀₀. (values were expressed as mean ± SEM, where n=6; *p<0.05; **p<0.01 when compared to control)

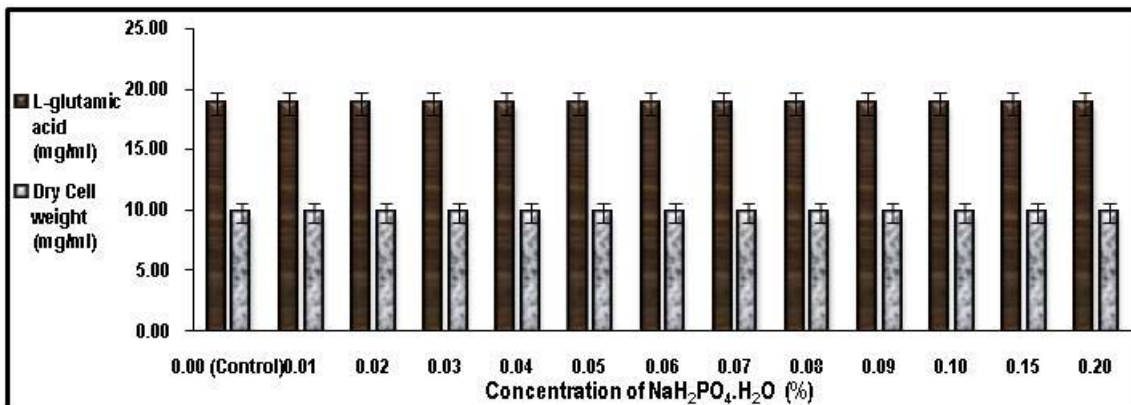


Figure 5: Effect of NaH₂PO₄.H₂O on growth and production of L-glutamic acid by *Micrococcus glutamicus* AB₁₀₀. (values were expressed as mean ± SEM, where n=6; *p<0.05; **p<0.01 when compared to control)

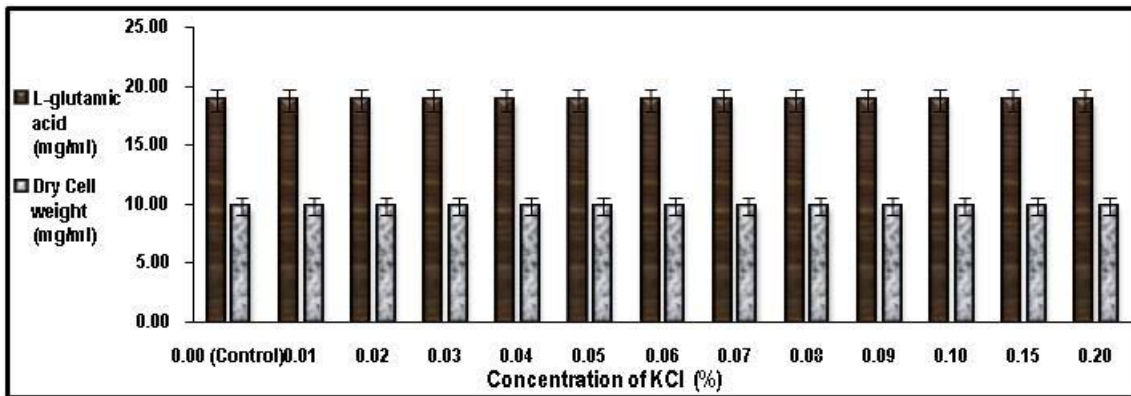


Figure 6: Effect of KCl on growth and production of L-glutamic acid by *Micrococcus glutamicus* AB₁₀₀. (values were expressed as mean ± SEM, where n=6; *p<0.05; **p<0.01 when compared to control)

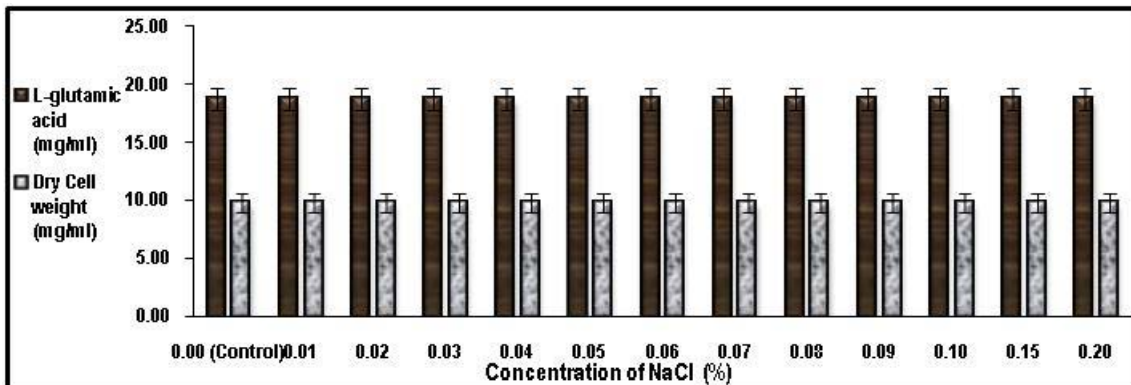


Figure 7: Effect of NaCl on growth and production of L-glutamic acid by *Micrococcus glutamicus* AB₁₀₀. (values were expressed as mean ± SEM, where n=6; *p<0.05; **p<0.01 when compared to control)

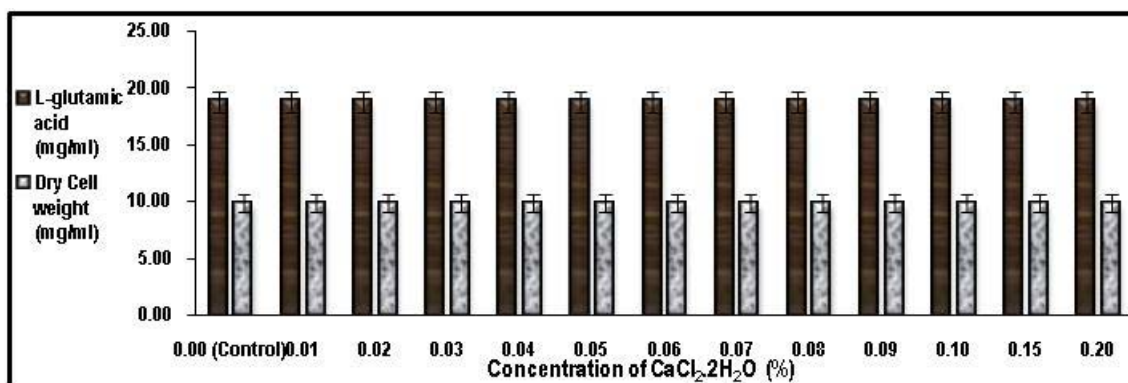


Figure 8: Effect of CaCl₂.2H₂O on growth and production of L-glutamic acid by *Micrococcus glutamicus* AB₁₀₀. (values were expressed as mean ± SEM, where n=6; *p<0.05; **p<0.01 when compared to control)

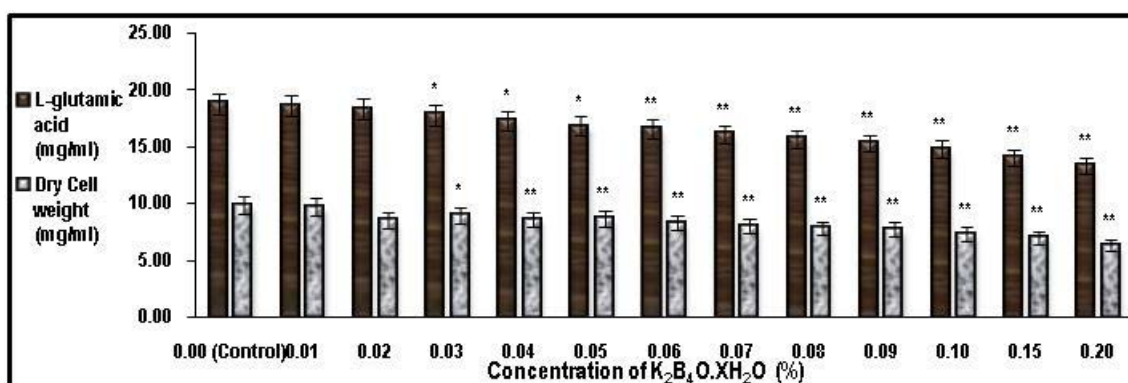


Figure 9: Effect of K₂B₄O₇.XH₂O on growth and production of L-glutamic acid by *Micrococcus glutamicus* AB₁₀₀. (values were expressed as mean ± SEM, where n=6; *p<0.05; **p<0.01 when compared to control)

Carito et al (1966) reported that *Fusarium moniliforme* required K₂HPO₄, 0.6%; KH₂PO₄, 0.4%; MgSO₄.7H₂O, 0.4%; NaCl, 0.2% and CaCl₂.2H₂O, 0.2% for l-alanine production¹⁶. Birnbacum et al (1969) used MgSO₄.7H₂O, 0.05% as macro-mineral element for l-glutamic acid production by *Corynebacterium glutamicum*¹⁶. Kase and Nakayama (1974) have reported K₂HPO₄, 0.05%; KH₂PO₄, 0.1%; MgSO₄.7H₂O, 0.01% were needed for l-methionine production¹⁸. Banik and Majumdar (1975) reported that production of l-methionine required K₂HPO₄, 0.03%; MgSO₄.7H₂O, 0.1% as macro-mineral element⁷. Lee et al (2006) used KH₂PO₄, 0.85%; MgSO₄.7H₂O, 0.1%; CaCl₂.2H₂O, 1.32%; K₂B₄O₇.XH₂O, 0.006% as macro-mineral element for l-threonine production *E. Coli* mutant⁶. Yugandhar et al (2007) reported that *Brevibacterium roseum* required K₂HPO₄, 0.12%; CaCO₃, 0.16%; MgSO₄.7H₂O, 0.01% and NaCl, 0.01% for l-glutamic acid production¹⁹. But there is no review available on the effect of KCl on growth and l-glutamic acid production by micro-organisms. In our present study we have observed that KCl showed no significant effect on growth and l-glutamic acid production by this mutant.

Thus, in this present study it was concluded that production of l-glutamic acid was increased significantly (p<0.01) from 18.8 mg/ml to 18.8 mg/ml after addition of K₂HPO₄, 0.15%; MgSO₄.7H₂O, 0.03% and CaCO₃, 0.04% in the fermentation broth.

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